

Appendix

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Appendix S1. The aim of each visit during the update of the red list of wild bees from Switzerland was to maximise the list of species on the corresponding day for the entire plot. Therefore, the collaborators decided themselves, taking into account sunlight, phenology, floral offer, small structures, etc. during each field visit, which habitats, structures or floral populations should be surveyed and for how long. For instance, if a large colony of *Lasioglossum malachurum* is found during a visit or if many patrolling males of *Andrena flavipes* are observed on a flower-rich slope, at least 5 individuals should be collected in the corresponding sub-square or noted in the data file. If on the same day many males of *Andrena flavipes* are again observed in another sub-square, at least 5 individuals should again be collected or recorded in this sub-square. Food plants of particular importance to wild bees were listed and were given special attention, as all mass-flowering plants (e.g., umbellifers). Besides flowers, small structures (earthen edges, steepy walls, patchy soil, sunny dead wood,...) were also checked during fieldwork.

Appendix S2. The 547 bee species found in the community plots used in this study. Species are sorted according to family. For each species, the number of occurrences in the sampled plots is provided. Not that the occurrences are only a presence indicator, and thus do not relate to the species local abundance.

Andrenidae		51 <i>Andrena intermedia</i>	180	102 <i>Andrena trimmerana</i>	676
1 <i>Andrena aeneiventris</i>	8	52 <i>Andrena labialis</i>	44	103 <i>Andrena vaga</i>	180
2 <i>Andrena afrensis</i>	16	53 <i>Andrena labiata</i>	224	104 <i>Andrena varians</i>	44
3 <i>Andrena agilissima</i>	25	54 <i>Andrena lagopus</i>	76	105 <i>Andrena ventralis</i>	224
4 <i>Andrena alftanella</i>	62	55 <i>Andrena lapponica</i>	68	106 <i>Andrena viridescens</i>	76
5 <i>Andrena allosa</i>	11	56 <i>Andrena lathyri</i>	136	107 <i>Andrena vulpecula</i>	68
6 <i>Andrena alutacea</i>	1	57 <i>Andrena limata</i>	3	108 <i>Andrena wilkella</i>	136
7 <i>Andrena amieti</i>	73	58 <i>Andrena marginata</i>	23	109 <i>Melitturga clavicornis</i>	3
8 <i>Andrena ampla</i>	3	59 <i>Andrena minutula</i>	912	110 <i>Panurginus herzi</i>	23
9 <i>Andrena anthrisci</i>	16	60 <i>Andrena minutuloides</i>	377	111 <i>Panurginus montanus</i>	912
10 <i>Andrena apicata</i>	25	61 <i>Andrena mitis</i>	30	112 <i>Panurgus banksianus</i>	377
11 <i>Andrena argentata</i>	4	62 <i>Andrena montana</i>	11	113 <i>Panurgus calcaratus</i>	30
12 <i>Andrena barbareae</i>	58	63 <i>Andrena nana</i>	34	114 <i>Panurgus dentipes</i>	11
13 <i>Andrena barbilabris</i>	76	64 <i>Andrena nanula</i>	2	Apidae	34
14 <i>Andrena bicolor</i>	692	65 <i>Andrena nigroaenea</i>	160	115 <i>Amegilla albigena</i>	2
15 <i>Andrena bimaculata</i>	23	66 <i>Andrena nigroolivacea</i>	41	116 <i>Amegilla garrula</i>	160
16 <i>Andrena bucephala</i>	111	67 <i>Andrena nitida</i>	630	117 <i>Ammobates punctatus</i>	41
17 <i>Andrena carantonica</i>	242	68 <i>Andrena nitidiuscula</i>	33	118 <i>Anthophora aestivalis</i>	630
18 <i>Andrena chrysopus</i>	3	69 <i>Andrena niveata</i>	2	119 <i>Anthophora balnearum</i>	33
19 <i>Andrena chrysosceles</i>	415	70 <i>Andrena nuptialis</i>	4	120 <i>Anthophora bimaculata</i>	2
20 <i>Andrena cineraria</i>	328	71 <i>Andrena ovatula</i>	865	121 <i>Anthophora crassipes</i>	4
21 <i>Andrena clarkella</i>	24	72 <i>Andrena pandellei</i>	89	122 <i>Anthophora crinipes</i>	865
22 <i>Andrena coitana</i>	23	73 <i>Andrena parviceps</i>	21	123 <i>Anthophora dispar</i>	89
23 <i>Andrena combinata</i>	93	74 <i>Andrena pilipes</i>	15	124 <i>Anthophora furcata</i>	21
24 <i>Andrena confinis</i>	1	75 <i>Andrena polita</i>	24	125 <i>Anthophora mucida</i>	15
25 <i>Andrena congruens</i>	42	76 <i>Andrena potentillae</i>	16	126 <i>Anthophora plagiata</i>	24
26 <i>Andrena curvungula</i>	168	77 <i>Andrena praecox</i>	87	127 <i>Anthophora plumipes</i>	16
27 <i>Andrena denticulata</i>	12	78 <i>Andrena probata</i>	24	128 <i>Anthophora pubescens</i>	87
28 <i>Andrena distinguenda</i>	2	79 <i>Andrena propinqua</i>	58	129 <i>Anthophora quadrimaculata</i>	24
29 <i>Andrena dorsata</i>	325	80 <i>Andrena proxima</i>	169	130 <i>Anthophora retusa</i>	58
30 <i>Andrena falsifica</i>	239	81 <i>Andrena ranunculorum</i>	4	131 <i>Apis mellifera</i>	169
31 <i>Andrena ferox</i>	4	82 <i>Andrena rogenhoferi</i>	26	132 <i>Biastes emarginatus</i>	4
32 <i>Andrena flavipes</i>	773	83 <i>Andrena rosae</i>	29	133 <i>Bombus alpinus</i>	26
33 <i>Andrena florea</i>	93	84 <i>Andrena ruficrus</i>	44	134 <i>Bombus argillaceus</i>	29
34 <i>Andrena floricola</i>	6	85 <i>Andrena rufizona</i>	30	135 <i>Bombus barbutellus</i>	44
35 <i>Andrena florivaga</i>	6	86 <i>Andrena rufula</i>	5	136 <i>Bombus bohemicus</i>	30
36 <i>Andrena freygessneri</i>	6	87 <i>Andrena rugulosa</i>	3	137 <i>Bombus campestris</i>	5
37 <i>Andrena fucata</i>	48	88 <i>Andrena schencki</i>	1	138 <i>Bombus cryptarum</i>	1
38 <i>Andrena fulva</i>	130	89 <i>Andrena semilaevis</i>	28	139 <i>Bombus distinguendus</i>	28
39 <i>Andrena fulvago</i>	337	90 <i>Andrena similis</i>	41	140 <i>Bombus flavidus</i>	41
40 <i>Andrena fulvata</i>	327	91 <i>Andrena similima</i>	56	141 <i>Bombus gerstaeckeri</i>	56
41 <i>Andrena fubicornis</i>	3	92 <i>Andrena simontornyella</i>	7	142 <i>Bombus hortorum</i>	7
42 <i>Andrena fubida</i>	6	93 <i>Andrena strohmella</i>	14	143 <i>Bombus humilis</i>	14
43 <i>Andrena fuscipes</i>	4	94 <i>Andrena subopaca</i>	326	144 <i>Bombus hypnorum</i>	326
44 <i>Andrena gelriae</i>	10	95 <i>Andrena suerinensis</i>	690	145 <i>Bombus inexpectatus</i>	690
45 <i>Andrena gravida</i>	501	96 <i>Andrena symphyti</i>	1	146 <i>Bombus jonellus</i>	1
46 <i>Andrena haemorrhhoa</i>	801	97 <i>Andrena symadelpa</i>	4	147 <i>Bombus lapidarius</i>	4
47 <i>Andrena hattorfiana</i>	279	98 <i>Andrena taraxaci</i>	3	148 <i>Bombus lucorum</i>	3
48 <i>Andrena helvola</i>	159	99 <i>Andrena tarsata</i>	12	149 <i>Bombus mendax</i>	12
49 <i>Andrena hesperia</i>	1	100 <i>Andrena thoracica</i>	11	150 <i>Bombus mesomelas</i>	11
50 <i>Andrena humilis</i>	677	101 <i>Andrena tibialis</i>	57	151 <i>Bombus monticola</i>	57

Appendix S2. Cont.

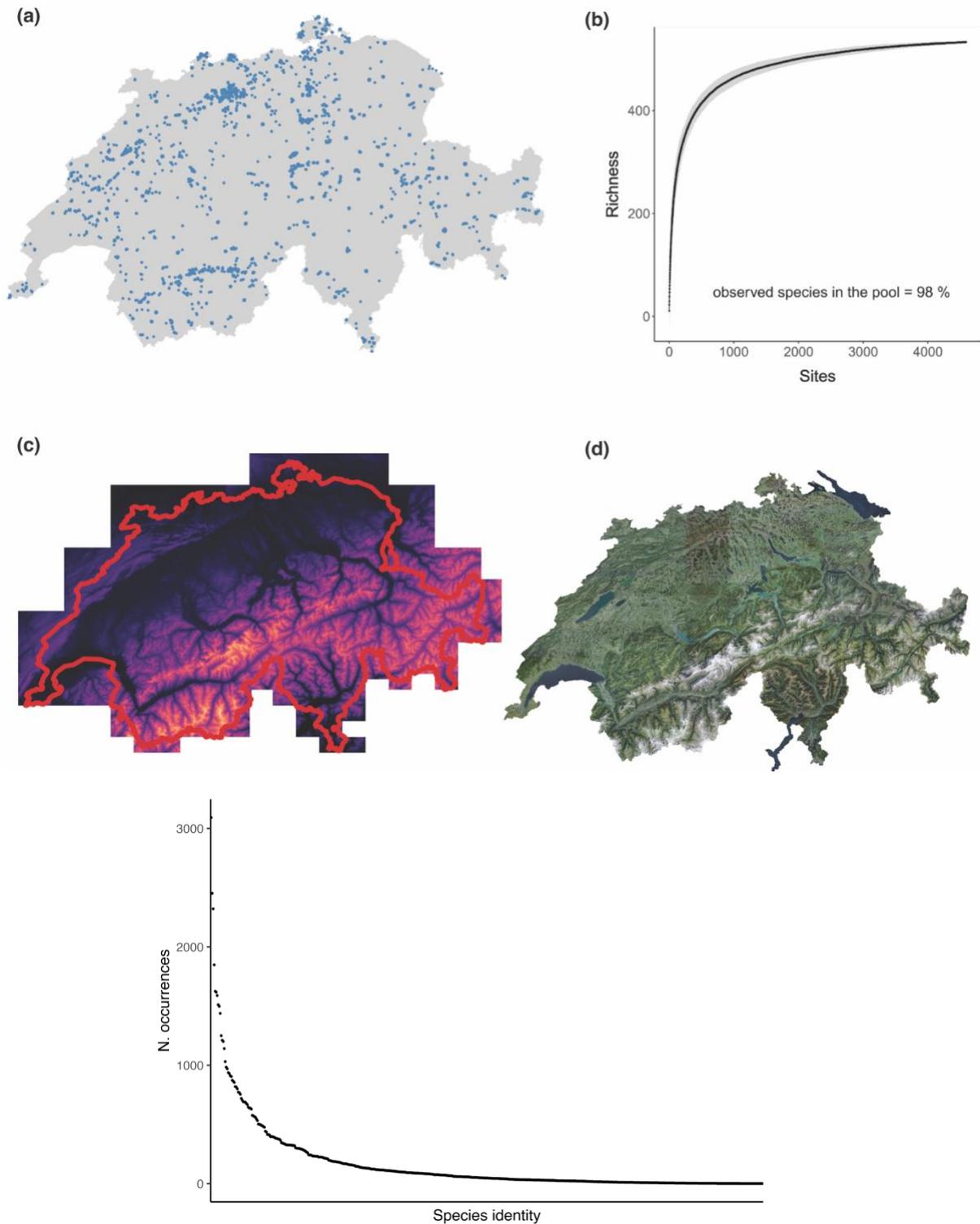
152	<i>Bombus mucidus</i>	161	203 <i>Nomada flava</i>	365	Colletidae	
153	<i>Bombus muscorum</i>	35	204 <i>Nomada flavilabris</i>	2	254 <i>Colletes cunicularius</i>	48
154	<i>Bombus norvegicus</i>	61	205 <i>Nomada flavoguttata</i>	572	255 <i>Colletes daviesanus</i>	344
155	<i>Bombus pascuorum</i>	3092	206 <i>Nomada flavopicta</i>	158	256 <i>Colletes fodiens</i>	7
156	<i>Bombus pratorum</i>	1617	207 <i>Nomada fucata</i>	80	257 <i>Colletes hederæ</i>	25
157	<i>Bombus pyrenaicus</i>	218	208 <i>Nomada fulvicornis</i>	76	258 <i>Colletes impunctatus</i>	99
158	<i>Bombus quadricolor</i>	76	209 <i>Nomada furva</i>	2	259 <i>Colletes marginatus</i>	45
159	<i>Bombus ruderarius</i>	394	210 <i>Nomada fuscicornis</i>	12	260 <i>Colletes mlkossewiczii</i>	18
160	<i>Bombus ruderatus</i>	67	211 <i>Nomada goodeniana</i>	338	261 <i>Colletes nigricans</i>	15
161	<i>Bombus rupestris</i>	477	212 <i>Nomada gransassoi</i>	2	262 <i>Colletes sierrensis</i>	40
162	<i>Bombus sichelii</i>	321	213 <i>Nomada guttulata</i>	25	263 <i>Colletes similis</i>	4
163	<i>Bombus soroeensis</i>	984	214 <i>Nomada hirtipes</i>	93	264 <i>Colletes succinctus</i>	288
164	<i>Bombus subterraneus</i>	53	215 <i>Nomada integra</i>	146	265 <i>Hylaeus alpinus</i>	115
165	<i>Bombus sylvarum</i>	489	216 <i>Nomada kohli</i>	7	266 <i>Hylaeus angustatus</i>	60
166	<i>Bombus sylvestris</i>	855	217 <i>Nomada lathburiana</i>	119	267 <i>Hylaeus annulatus</i>	93
167	<i>Bombus terrestris</i>	1141	218 <i>Nomada leucophthalma</i>	12	268 <i>Hylaeus brevicornis</i>	67
168	<i>Bombus vestalis</i>	164	219 <i>Nomada marshamella</i>	156	269 <i>Hylaeus chypensis</i>	215
169	<i>Bombus veteranus</i>	28	220 <i>Nomada moeschleri</i>	20	270 <i>Hylaeus communis</i>	50
170	<i>Bombus wurflenii</i>	755	221 <i>Nomada mutica</i>	3	271 <i>Hylaeus confusus</i>	970
171	<i>Ceratina chalybea</i>	110	222 <i>Nomada nobilis</i>	2	272 <i>Hylaeus cornutus</i>	720
172	<i>Ceratina cucurbitina</i>	198	223 <i>Nomada obscura</i>	5	273 <i>Hylaeus crassanus</i>	52
173	<i>Ceratina cyanea</i>	577	224 <i>Nomada obtusifrons</i>	3	274 <i>Hylaeus difformis</i>	11
174	<i>Ceratina gravidula</i>	2	225 <i>Nomada panzeri</i>	225	275 <i>Hylaeus dilatatus</i>	154
175	<i>Ceratina nigrolabiata</i>	1	226 <i>Nomada piccioliana</i>	3	276 <i>Hylaeus duckei</i>	145
176	<i>Epeoloides coecutiens</i>	33	227 <i>Nomada pleurosticta</i>	4	277 <i>Hylaeus gibbus</i>	10
177	<i>Epeolus alpinus</i>	13	228 <i>Nomada posthuma</i>	9	278 <i>Hylaeus glacialis</i>	116
178	<i>Epeolus cruciger</i>	33	229 <i>Nomada roberjeotiana</i>	3	279 <i>Hylaeus gredleri</i>	9
179	<i>Epeolus productulus</i>	3	230 <i>Nomada ruficornis</i>	231	280 <i>Hylaeus hyalinatus</i>	375
180	<i>Epeolus variegatus</i>	59	231 <i>Nomada rufipes</i>	6	281 <i>Hylaeus incongruus</i>	327
181	<i>Eucera interrupta</i>	34	232 <i>Nomada sexfasciata</i>	116	282 <i>Hylaeus kahri</i>	53
182	<i>Eucera longicornis</i>	293	233 <i>Nomada sheppardana</i>	37	283 <i>Hylaeus leptoccephalus</i>	83
183	<i>Eucera nigrescens</i>	769	234 <i>Nomada signata</i>	49	284 <i>Hylaeus moricei</i>	42
184	<i>Eucera nigrifacies</i>	1	235 <i>Nomada similis</i>	17	285 <i>Hylaeus nigrinus</i>	1
185	<i>Melecta albifrons</i>	76	236 <i>Nomada stigma</i>	1	286 <i>Hylaeus nivalis</i>	298
186	<i>Melecta festiva</i>	6	237 <i>Nomada striata</i>	84	287 <i>Hylaeus paulus</i>	46
187	<i>Melecta luctuosa</i>	33	238 <i>Nomada succincta</i>	58	288 <i>Hylaeus pectoralis</i>	30
188	<i>Nomada alboguttata</i>	73	239 <i>Nomada villosa</i>	14	289 <i>Hylaeus pfankuchii</i>	19
189	<i>Nomada argentata</i>	2	240 <i>Nomada zonata</i>	19	290 <i>Hylaeus pictipes</i>	35
190	<i>Nomada armata</i>	38	241 <i>Pasites maculatus</i>	30	291 <i>Hylaeus pilosulus</i>	37
191	<i>Nomada atroscutellaris</i>	59	242 <i>Pseudapis diversipes</i>	99	292 <i>Hylaeus punctatus</i>	1
192	<i>Nomada bifasciata</i>	171	243 <i>Tetralonia dentata</i>	34	293 <i>Hylaeus punctulatus</i>	153
193	<i>Nomada braunsiana</i>	3	244 <i>Tetralonia fulvescens</i>	6	294 <i>Hylaeus rinki</i>	58
194	<i>Nomada castellana</i>	17	245 <i>Tetralonia salicariae</i>	29	295 <i>Hylaeus signatus</i>	36
195	<i>Nomada conjungens</i>	25	246 <i>Thyreus hirtus</i>	9	296 <i>Hylaeus simiatus</i>	242
196	<i>Nomada discrepans</i>	2	247 <i>Thyreus histrionicus</i>	1	297 <i>Hylaeus styriacus</i>	233
197	<i>Nomada distinguenda</i>	24	248 <i>Thyreus orbatus</i>	15	298 <i>Hylaeus taeniolatus</i>	168
198	<i>Nomada emarginata</i>	24	249 <i>Thyreus ramosus</i>	9	299 <i>Hylaeus tyrolensis</i>	71
199	<i>Nomada fabriciana</i>	436	250 <i>Thyreus truncatus</i>	5	300 <i>Hylaeus variegatus</i>	22
200	<i>Nomada facilis</i>	31	251 <i>Xyllocopa iris</i>	20	Halictidae	
201	<i>Nomada femoralis</i>	25	252 <i>Xyllocopa valga</i>	42	301 <i>Dufourea alpina</i>	41
202	<i>Nomada ferruginata</i>	7	253 <i>Xyllocopa violacea</i>	183	302 <i>Dufourea dentiventris</i>	60

Appendix S2. Cont.

303	<i>Dufourea halictula</i>	83	354	<i>Lasioglossum lineare</i>	54	405	<i>Sphecodes hyalinatus</i>	384
304	<i>Dufourea inermis</i>	5	355	<i>Lasioglossum lissonotum</i>	123	406	<i>Sphecodes longulus</i>	167
305	<i>Dufourea minuta</i>	11	356	<i>Lasioglossum lucidulum</i>	12	407	<i>Sphecodes majalis</i>	68
306	<i>Dufourea paradoxa</i>	23	357	<i>Lasioglossum majus</i>	112	408	<i>Sphecodes marginatus</i>	8
307	<i>Halictus carinthiacus</i>	1	358	<i>Lasioglossum malachurum</i>	47	409	<i>Sphecodes minimatus</i>	2
308	<i>Halictus confusus</i>	5	359	<i>Lasioglossum marginatum</i>	1031	410	<i>Sphecodes monilicornis</i>	31
309	<i>Halictus eurygnathus</i>	110	360	<i>Lasioglossum marginellum</i>	108	411	<i>Sphecodes niger</i>	297
310	<i>Halictus langobardicus</i>	16	361	<i>Lasioglossum minutissimum</i>	1	412	<i>Sphecodes pellucidus</i>	106
311	<i>Halictus leucaheneus</i>	123	362	<i>Lasioglossum minutulum</i>	45	413	<i>Sphecodes pseudofuscatus</i>	108
312	<i>Halictus maculatus</i>	22	363	<i>Lasioglossum morio</i>	25	414	<i>Sphecodes puncticeps</i>	29
313	<i>Halictus quadricinctus</i>	322	364	<i>Lasioglossum nigripes</i>	1589	415	<i>Sphecodes reticulatus</i>	187
314	<i>Halictus rubicundus</i>	86	365	<i>Lasioglossum nitidiusculum</i>	299	416	<i>Sphecodes rubicundus</i>	103
315	<i>Halictus scabiosae</i>	815	366	<i>Lasioglossum nitidulum</i>	82	417	<i>Sphecodes ruficrus</i>	6
316	<i>Halictus seladonius</i>	904	367	<i>Lasioglossum pallens</i>	303	418	<i>Sphecodes rufiventris</i>	44
317	<i>Halictus sexcinctus</i>	10	368	<i>Lasioglossum parvulum</i>	70	419	<i>Sphecodes scabricollis</i>	60
318	<i>Halictus simplex</i>	181	369	<i>Lasioglossum pauxillum</i>	184	420	<i>Sphecodes schenckii</i>	133
319	<i>Halictus subauratus</i>	638	370	<i>Lasioglossum pleurospeculum</i>	1510	421	<i>Sphecodes spinulosus</i>	96
320	<i>Halictus submediterraneus</i>	502	371	<i>Lasioglossum podolicum</i>	9	422	<i>Sphecodes zangherii</i>	2
321	<i>Halictus tumulorum</i>	60	372	<i>Lasioglossum politum</i>	3	423	<i>Systropha curvicornis</i>	1
322	<i>Lasioglossum aeratum</i>	48	373	<i>Lasioglossum punctatissimum</i>	560	Megachilidae		
323	<i>Lasioglossum albipes</i>	84	374	<i>Lasioglossum puncticolle</i>	231	424	<i>Anthidellum strigatum</i>	327
324	<i>Lasioglossum albocinctum</i>	930	375	<i>Lasioglossum pygmaeum</i>	85	425	<i>Anthidium florentinum</i>	345
325	<i>Lasioglossum alpigenum</i>	9	376	<i>Lasioglossum quadrinotatum</i>	28	426	<i>Anthidium manicatum</i>	8
326	<i>Lasioglossum bavaricum</i>	10	377	<i>Lasioglossum quadrisignatum</i>	4	427	<i>Anthidium montanum</i>	390
327	<i>Lasioglossum bluethgeni</i>	14	378	<i>Lasioglossum rufitarse</i>	1	428	<i>Anthidium oblongatum</i>	56
328	<i>Lasioglossum brevicorne</i>	34	379	<i>Lasioglossum sabulosum</i>	91	429	<i>Anthidium punctatum</i>	482
329	<i>Lasioglossum breviventre</i>	73	380	<i>Lasioglossum semilucens</i>	23	430	<i>Anthidium septemdentatum</i>	284
330	<i>Lasioglossum buccale</i>	5	381	<i>Lasioglossum setulosum</i>	73	431	<i>Anthidium septemspinatum</i>	31
331	<i>Lasioglossum calceatum</i>	8	382	<i>Lasioglossum sexnotatum</i>	3	432	<i>Chelostoma campanularum</i>	1
332	<i>Lasioglossum clypeare</i>	2452	383	<i>Lasioglossum sexstrigatum</i>	22	433	<i>Chelostoma distinctum</i>	442
333	<i>Lasioglossum convexiusculum</i>	9	384	<i>Lasioglossum subfasciatum</i>	73	434	<i>Chelostoma florissomme</i>	196
334	<i>Lasioglossum costulatum</i>	26	385	<i>Lasioglossum subfulvicorne</i>	53	435	<i>Chelostoma foveolatum</i>	704
335	<i>Lasioglossum cupromicans</i>	87	386	<i>Lasioglossum subhirtum</i>	23	436	<i>Chelostoma grande</i>	4
336	<i>Lasioglossum discum</i>	118	387	<i>Lasioglossum tarsatum</i>	5	437	<i>Chelostoma rapunculi</i>	32
337	<i>Lasioglossum elegans</i>	13	388	<i>Lasioglossum tricinctum</i>	16	438	<i>Coelioxys afra</i>	499
338	<i>Lasioglossum euboensis</i>	5	389	<i>Lasioglossum villosulum</i>	98	439	<i>Coelioxys alata</i>	81
339	<i>Lasioglossum fratellum</i>	15	390	<i>Lasioglossum xanthopus</i>	1201	440	<i>Coelioxys aurolobata</i>	3
340	<i>Lasioglossum fulvicorne</i>	97	391	<i>Lasioglossum zonulum</i>	89	441	<i>Coelioxys conica</i>	29
341	<i>Lasioglossum glabriusculum</i>	938	392	<i>Rophitoides canus</i>	12	442	<i>Coelioxys conoidea</i>	106
342	<i>Lasioglossum griseolum</i>	411	393	<i>Rophites algius</i>	3	443	<i>Coelioxys echinata</i>	31
343	<i>Lasioglossum intermedium</i>	13	394	<i>Rophites quinquespinosus</i>	31	444	<i>Coelioxys elongata</i>	3
344	<i>Lasioglossum interruptum</i>	45	395	<i>Sphecodes albilabris</i>	6	445	<i>Coelioxys inermis</i>	38
345	<i>Lasioglossum laeve</i>	138	396	<i>Sphecodes alternatus</i>	194	446	<i>Coelioxys lanceolata</i>	31
346	<i>Lasioglossum laev dorsum</i>	8	397	<i>Sphecodes crassus</i>	1	447	<i>Coelioxys mandibularis</i>	11
347	<i>Lasioglossum laevigatum</i>	2	398	<i>Sphecodes cristatus</i>	233	448	<i>Coelioxys rufescens</i>	19
348	<i>Lasioglossum laterale</i>	265	399	<i>Sphecodes croaticus</i>	1	449	<i>Dioxys cincta</i>	46
349	<i>Lasioglossum laticeps</i>	2	400	<i>Sphecodes dusmeti</i>	12	450	<i>Dioxys tridentata</i>	10
350	<i>Lasioglossum lativentre</i>	1438	401	<i>Sphecodes ephippius</i>	1	451	<i>Heriades crenulatus</i>	1848
351	<i>Lasioglossum leucopus</i>	323	402	<i>Sphecodes ferruginatus</i>	639	452	<i>Heriades rubicola</i>	62
352	<i>Lasioglossum leucozonium</i>	225	403	<i>Sphecodes geoffrellus</i>	380	453	<i>Heriades truncorum</i>	1
353	<i>Lasioglossum limbellum</i>	1250	404	<i>Sphecodes gibbus</i>	190	454	<i>Hoplitis acuticornis</i>	533

Appendix S2. Cont.

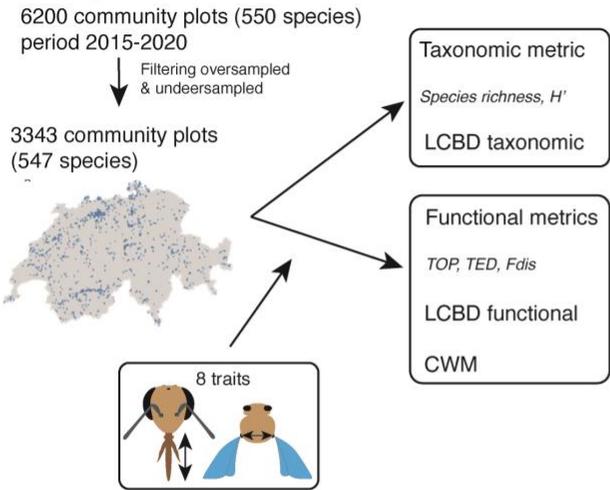
455	<i>Hoplitis adunca</i>	2	506	<i>Osmia labialis</i>	35
456	<i>Hoplitis anthocopoides</i>	397	507	<i>Osmia latreillei</i>	50
457	<i>Hoplitis claviventris</i>	36	508	<i>Osmia leaiana</i>	1
458	<i>Hoplitis dalmatica</i>	212	509	<i>Osmia mustelina</i>	210
459	<i>Hoplitis lepeletieri</i>	27	510	<i>Osmia nigriventris</i>	110
460	<i>Hoplitis leucomelana</i>	123	511	<i>Osmia niveata</i>	28
461	<i>Hoplitis loti</i>	298	512	<i>Osmia parietina</i>	58
462	<i>Hoplitis mitis</i>	130	513	<i>Osmia pilicornis</i>	144
463	<i>Hoplitis praestans</i>	84	514	<i>Osmia rufohirta</i>	17
464	<i>Hoplitis ravouxi</i>	2	515	<i>Osmia scutellaris</i>	125
465	<i>Hoplitis robusta</i>	43	516	<i>Osmia spinulosa</i>	2
466	<i>Hoplitis tridentata</i>	21	517	<i>Osmia steinmanni</i>	93
467	<i>Hoplitis tuberculata</i>	66	518	<i>Osmia submicans</i>	5
468	<i>Hoplitis villosa</i>	118	519	<i>Osmia tergestensis</i>	138
469	<i>Icteranthisidium laterale</i>	37	520	<i>Osmia uncinata</i>	28
470	<i>Lithurgus chrysurus</i>	820	521	<i>Osmia viridana</i>	71
471	<i>Megachile alpicola</i>	117	522	<i>Osmia xanthomelana</i>	3
472	<i>Megachile analis</i>	98	523	<i>Protosmia minutula</i>	30
473	<i>Megachile apicalis</i>	53	524	<i>Pseudoanthidium scapulare</i>	99
474	<i>Megachile centuncularis</i>	3	525	<i>Rhodanthidium caturigense</i>	87
475	<i>Megachile circumcincta</i>	130	526	<i>Stelis breviuscula</i>	9
476	<i>Megachile ericetorum</i>	244	527	<i>Stelis franconica</i>	52
477	<i>Megachile flabellipes</i>	415	528	<i>Stelis minima</i>	8
478	<i>Megachile genalis</i>	7	529	<i>Stelis minuta</i>	2
479	<i>Megachile lagopoda</i>	2	530	<i>Stelis nasuta</i>	8
480	<i>Megachile lapponica</i>	34	531	<i>Stelis odontopyga</i>	8
481	<i>Megachile leachella</i>	1	532	<i>Stelis ornata</i>	2
482	<i>Megachile ligniseca</i>	34	533	<i>Stelis phaeoptera</i>	44
483	<i>Megachile maritima</i>	100	534	<i>Stelis punctulatisima</i>	21
484	<i>Megachile melanopyga</i>	49	535	<i>Stelis signata</i>	116
485	<i>Megachile nigriventris</i>	89	536	<i>Stelis similima</i>	23
486	<i>Megachile parietina</i>	115	537	<i>Trachusa byssina</i>	5
487	<i>Megachile pilicrus</i>	134	538	<i>Trachusa interrupta</i>	383
488	<i>Megachile pilidens</i>	14	Melittidae		
489	<i>Megachile pyrenaica</i>	262	539	<i>Dasypoda argentata</i>	11
490	<i>Megachile pyrenaica</i>	35	540	<i>Dasypoda hirtipes</i>	8
491	<i>Megachile rotundata</i>	99	541	<i>Macropis europaea</i>	6
492	<i>Megachile sculpturalis</i>	89	542	<i>Macropis fulvipes</i>	187
493	<i>Megachile versicolor</i>	11	543	<i>Melitta dimidiata</i>	33
494	<i>Megachile willughbiella</i>	231	544	<i>Melitta haemorrhoidalis</i>	17
495	<i>Osmia alticola</i>	19	545	<i>Melitta leporina</i>	214
496	<i>Osmia anceyi</i>	10	546	<i>Melitta nigricans</i>	222
497	<i>Osmia andrenoides</i>	4	547	<i>Melitta tricincta</i>	106
498	<i>Osmia aurulenta</i>	28			
499	<i>Osmia bicolor</i>	688			
500	<i>Osmia bicornis</i>	398			
501	<i>Osmia brevicornis</i>	668			
502	<i>Osmia caeruleascens</i>	32			
503	<i>Osmia cornuta</i>	331			
504	<i>Osmia gallarum</i>	112			
505	<i>Osmia inermis</i>	91			



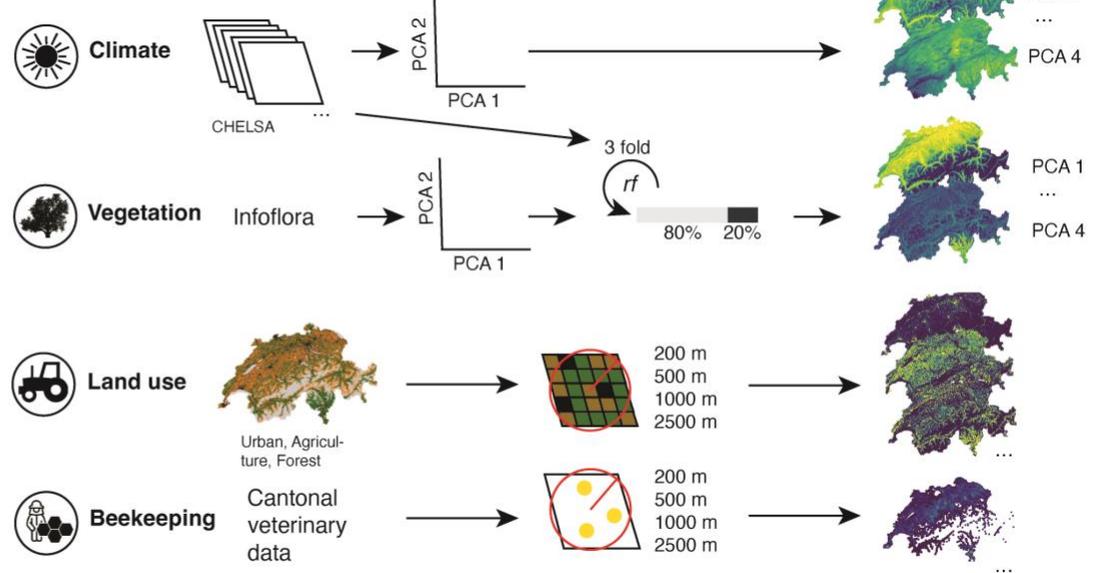
Appendix S3. Summary of the occurrence data: (a) spatial distribution of sampled plots and (b) species accumulation curves. These curves were used to estimate the total number of species present in Switzerland (extrapolated species richness in the species pool based on bootstrap resampling). In total, our data include more than 98% of the bee species predicted to be present in Switzerland. (c) Digital

height model (dark colours depict lower elevation, warm colours depict higher elevation). (d) Aerial photograph of Switzerland. (e) Rank-occurrence diagram of the wild bee species collected in the community plots. Every dot corresponds to one species, which is sorted decreasingly according to the number of occurrences (i.e., the number of community plots where the species was sam

(a) Responses

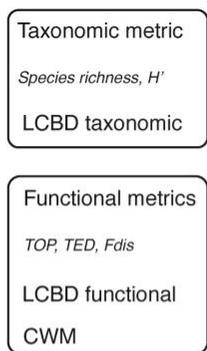


(b) Predictors

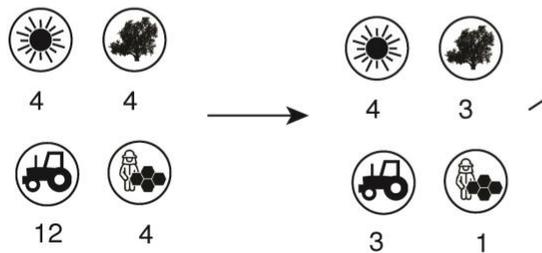


(c)

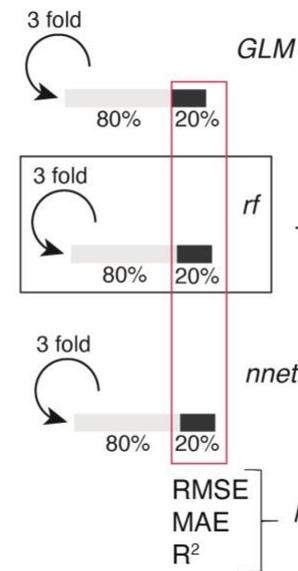
Responses



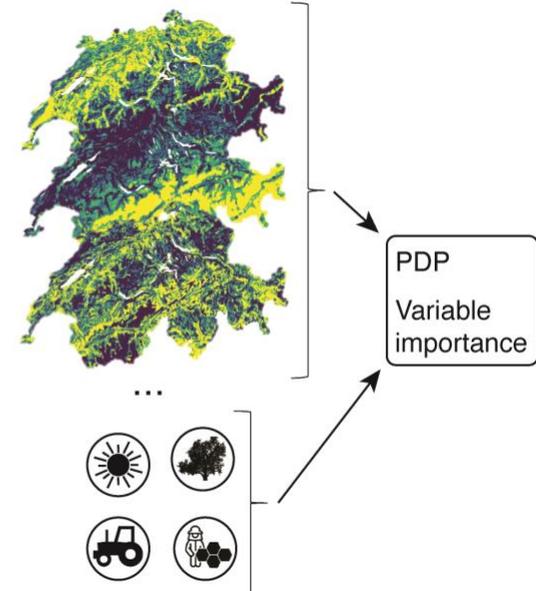
Predictors



Model calibration and validation



Projection



Appendix S4. Methodological framework depicting the computation of the responses considered (a), the predictors (b) and the modelling pipeline. (a) Using 4591 community plots and 8 functional traits (i.e. intertegular distance ITD, tongue length, solitary, belowground, cleptoparasite, phenology start, phenology duration, and feeding specialisation, see also Appendix S2), we calculated α taxonomic (i.e. species richness, H') and α functional (i.e. TOP for functional richness, TED for functional evenness, FDis for functional divergence) metrics, as well as LCBD as a metric of taxonomic and functional uniqueness. (b) We used four types of predictors: climate, vegetation, land use and beekeeping. For climate, we used the 16 climatic variables from the CHELSA dataset (Kraeger et al., 2015), conducted a principal components analysis (PCA) and kept the first four axes (Appendix S3). For vegetation metrics, we used plant community plots, applied a PCA, and kept the first four axes. We then modelled the countrywide distribution as random forests, using climatic variables as predictors (see Methods). We calculated the land-use metrics by measuring the amount of agricultural, forest and urban land use within buffers of 200, 500, 1000 and 2500 m. Beekeeping intensity maps were produced by calculating the number of honeybee hives within the buffers. (c) Modelling pipeline. Note that although we considered three different algorithms (GLM, rf, nnet), we only used rf because it outperformed the other algorithms (see Methods). H' = Shannon diversity; TOP = trait onion peeling; TED = trait evenness distribution; FDis = functional dispersion; LCBD = local community contributions to β -diversity; rf = random forest; PCA = principal components analysis; GLM = generalised linear model; nnet = neural network; RMSE = root mean squared error; MAE = mean absolute error; PDP = partial dependence pl

Appendix S5. Trait data for the different species were collected within a European level effort. We used eight functional traits: (1) *intertegular distance* (ITD) reflects species dispersal capacity and body size and is defined as the distance in millimetres between the two tegula. It is obtained by measuring the SPAN between the wing bases in a bee's thorax (Cariveau et al., 2016) (2) *Feeding specialisation* provides information about the number of plant families exploited by a species. We classified species into two categories, polylectic species (feeding on more than five plant genus from more than three plant families, classified as "1") and species with other strategies, including oligolectic and monolectic (species feeding on fewer than five plant genus or a single plant species, classified as "0"). This definition corresponds to "Polylecty sensu lato" in Robertson (1925) and to "meso-lecty + Polylecty" in Cane and Sipes (2006) and Müller and Kuhlmann (2008). (3) *Tongue length* indicates the diversity of floral resources a species can access. Species were classified as having either a long tongue (indicating a narrower range of resources, classified as "0") or short tongue (indicating a broader range of resources, classified as "1"). We summarised the phenology of wild bee species as two traits, (4) *the month when the activity period of the adults starts* and (5) *the duration of the activity period in months*. Phenology of species is obtained from individuals from within their ranges in Europe. The nesting or reproductive strategy is an important trait determining, for instance, the response of wild bees to land-use change (Buchholz & Egerer, 2020). In that regard, species were classified as below-ground nesters, above-ground nesters and kleptoparasites. To include the information contained in this categorical variable in our analyses, we created dummy variables. Specifically, we created three numerical variables (one for each category) where a species belonging to a given category takes the value of 1 and 0 otherwise. When defining dummy variables, a common mistake is to define too many variables (commonly referred to as the "dummy variable trap", see Gujarati, 2012). In our case, the categorical variable can take on 3 values. To include all the information in our analysis, we only need 2 dummy variables (n-1). Specifically, we defined nesting mode and kleptoparasitic behaviour. Nesting mode was obtained by classifying non-kleptoparasitic species as either belowground nesters (value 1) or not belowground nesters, that is, aboveground nesters (value 0). Belowground nesters build a nest below ground or use existing cavities below ground. Thus, aboveground nesters were not included to avoid collinearity. We also included the kleptoparasitic behaviour (7). Kleptoparasite bees lay their eggs

inside nests constructed by other bee species, and their larvae feed on pollen provided by the host. (8)

Solitary describes the degree of social organisation of each wild bee species. We used two main categories, solitary bees (non-social bees that do not show any type of social organisation, classified as “1”) and social bees (including primitive social, subsocial and eusocial wild bee species, classified as “0”).

Appendix S6. Functional richness, i.e. the amount of multidimensional functional trait space filled by the community, and functional evenness, i.e. the regularity in the spacing of species within that functional trait space, were measured using the trait onion peeling (TOP) and trait evenness diversity (TED) indices, respectively, both introduced by Fontana et al. (2016). Functional dispersion, i.e. the multivariate degree of spread around the centre of gravity of the species trait distribution, was measured using the FDis index proposed by Laliberté and Legendre (2010).

For taxonomic LCBD, we used the matrix of species occurrence to compute a pairwise dissimilarity matrix using the Sorensen index and the function 'beta.pair' from the package betapart (Baselga, 2010) in R v. 4.0.3 (R Core Team, 2020). We then used the resulting dissimilarity matrix to calculate LCBD using the function 'LCBD.comp' from the R package adespatial (Legendre & De Cáceres, 2013). For functional LCBD, we used the CWM matrix to calculate a Sorensen derived pairwise functional dissimilarity matrix, which was used to calculate functional LCBD as done for taxonomic LCBD. We used principal components analysis (PCA) to reduce the dimensionality of the trait matrix to allow the computation of LCBD in communities with fewer species than traits. Three PCA axes were kept, representing 86% of the total variation in the original standardised trait matrix (PCA1 = 63%; PCA2 = 14%, PCA3 = 9%; other axes < 5%).

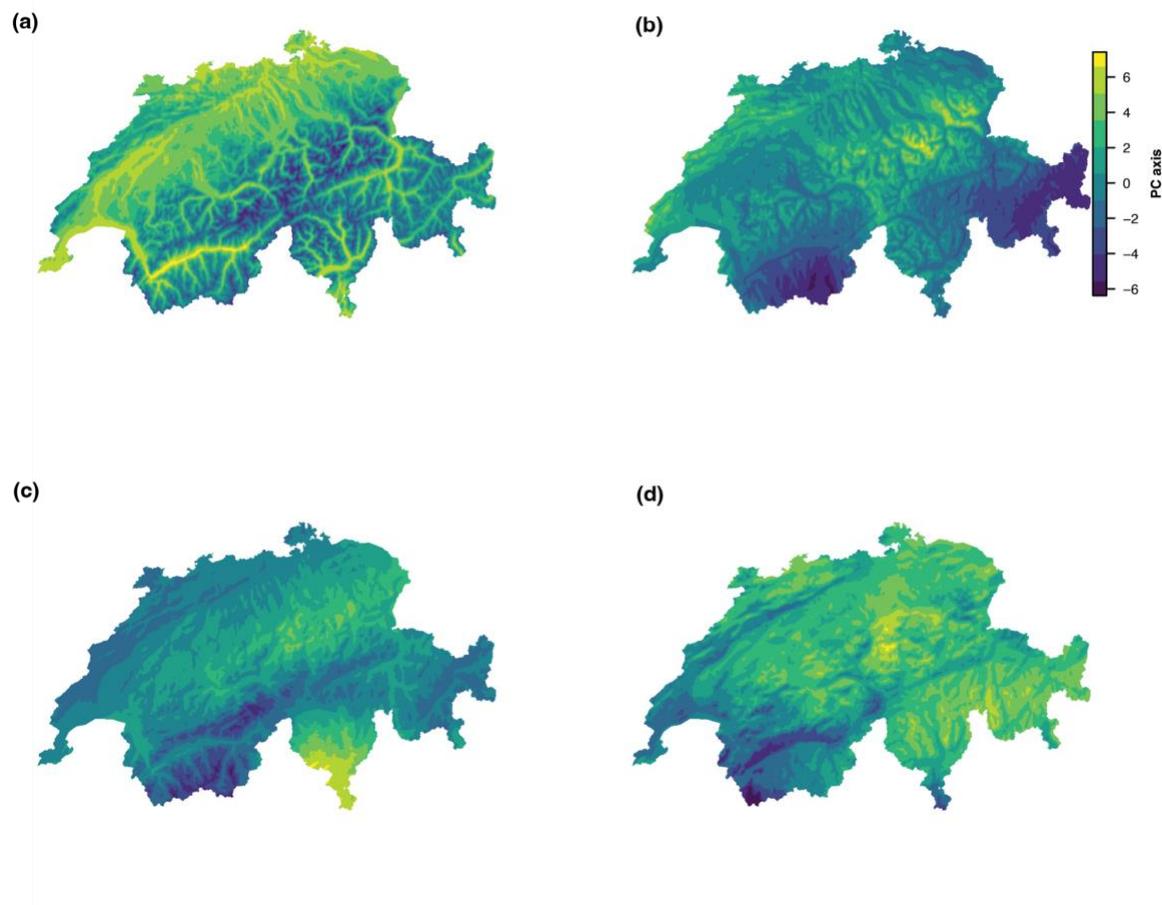
Appendix S7. The first principal components analysis (PCA) axis was related to elevation-driven temperature, with higher values indicating warmer conditions (Appendix S3, hereafter referred to as temperature). The second PCA axis was related to precipitation, with lower PCA values indicating lower precipitation (Appendix S3, hereafter precipitation). The third axis was related to variability in both temperature and precipitation, with higher PCA values indicating higher variability (Appendix S3, hereafter seasonality). The fourth axis was related to changes in temperature range (Appendix S3, temperature range), where places with positive values indicating lower temperature range. (Appendix S3, hereafter temperature range). The description of each axis is based on the contribution of the raw variables (or factor loading).

Appendix S8. Factor loading in climate principal components analysis (PCA) on the CHELSA climate variables (Kraeger et al., 2017). Bold = 3 variables with the highest absolute loading on each axis. PCA1 represents changes in temperature. PCA2 represents changes in precipitation. PCA3 represents changes in climate (temperature and precipitation) seasonality. PCA4 represents changes in temperature range, where places with a higher PCA values having lower temperature range.

CHELSA climate variable	PCA1 (39%)	PCA2 (28%)	PCA3 (20%)	PCA4 (6%)
clim1: Annual Mean Temperature	0.3211	0.1770	0.1341	0.0141
clim2: Mean Diurnal Range	-0.1238	-0.2319	0.2647	-0.4788
clim3: Isothermality	-0.2655	0.0363	-0.2153	-0.1058
clim4: Temperature Seasonality	0.1005	-0.2037	0.3973	-0.2886
clim5: Max Temperature of Warmest Month	0.3225	0.1607	0.1549	-0.0273
clim6: Min Temperature of Coldest Month	0.3178	0.2023	0.0893	0.0442
clim7: Temperature Annual Range	0.0135	-0.2413	0.3639	-0.4022
clim8: Mean Temperature of Wettest Quarter	0.1556	0.0012	0.2525	0.3495
clim9: Mean Temperature of Driest Quarter	0.2408	0.2322	-0.0551	-0.2778
clim10: Mean Temperature of Warmest Quarter	0.3224	0.1620	0.1545	-0.0068
clim11: Mean Temperature of Coldest Quarter	0.3187	0.1934	0.1092	0.0291
clim12: Annual Precipitation	-0.2286	0.2615	0.2358	-0.0148
clim13: Precipitation of Wettest Month	-0.2393	0.1699	0.3154	0.1074
clim14: Precipitation of Driest Month	-0.1644	0.3720	0.0134	-0.1388
clim15: Precipitation Seasonality	-0.0913	-0.2307	0.3215	0.3674

clim16: Precipitation of Wettest Quarter	-0.2407	0.1800	0.3090	0.1182
clim17: Precipitation of Driest Quarter	-0.1737	0.3682	0.0198	-0.1498
clim18: Precipitation of Warmest Quarter	-0.2412	0.1708	0.2886	0.2221
clim19: Precipitation of Coldest Quarter	-0.1300	0.3789	-0.0546	-0.2443

CHELSA codes: clim1 = Annual Mean Temperature [$^{\circ}\text{C} \cdot 10$]; clim2 = Mean Diurnal Range [$^{\circ}\text{C}$]; clim3 = Isothermality; clim4 = Temperature Seasonality [standard deviation]; clim5 = Max Temperature of Warmest Month [$^{\circ}\text{C} \cdot 10$]; clim6 = Min Temperature of Coldest Month [$^{\circ}\text{C} \cdot 10$]; clim7 = Temperature Annual Range [$^{\circ}\text{C} \cdot 10$]; clim8 = Mean Temperature of Wettest Quarter [$^{\circ}\text{C} \cdot 10$]; clim9 = Mean Temperature of Driest Quarter [$^{\circ}\text{C} \cdot 10$]; clim10 = Mean Temperature of Warmest Quarter [$^{\circ}\text{C} \cdot 10$]; clim11 = Mean Temperature of Coldest Quarter [$^{\circ}\text{C} \cdot 10$]; clim12 = Annual Precipitation [mm/year]; clim13 = Precipitation of Wettest Month [mm/month]; clim14 = Precipitation of Driest Month [mm/month]; clim15 = Precipitation Seasonality [coefficient of variation]; clim16 = Precipitation of Wettest Quarter [mm/quarter]; clim17 = Precipitation of Driest Quarter [mm/quarter]; clim18 = Precipitation of Warmest Quarter [mm/quarter]; clim19 = Precipitation of Coldest Quarter [mm/quarter]



Appendix S9. Maps of the four climatic variables used in the model. (a) Map corresponding to PCA1, which depicts changes in temperature (Appendix S3), with blue values indicating lower temperatures and yellow values indicating higher temperatures. (b) Map corresponding to PCA2, which depicts changes in precipitation (Appendix S3), with blue values indicating higher precipitation and yellow ones indicating lower precipitation. (c) Map corresponding to PCA3 (climate seasonality), which depicts changes in temperature and precipitation seasonality (Appendix S3), with blue values indicating lower seasonality and yellow ones indicating higher seasonality. (d) Map corresponding to PCA4 (temperature ranges), which depicts changes in temperature range, with blue values indicating a wider temperature range but low precipitation seasonality, and yellow values indicating a smaller temperature range and higher precipitation seasonality.

Appendix S10. To model vegetation in relation to the 19 raw CHELSA variables, the random forest algorithm was trained on 80% of the data and evaluated on the remaining 20%, stratified according to the response variable (function ‘createDataPartition’ in R package caret v. 6.0-86, Kuhn, 2008). Model training and parameter tuning were done using three times three-fold cross-validation (function ‘train’ in R package caret). The best model was chosen based on the root mean squared error (RMSE), mean absolute error (MAE) and R² measured on the trained dataset. The performance of the selected model was further evaluated on the test dataset using the same metrics. Predictions across the whole of Switzerland were performed based on the raw CHELSA data (downscaled version, 100 m).

The first principal components analysis (PCA) axis was related to the elevation gradient and was removed from the analysis because it was strongly correlated with climate PCA1 ($r = 0.85$). Pearson correlations (r) between climate and all other vegetation principal components were between 0.39 and 0.01. The second PCA axis (named mid-elevation coniferous) represents a gradient in the dominance of coniferous plant communities, with negative values indicating coniferous plant communities (fir, spruce and Scots pine forest with the associated herbaceous plants, e.g. *Silene dioica*, *Petasites albus*, *Veronica urticifolia*, Appendix S4) and positive values indicating other vegetation types, including lowland vegetation at intermediate positive values and high-elevation plants at large positive values (e.g. *Salix herbacea*, *Leucanthemopsis alpina*, *Gnaphalium supinum*). The third PCA axis (named dry–wet) depicts a dry to wet gradient, with positive values indicating ruderal and/or dry open vegetation (e.g. *Echium vulgare*, *Silene vulgaris*, *Artemisia vulgaris*, Appendix S4) and negative values indicating wet forest vegetation (e.g. *Veronica montana*, *Carex remota*, *Lysimachia nemorum*, Appendix S4). The fourth PCA axes (named forest) depicts a gradient from woody to open vegetation, with positive values indicating plant assemblages from forests and shrublands (e.g. *Polypodium vulgare*, *Hypericum montanum*, *Salvia glutinosa*, Appendix S4) and negative values indicating plant assemblages from meadows, pastures and other types of open vegetation (e.g. *Plantago atrata*, *Carex flacca*, *Centaurea jacea*, Appendix S4).

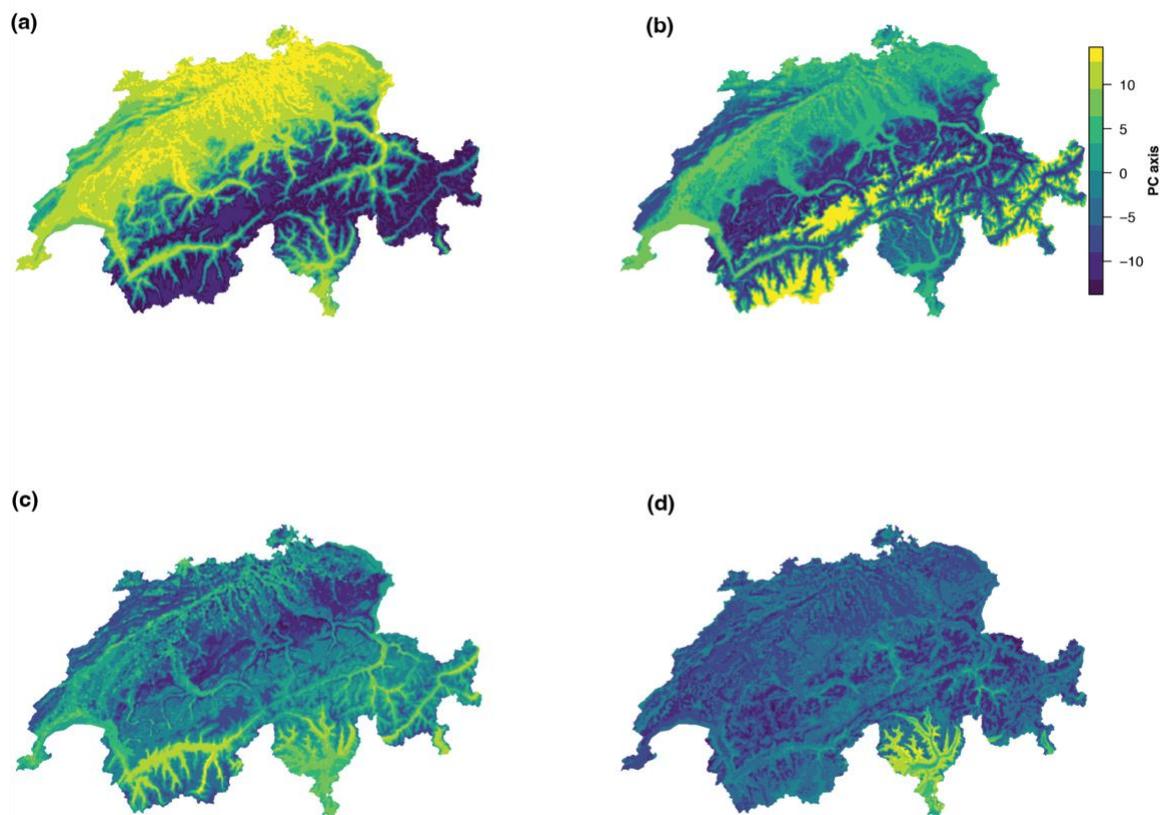
Appendix S11. Factor loading in plant principal components analysis (PCA) for selected species. The five species with the highest/lowest loading on each axis are shown (bold values). The most common tree species in our dataset are also included.

Species	PCA1 (24%)	PCA2 (12%)	PCA3 (4%)	PCA4 (3%)
<i>Abies alba</i>	0.0376	-0.0547	-0.0810	0.0472
<i>Acer campestre</i>	0.0606	0.0183	0.0047	0.0013
<i>Acer platanoides</i>	0.0512	0.0056	-0.0164	-0.0064
<i>Acer pseudoplatanus</i>	0.0563	-0.0523	-0.0150	0.0065
<i>Alnus glutinosa</i>	0.0292	0.0116	-0.0105	0.0142
<i>Alnus incana</i>	0.0174	-0.0461	-0.0232	0.0509
<i>Alnus viridis</i>	-0.0475	-0.0503	0.0497	-0.0071
<i>Artemisia vulgaris</i>	0.0200	0.0032	0.0995	0.0245
<i>Campanula scheuchzeri</i>	-0.0710	-0.0051	0.0118	0.0006
<i>Carex flacca</i>	0.0238	-0.0643	-0.0368	-0.0903
<i>Carex remota</i>	0.0416	-0.0080	-0.1030	0.0285
<i>Carpinus betulus</i>	0.0508	0.0349	0.0002	-0.0393
<i>Centaurea jacea</i>	0.0448	-0.0397	-0.0012	-0.0843
<i>Crataegus laevigata</i>	0.0324	-0.0028	-0.0450	0.0079
<i>Crataegus monogyna</i>	0.0610	0.0051	0.0058	0.0011
<i>Dryopteris affinis</i>	0.0010	-0.0337	-0.0273	0.1286

<i>Echium vulgare</i>	0.0258	-0.0025	0.1243	0.0250
<i>Euphrasia rostkoviana</i>	-0.0256	-0.0842	0.0118	-0.0152
<i>Fagus sylvatica</i>	0.0486	-0.0421	-0.0746	0.0432
<i>Frangula alnus</i>	0.0306	0.0100	-0.0152	0.0305
<i>Fraxinus excelsior</i>	0.0705	-0.0274	0.0002	0.0268
<i>Gentiana verna</i>	-0.0482	-0.0362	0.0196	-0.0831
<i>Geum urbanum</i>	0.0721	-0.0203	-0.0024	0.0268
<i>Glechoma hederacea</i>	0.0706	-0.0074	-0.0184	-0.0093
<i>Gnaphalium supinum</i>	-0.0566	0.0624	-0.0389	0.0188
<i>Homogyne alpina</i>	-0.0720	-0.0118	-0.0037	-0.0213
<i>Hypericum montanum</i>	0.0072	-0.0395	0.0396	0.1303
<i>Ilex aquifolium</i>	0.0351	-0.0032	-0.0499	0.0439
<i>Juglans regia</i>	0.0603	0.0273	0.0127	0.0212
<i>Lapsana communis</i>	0.0716	-0.0043	-0.0235	0.0025
<i>Larix decidua</i>	-0.0171	-0.0192	0.0498	0.0506
<i>Leucanthemopsis alpina</i>	-0.0563	0.0628	-0.0385	0.0185
<i>Luzula nivea</i>	-0.0132	-0.0312	0.0950	0.1296
<i>Lysimachia nemorum</i>	0.0279	-0.0541	-0.1092	-0.0140
<i>Petasites albus</i>	-0.0036	-0.0860	-0.0377	0.0197
<i>Picea abies</i>	0.0140	-0.0724	-0.0238	0.0117

<i>Pinus sylvestris</i>	0.0287	-0.0055	-0.0064	0.0234
<i>Plantago alpina</i>	-0.0508	-0.0199	0.0138	-0.0810
<i>Plantago atrata</i>	-0.0434	-0.0340	0.0244	-0.0940
<i>Poa alpina</i>	-0.0730	-0.0173	0.0031	-0.0288
<i>Polygonum viviparum</i>	-0.0717	-0.0012	0.0058	-0.0475
<i>Polypodium vulgare</i>	-0.0154	-0.0302	0.0794	0.1333
<i>Populus tremula</i>	0.0207	-0.0361	0.0323	0.0563
<i>Potentilla erecta</i>	-0.0232	-0.0845	0.0157	0.0320
<i>Primula elatior</i>	0.0280	-0.0566	-0.0983	-0.0578
<i>Quercus petraea</i>	0.0268	-0.0024	0.0365	0.0633
<i>Quercus robur</i>	0.0604	0.0185	-0.0333	0.0029
<i>Ranunculus montanus aggr.</i>	-0.0699	-0.0170	0.0136	-0.0004
<i>Salix appendiculata</i>	-0.0255	-0.0714	0.0029	0.0134
<i>Salix caprea</i>	0.0371	-0.0575	-0.0152	-0.0190
<i>Salix herbacea</i>	-0.0556	0.0584	-0.0387	0.0042
<i>Salix myrsinifolia</i>	-0.0247	-0.0322	0.0237	-0.0057
<i>Salix purpurea</i>	0.0103	-0.0293	0.0047	-0.0091
<i>Salix retusa</i>	-0.0548	0.0257	-0.0262	-0.0495
<i>Salvia glutinosa</i>	0.0092	-0.0360	0.0388	0.1300
<i>Salvia pratensis</i>	0.0438	0.0112	0.0990	0.0009

<i>Sibbaldia procumbens</i>	-0.0563	0.0610	-0.0362	0.0141
<i>Silene dioica</i>	-0.0057	-0.0850	-0.0003	0.0136
<i>Silene exscapa</i>	-0.0504	0.0572	-0.0306	0.0196
<i>Silene nutans</i>	-0.0191	-0.0451	0.1051	0.0462
<i>Silene vulgaris</i>	-0.0078	-0.0434	0.1059	-0.0055
<i>Sorbus aria</i>	0.0128	-0.0740	0.0112	0.0658
<i>Sorbus aucuparia</i>	0.0079	-0.0776	0.0058	0.0451
<i>Tilia cordata</i>	0.0353	0.0142	0.0305	0.0494
<i>Tilia platyphyllos</i>	0.0485	0.0056	-0.0134	-0.0116
<i>Ulmus glabra</i>	0.0407	-0.0237	-0.0452	0.0366
<i>Veronica beccabunga</i>	0.0195	-0.0551	-0.0992	-0.0368
<i>Veronica montana</i>	0.0375	-0.0084	-0.1017	0.0039
<i>Veronica persica</i>	0.0712	0.0060	-0.0101	-0.0208
<i>Veronica urticifolia</i>	-0.0090	-0.0864	0.0026	0.0655

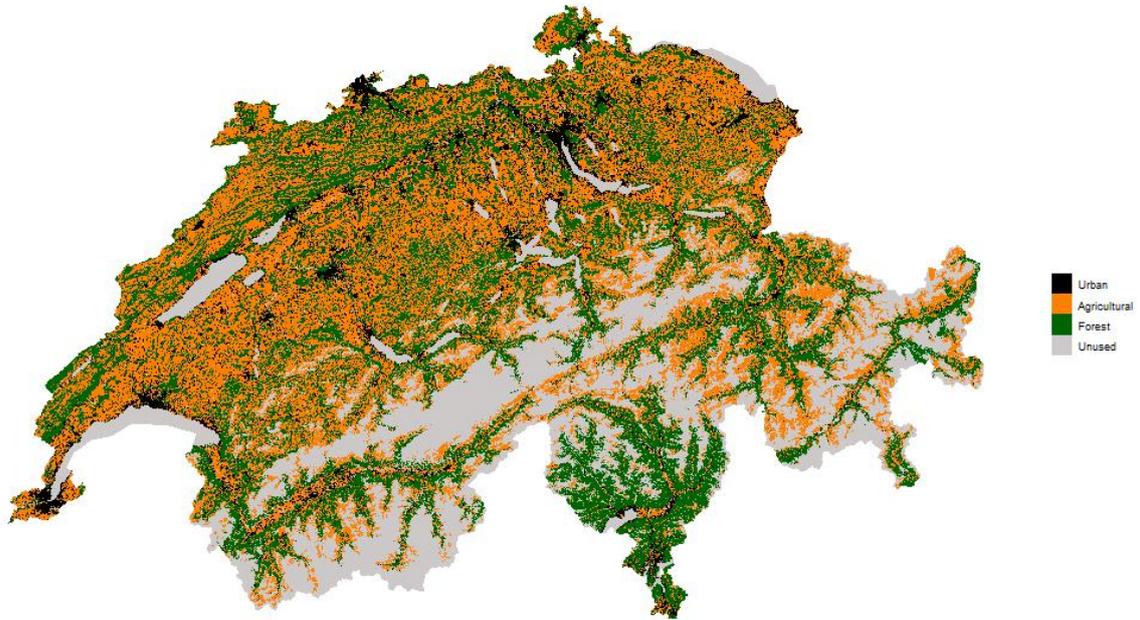


Appendix S12. Maps of the four vegetation variables used in the model. (a) Map corresponding to PCA1, which reflects the elevation gradient, with positive values indicating more colline–montane species (e.g. *Geum urbanum*, *Lapsana communis*, *Veronica persica*) and negative values indicating subalpine–alpine species (e.g. *Poa alpina*, *Homogyne alpina*, *Polygonum viviparum*). This variable was not used because it was highly correlated with climate PCA1. (b) Map corresponding to PCA2 (named mid-elevation coniferous), which shows a gradient in the dominance of coniferous plant communities, with negative values indicating coniferous plant communities (fir, spruce and Scots pine forest with the associated herbaceous plants, e.g. *Silene dioica*, *Petasites albus*, *Veronica urticifolia*) and positive values indicating other vegetation types, including lowland vegetation at intermediate positive values and high-elevation plants at large positive values (e.g. *Salix herbacea*, *Leucanthemopsis alpina*, *Gnaphalium supinum*). (c) Map corresponding to PCA3 (named dry–wet), which depicts a dry to wet gradient, with positive values indicating ruderal and/or dry open vegetation (e.g. *Echium vulgare*, *Silene*

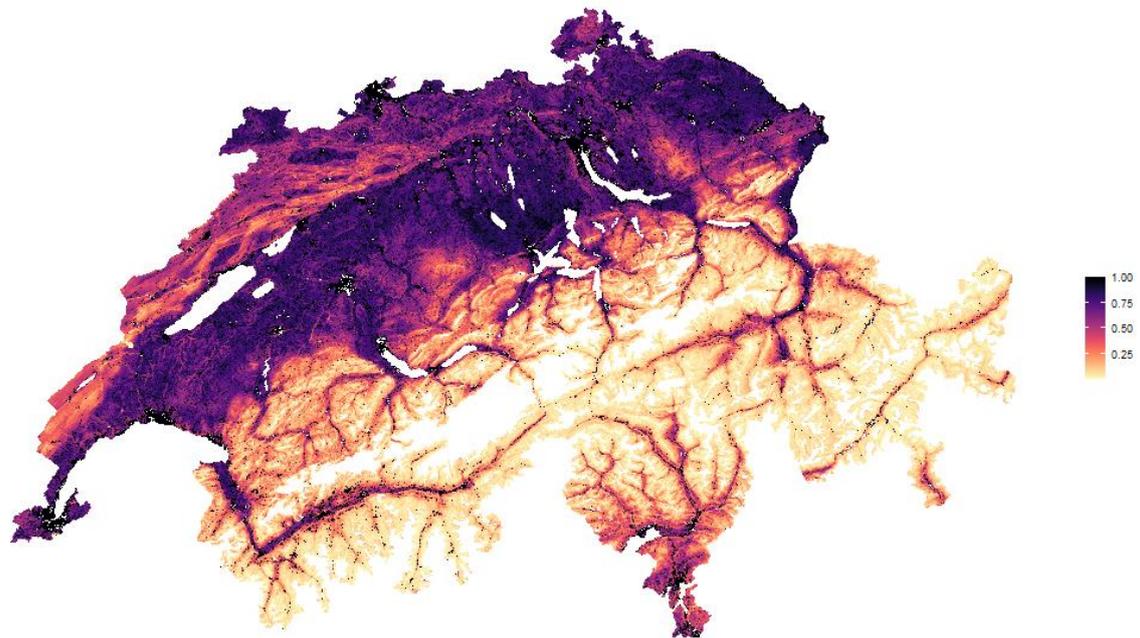
vulgaris, *Artemisia vulgaris*) and negative values indicating wet forest vegetation (e.g. *Veronica montana*, *Carex remota*, *Lysimachia nemorum*). (d) Map corresponding to PCA4 (named forest), which depicts a gradient from woody to open vegetation, with positive values indicating plant assemblages from forests and shrublands (e.g. *Polypodium vulgare*, *Hypericum montanum*, *Salvia glutinosa*) and negative values indicating plant assemblages from meadows, pastures and other types of open vegetation (e.g. *Plantago atrata*, *Carex flacca*, *Centaurea jacea*).

Appendix S13. The Land Use and Land Cover data include 4 principal types (urban, rural, forest, unproductive), 10 classes, and 46 basic categories, and they have a resolution of 100 m by 100 m, thus matching that of the bee data. We focused on three of the four principal domains: urban, agricultural and forest. Urban (Buchholz et al., 2020; Fournier et al., 2020), agricultural (Evans et al., 2018), and forest (Brosi et al., 2007) land-use types were all shown to be important drivers of the functional and taxonomic composition of bee communities. Urban land-use (7.5 % of the total surface area) includes all types of settlements and build-up infrastructure, including buildings, transportation infrastructure, special infrastructure, sport and leisure areas and urban green areas (e.g. gardens, cemeteries, parks). Thus, they not only indicate major settlements but also, e.g., sparse infrastructure, towns, transportation networks, which is the main case at higher elevations. Agricultural land-use (35.9 % of the total surface area) encompasses productive surfaces relating to crop growing, livestock and fruit cultivation. They include all herbaceous, arbustive and arboreous crops, as well as alpine meadows, pastures and other grazing areas. Finally, forest land use (31.3 % of the total surface area) includes all the different types of forest found in Switzerland except agroforestry. For details, see Altwegg and Weibel (2015) and FSO (2013). We excluded unproductive land-use types, such as water bodies, glaciers and permanent snow, as they mostly represent non-bee habitat.

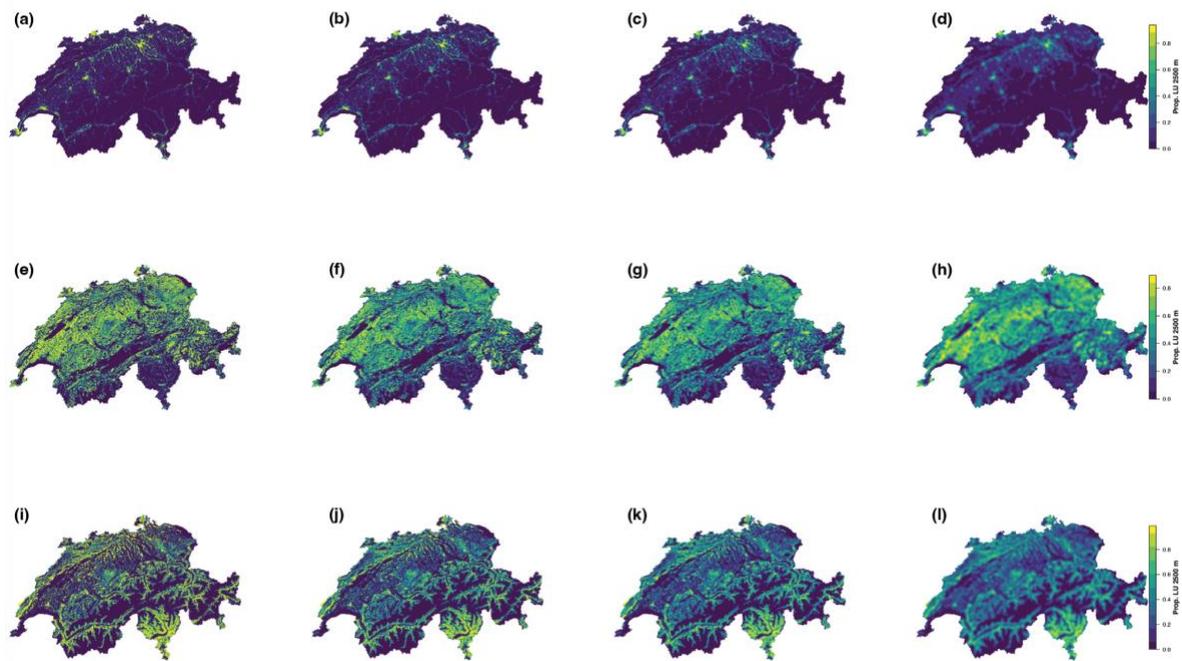
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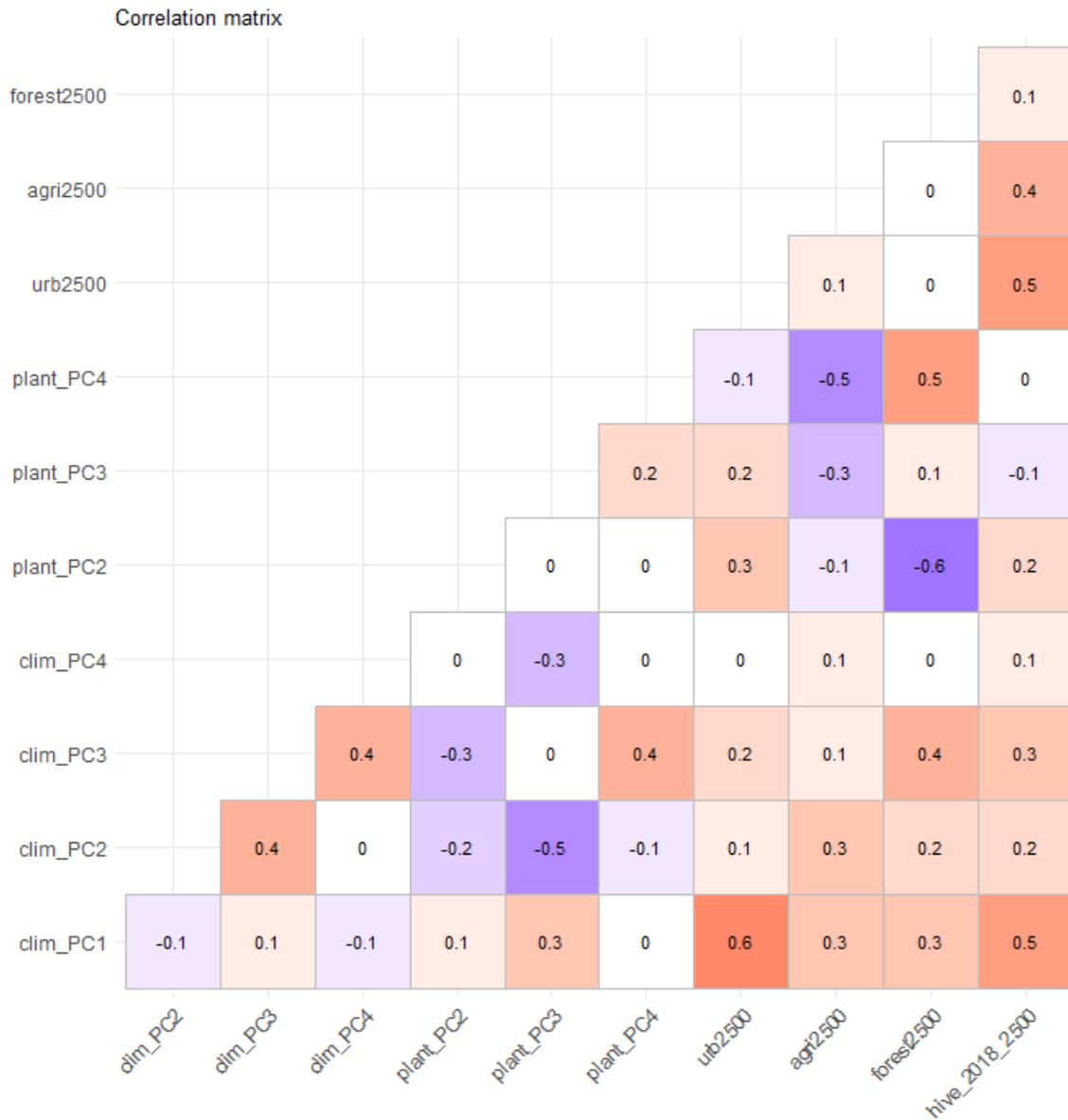
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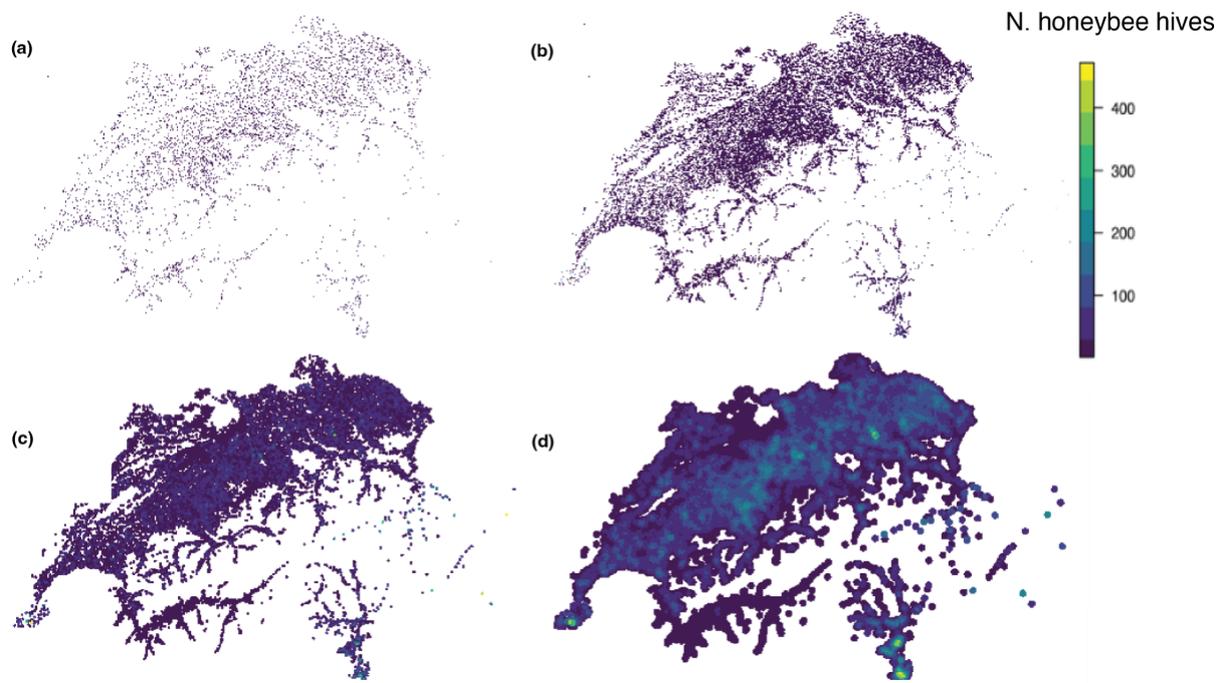
Appendix S14. Maps of (a) land use and (b) land-use intensity in Switzerland, based on Meier et al. (2020).



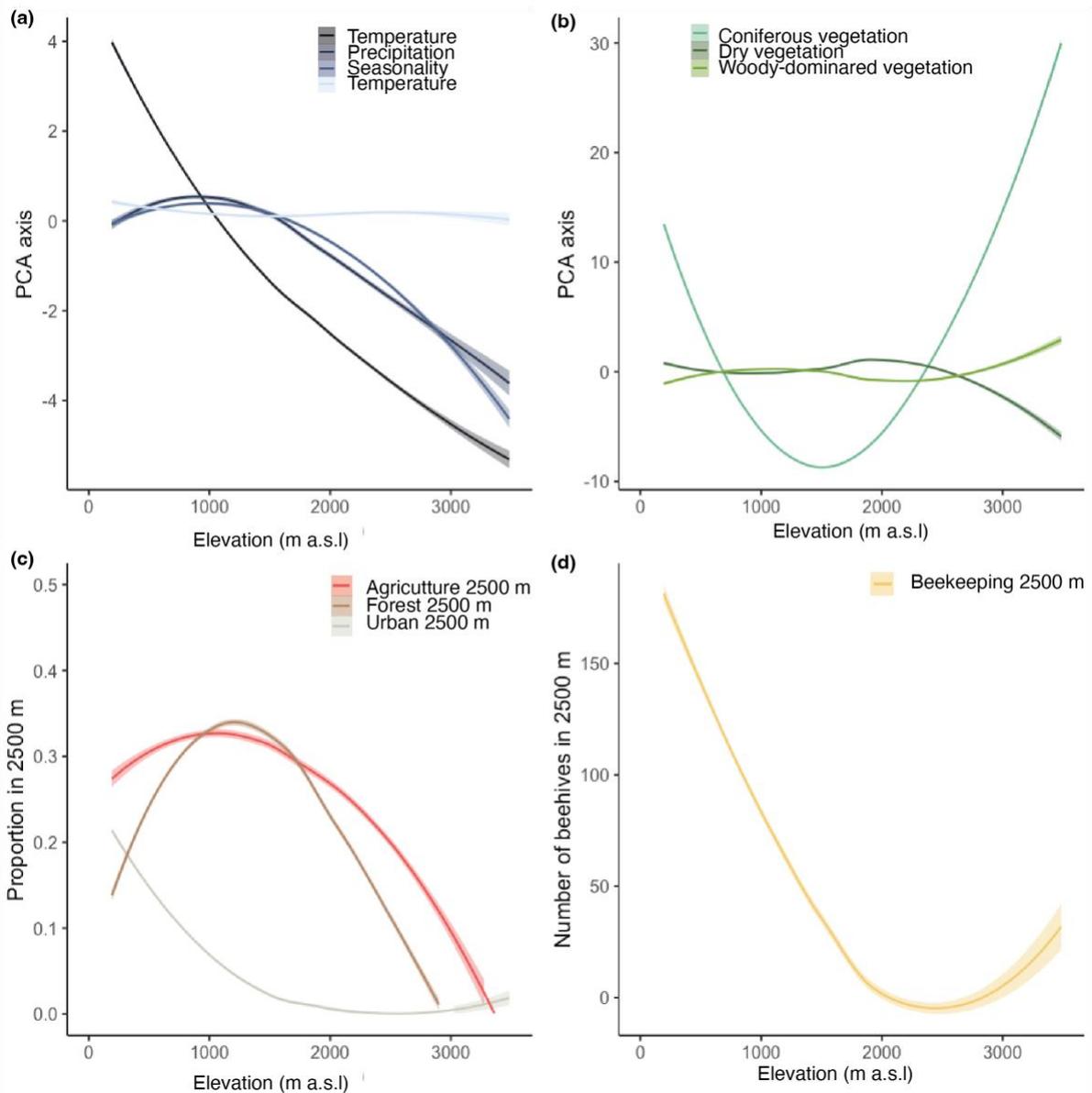
Appendix S15. Maps depicting the landscape metrics in the three land-use categories (agricultural, forest, urban). (a–d) Proportion of urban land use in 200, 500, 1000 and 2500 m radii around each raster cell centre. (e–h) Proportion of agricultural land use in 200, 500, 1000 and 2500 m radii. (i–l) Proportion of forest land use in 200, 500, 1000 and 2500 m radii.



Appendix S16. Correlation matrices between predictors. (a) Correlation matrix between all candidate predictors and (b) correlation matrix among the selected predictors. Red shading indicates positive correlations and blue indicates negative correlations.



Appendix S17. Maps depicting beekeeping intensity in Switzerland. Number of beehives in a (a) 200 m, (b) 500 m, (c) 1000 m, and (d) 2500 m radius around each raster cell centre.



Appendix S18. Relationship between the selected climate (four predictors, a), vegetation (three predictors, b), land use (three predictors, c), and beekeeping intensity (one predictor, d) predictors and elevation. Climate predictors represent principal components analysis (PCA) axes 1–4, which are related to temperature, precipitation, seasonality and precipitation seasonality (see details in Appendix S3). Vegetation predictors represent PCA axes 2–4, which are related to mid-elevation coniferous plant communities, dry plant communities, and forest plant communities (see details in Appendix S4 and Fig. S7). Land-use metrics represent the proportion of each land-use type (i.e. agricultural, forest, urban) within a 2500 m radius from each focal cell. Beekeeping intensity represents the number of beehives within a 2500 m radius from each focal cell. Note that the upper value of the elevation range has been narrowed to 3500 m a.s.l. for simplicity.

Appendix S19. Types of protected areas considered. For each protected area type, we indicate in what group of protected areas were included according to their protective measures (Classification), that is, strict protection = Protected areas *sensu stricto*; less strict protection = Protected areas *sensu lato*. We also indicate the main legislative authority, the equivalence with the IUCN categories, when possible, the proportion of the Swiss surface they represent, a description, some indication on the protection provisions, the existing law and reference to the official administrative bodies.

Protected area	Classification	Authority	IUCN category	Proportion surface	Description	Protection provisions	Law	Reference
Biotopes of National Interest: dry meadows	Strictly protected areas	Confederation	Not assessed officially, but likely IV	2.17%	Highly mencaed biotop types, which are highly monitored to preserve them. The aim of these inventories is to effectively protect the habitats of endangered animals and plants.	Management intensity and anthropogenic influence to the minimum. E.g.: No agricultural activities No hunting No leisure or other anthropogenic activities Continuous monitoring	451: Loi fédérale sur la protection de la nature et du paysage	FOEN 2012a, FOEN, 2022
Biotopes of National Interest: bogs								
Biotopes of National Interest: floodplains								
Biotopes of National Interest: fens								
Biotopes of National Interest: amphibian reproduction sites								
Ramsar		International	Not assessed officially, but likely IV	0.20%	Areas aimed at guaranteeing the sustainable use and protection of waterfowl habitats.	Specific management to promote biodiversity and matian habitats might occur.	451: Loi fédérale sur la protection de la nature et du paysage 0.451.45: Convention relative aux zones humides d'importance internationale particulièrement comme habitats des oiseaux d'eau	Ramsar, 2022
National Park		Confederation	II	0.41%	The only national park of the country, which has been assessed by the IUCN and classified within category II		454 Loi fédérale sur le Parc national suisse dans le canton des Grisons	Swiss National Park, 2022
Forest reserves		Confederation	Not assessed officially, but likely IV	2.10%	Forest reserves are among the core areas that form the basis of the ecological infrastructure in forests. In natural forest reserves, no silvicultural intervention is carried out and the forest is left to develop naturally. In special forest reserves, targeted interventions are aimed at promoting threatened species, which are mainly light and heat demanding species.		921.0: Loi fédérale sur les forêts	FOEN, 2020, 2021b
Pro Natura forest reserves		NGO		0.09%	Privately owned forest reserves. These forests are either untouched or used in traditional ways, with special measures in place to support biodiversity. In this manner, many habitats are preserved along with all their fauna and flora.		921.0: Loi fédérale sur les forêts	Pronatura, 2022
Pro Natura nature reserves		NGO		0.65%	Privately owned Natural reserves, which are either untouched or		Private law	Pronatura, 2022

Appendix S19. Continuation

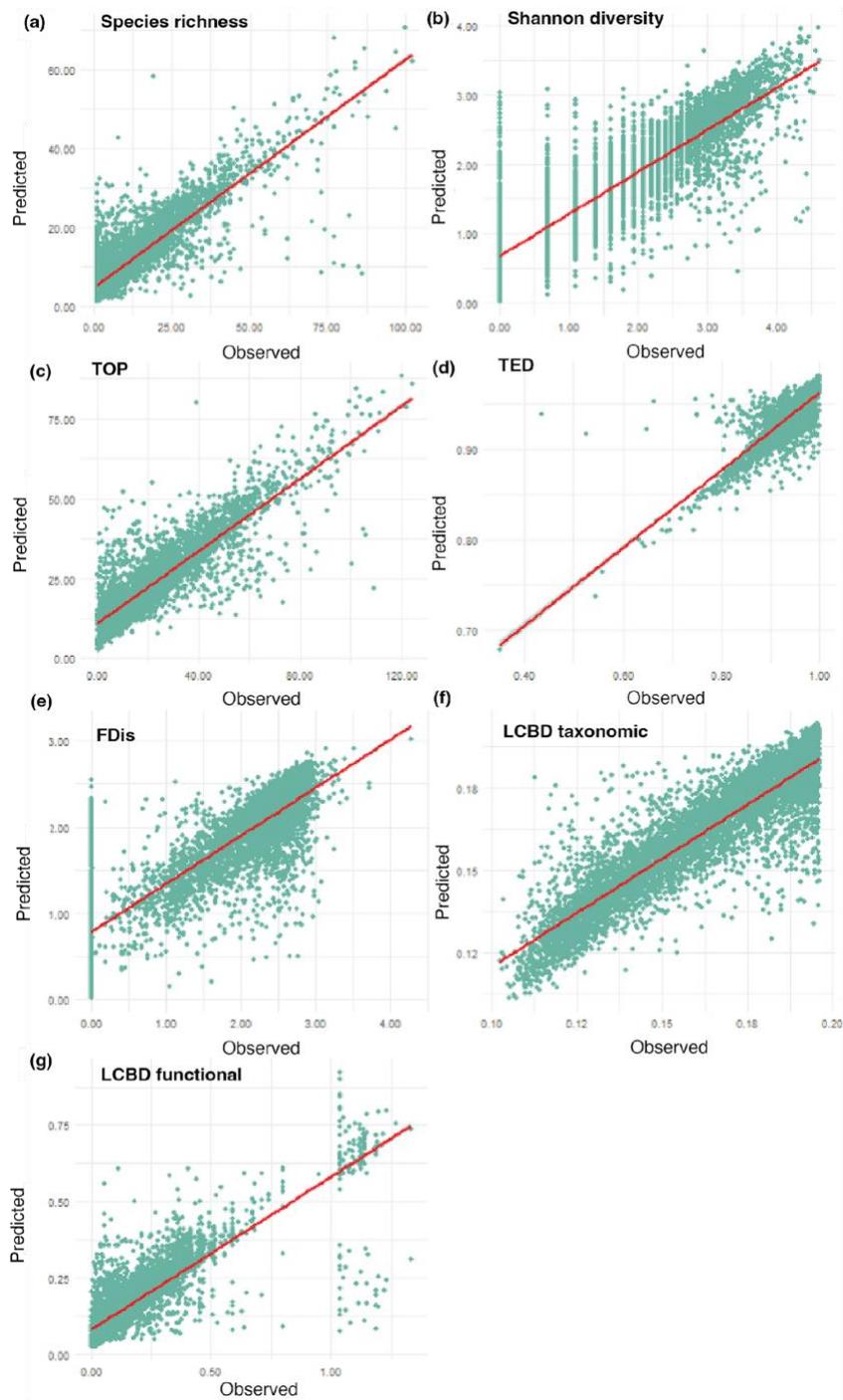
Protected area	Classification	Authority	IUCN category	Proportion surface	Description	Protection provisions	Law	Reference
Water and migrant bird reserves	Protected multiple use areas	Confederation / International	Not assessed officially, possible IV	4.84%	Areas aimed to protect the natural habitats of migratory birds that live in Switzerland throughout the year.	Variable management intensity and anthropogenic influence.	922.32: Ordonnance sur les réserves d'oiseaux d'eau et de migrateurs d'importance internationale et nationale	FOEN, 2002
Emerald sites		International	Not assessed	1.56%	The Emerald Network aims to protect species and natural environments of particularly high ecological value in Europe. Switzerland, as a Contracting Party to the Bern Convention, is also committed to this goal. So far, 37 areas proposed by Switzerland have been included in this international network.		0.455: Convention relative à la conservation de la vie sauvage et du milieu naturel de l'Europe	FOEN, 2003, 2012b
Swiss parks		Cantonal, municipal	Not assessed, some could be V or VI	12.28%	Regional nature parks are partly inhabited rural areas with a high level of natural, landscape and cultural wealth. They enhance the quality of nature and landscape and promote the sustainable development of the regional economy. Peri-urban nature parks are areas in the vicinity of a highly urbanised region which offer intact natural habitats for indigenous flora and fauna in their core area. The core area is surrounded by a transition zone that serves as a buffer and offers a variety of opportunities for education, discovery and recreation, thus making an important contribution to the quality of life of the urban population.		451: Loi fédérale sur la protection de la nature et du paysage	FOEN, 2018a
UNESCO Natural Heritage		International	Not assessed officially, possible III, V, VI	2.80%	The World Heritage List is an instrument of the Convention concerning the Protection of the World Cultural and Natural Heritage (World Heritage Convention). It lists properties classified by the World Heritage Committee for their outstanding universal value.		International law RO 1975 2221 Arrêté fédéral du 19 juin 1975 approuvant deux conventions de l'UNESCO en matière de protection du patrimoine culturel et naturel et de conservation des zones humides	FOEN, 2016
Game reserves		Confederation	Not assessed officially, some could be V	3.63%	In accordance with the provisions of the Ordinance on Federal Game Reserves, game reserves "are intended to protect and conserve rare and endangered wild mammals and birds and to protect and conserve their biotopes. They are also intended to maintain healthy populations of huntable species adapted to local conditions		922.0: Loi fédérale sur la chasse et la protection des mammifères et oiseaux sauvages	FOEN, 2018b
UNESCO Biosphere		International	Not assessed officially, possible III, V, VI	2.94%	The United Nations Educational, Scientific and Cultural Organisation (UNESCO) has set up the Man and Biosphere Programme (MAB) with the aim of developing new models for the sound management of natural environments. The MAB programme is implemented in UNESCO biosphere reserves. It applies the principles of sustainable and environmentally friendly development.		International law RO 1975 2221 Arrêté fédéral du 19 juin 1975 approuvant deux conventions de l'UNESCO en matière de protection du patrimoine culturel et naturel et de conservation des zones humides	UNESCO, 2022

Appendix S20. Performance of the different algorithms used to predict the α and β taxonomic and functional attributes of wild bee communities in Switzerland. Three performance metrics were calculated on the test dataset (i.e. data not used for model calibration): root mean squared error (RMSE), R^2 and mean absolute error (MAE).

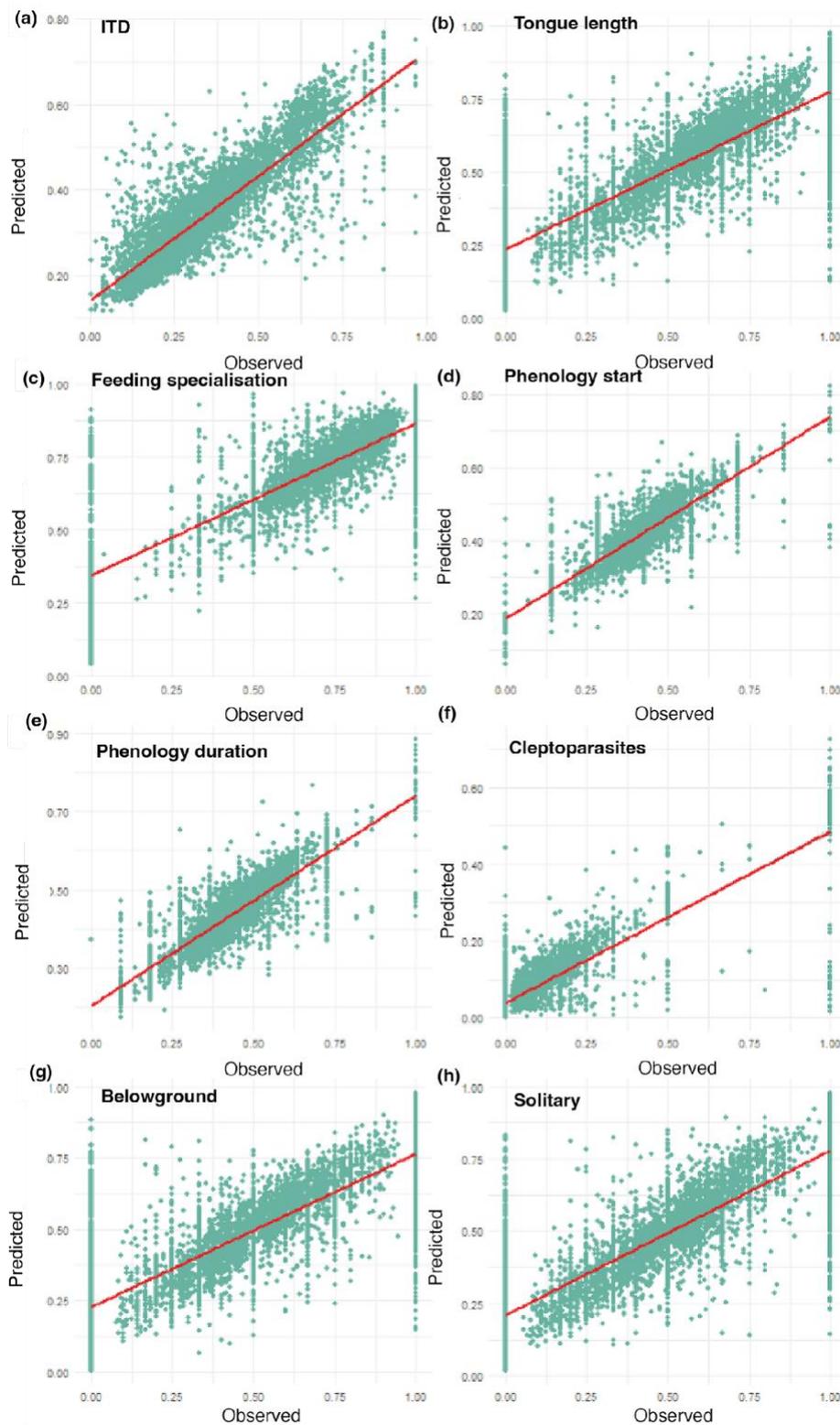
RMSE	R^2	MAE	Response	Model
5.03	0.91	3.36	Richness	Random forest
15.89	NA	9.59	Richness	Neural networks
12.40	0.04	8.83	Richness	Generalized linear models
0.45	0.92	0.37	Shannon	Random forest
1.40	0.02	1.18	Shannon	Neural networks
1.19	0.04	1.02	Shannon	Generalized linear models
0.43	0.91	0.35	FDis	Random forest
1.30	0.02	1.21	FDis	Neural networks
1.04	0.02	0.90	FDis	Generalized linear models
7.94	0.91	5.97	TOP	Random forest
30.62	NA	23.54	TOP	Neural networks
19.09	0.05	14.76	TOP	Generalized linear models
0.02	0.90	0.01	TED	Random forest
0.04	0.01	0.03	TED	Neural networks

0.04	0.01	0.03	TED	Generalised linear models
0.07	0.91	0.05	ITD	Random forest
0.16	0.14	0.12	ITD	Neural networks
0.16	0.11	0.12	ITD	Generalised linear models
0.12	0.91	0.09	Feeding specialisation	Random forest
0.28	0.05	0.21	Feeding specialisation	Neural networks
0.28	0.03	0.21	Feeding specialisation	Generalised linear models
0.14	0.91	0.11	Tongue length	Random forest
0.32	0.11	0.26	Tongue length	Neural networks
0.32	0.08	0.26	Tongue length	Generalised linear models
0.05	0.90	0.03	Phenology start	Random forest
0.11	0.12	0.08	Phenology start	Neural networks
0.11	0.09	0.08	Phenology start	Generalised linear models
0.05	0.90	0.04	Phenology duration	Random forest
0.11	0.12	0.08	Phenology duration	Neural networks
0.12	0.09	0.08	Phenology duration	Generalised linear models
0.07	0.90	0.04	Cleptoparasite	Random forest
0.15	0.03	0.09	Cleptoparasite	Neural networks

0.15	0.02	0.09	Cleptoparasite	Generalised linear models
0.14	0.91	0.11	Belowground	Random forest
0.32	0.08	0.25	Belowground	Neural networks
0.32	0.08	0.25	Belowground	Generalised linear models
0.13	0.91	0.10	Solitary	Random forest
0.30	0.16	0.24	Solitary	Neural networks
0.31	0.12	0.25	Solitary	Generalised linear models
0.00	0.92	0.00	LCBD taxonomic	Random forest
0.00	NA	0.00	LCBD taxonomic	Neural networks
0.00	0.14	0.00	LCBD taxonomic	Generalised linear models
0.00	0.90	0.00	LCBD functional	Random forest
0.00	NA	0.00	LCBD functional	Neural networks
0.00	0.01	0.00	LCBD functional	Generalised linear models

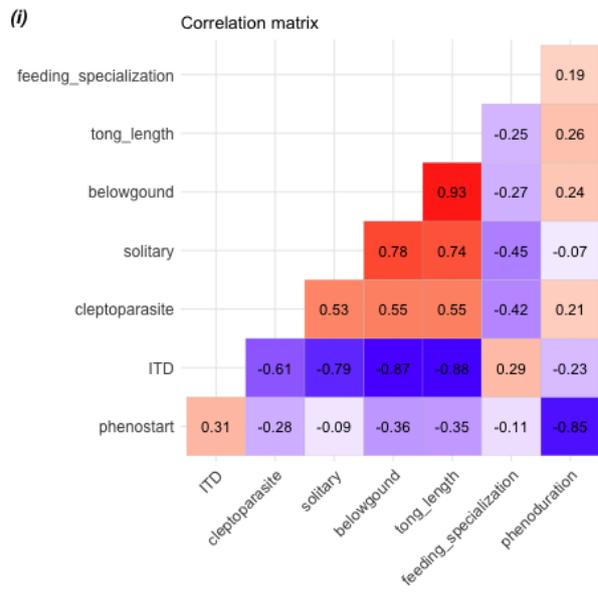
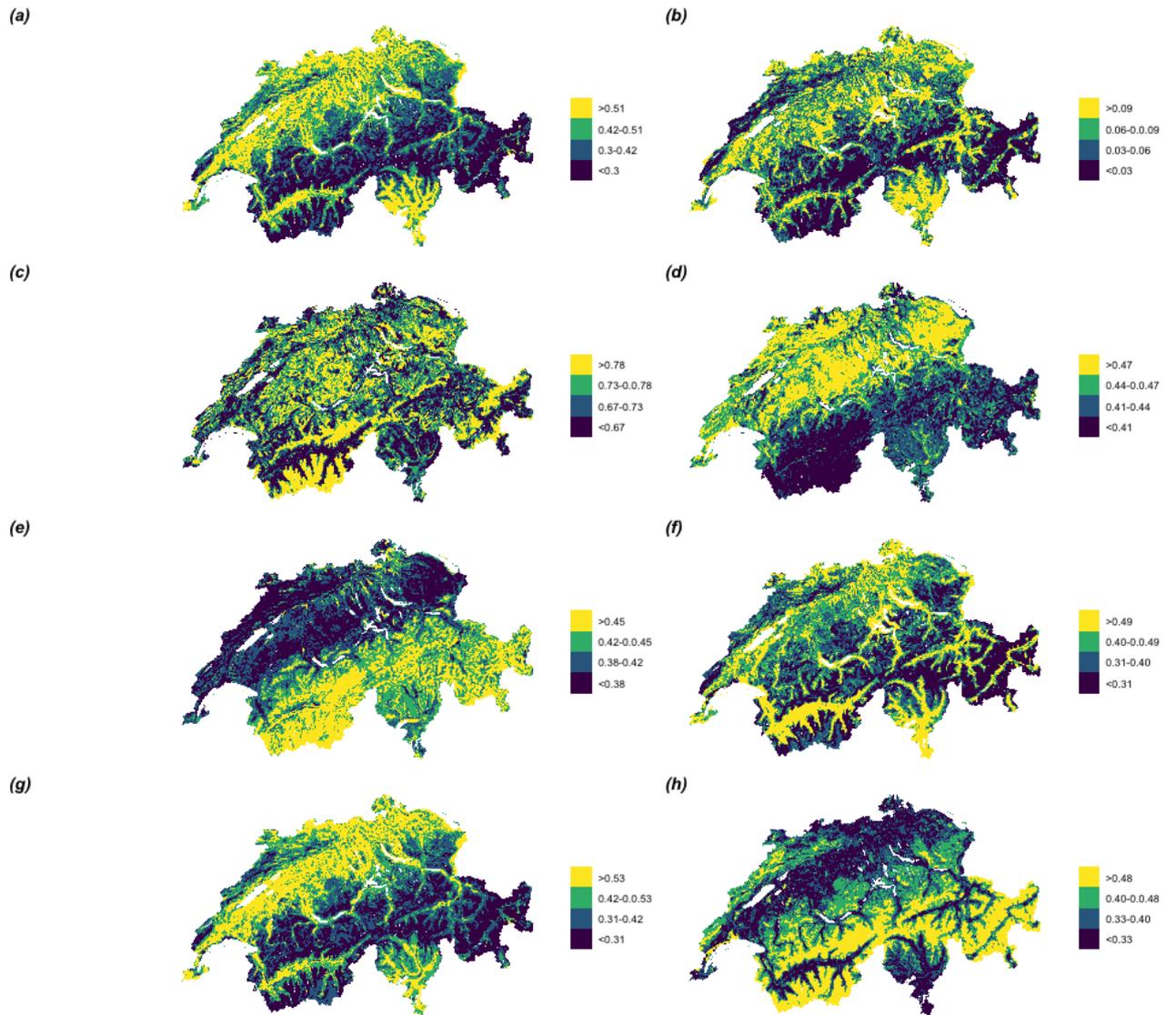


Appendix S21. Scatterplots of the predictive performance (observed vs. predicted values) of random forest models for diversity metrics. (a) species richness , (b) Shannon diversity, (c) trait onion peeling (TOP), (d) trait evenness distribution (TED), (e) functional dispersion (FDis), (f) local community contributions to the taxonomic β -diversity (LCBD taxonomic), and (g) local community contributions to the functional β -diversity (LCBD functional).

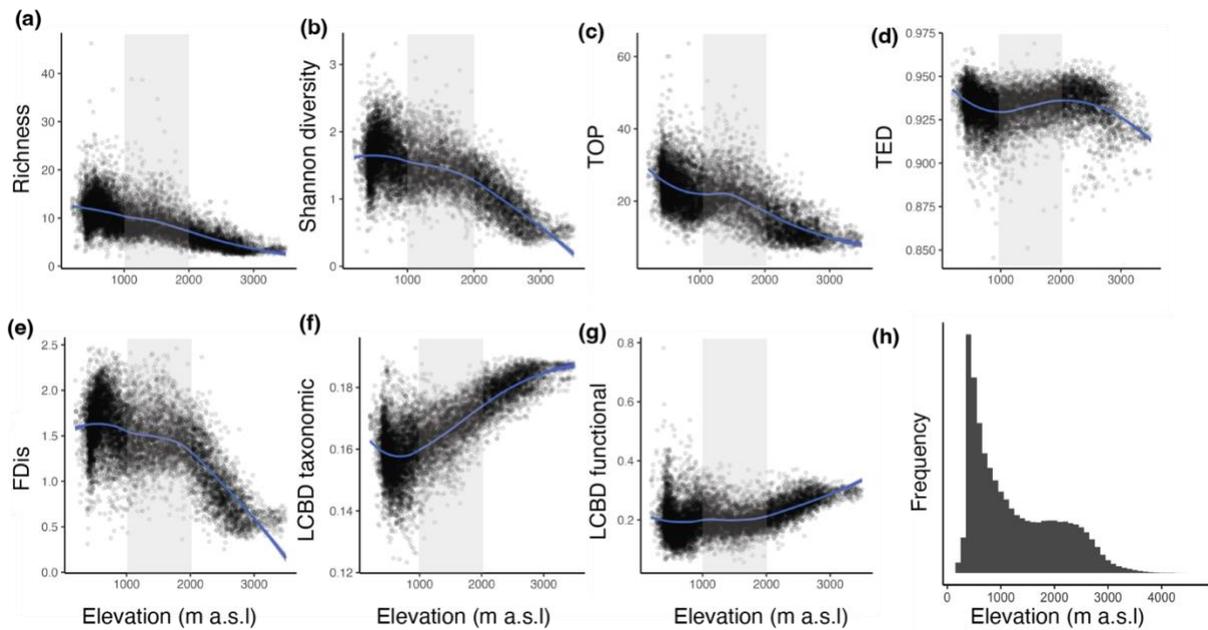


Appendix S22. Scatterplots of the predictive performance (observed vs. predicted values) of random forest models for community-weighted means of the eight studied wild bee traits. (a) intertegular

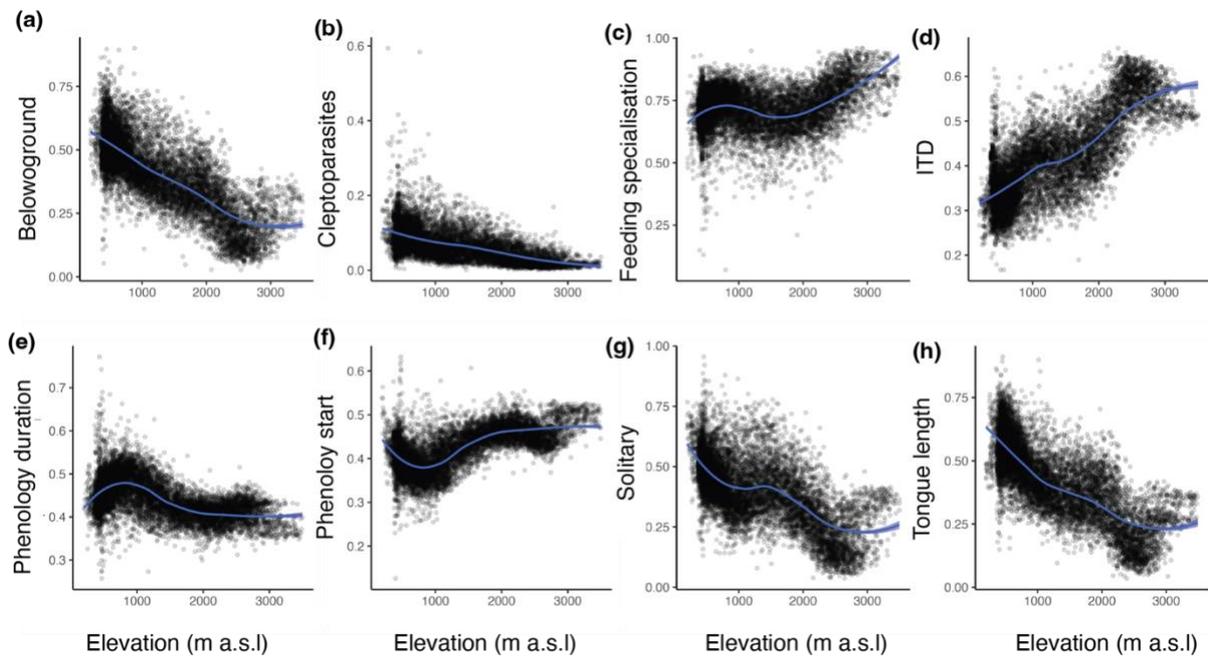
distance (ITD), (b) tongue length, (c) feeding specialisation, (d) start of phenology, (e) duration of phenology, (f) % cleptoparasite, (g) % belowground nester, and (h) % solitary.



Appendix S23. Wild bee trait distribution in Switzerland. Maps depict the community-weighted means (CWMs) of (a) proportion of belowground species, (b) proportion of cleptoparasitic species (cleptoparasite), (c) feeding specialisation (feeding_specialisation), (d) duration of phenology (phenoduration), (e) proportion of solitary species (solitary), (f) tongue length (tong_length), and (g) intertegular distance (ITD). (h) Pearson correlations between the CWMs of the seven studied traits.

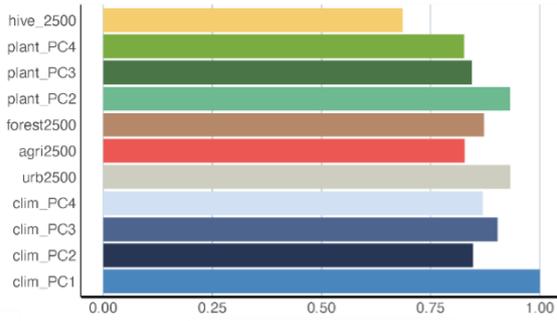


Appendix S24. Relationship between the predicted α and β taxonomic diversity and elevation and between functional community attributes and elevation based on 10000 randomly selected cells. Fitted lines and the 95% confidence intervals (shaded bands) were obtained using local polynomial regression (LOESS). Relationships are shown between elevation and wild bee (a) species richness (rich), (b) Shannon diversity (shannon), (c) trait onion peeling (TOP), (d) trait evenness distribution (TED), (e) functional dispersion (FDis), (f) local community contributions to the taxonomic β -diversity (LCBD_taxo), and (g) local community contributions to the functional β -diversity (LCBD_fun). (h) Histogram of the digital elevation model for Switzerland. Note that the upper value of the elevation range has been narrowed to 3500 m a.s.l. for simplicity.

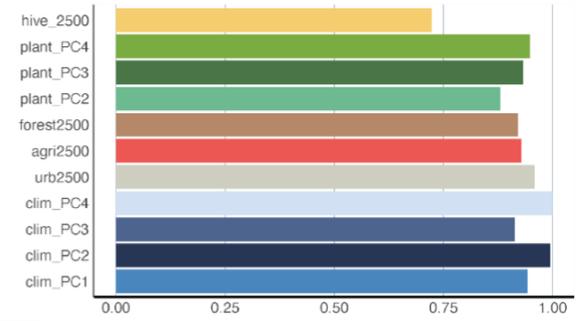


Appendix S25. Relationship between the community-weighted means (CWMs) of the eight studied traits and elevation based on 10000 randomly selected cells. Fitted lines and the 95% confidence intervals (shaded bands) were obtained from local polynomial regression (LOESS). Relationships are shown between elevation and (a) proportion of belowground species (belowground; high values = more belowground nesters, low values = fewer belowground nesters), (b) proportion of cleptoparasitic species (cleptoparasite; high values = more cleptoparasites, low values = less cleptoparasites), (c) feeding specialisation (feeding_specialisation; high values = less specialisation, low values = more specialisation), (d) intertegular distance (ITD), (e) duration of the phenology (phenoduration), (f) start of phenology (phenostart); (g) proportion of solitary species (solitary; high values = more solitary species, low values = fewer solitary species), and (h) tongue length (tongue_length; high values = longer tongues, low values = shorter tongues).

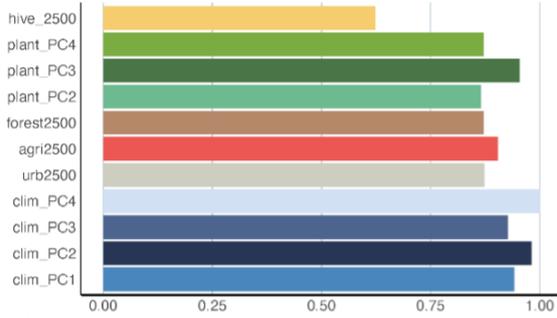
(a) Proportion belowground



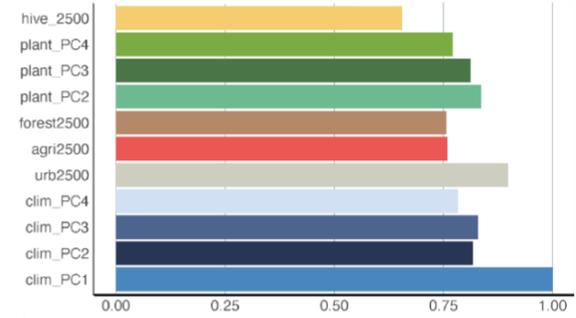
(b) Proportion cleptoparasite



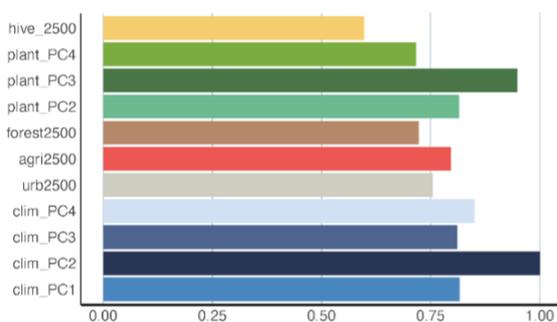
(c) Feeding specialisation



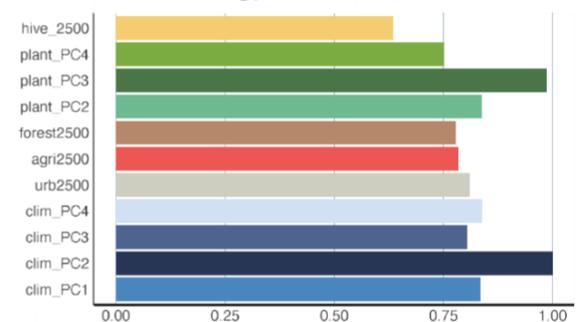
(d) Intertegular distance



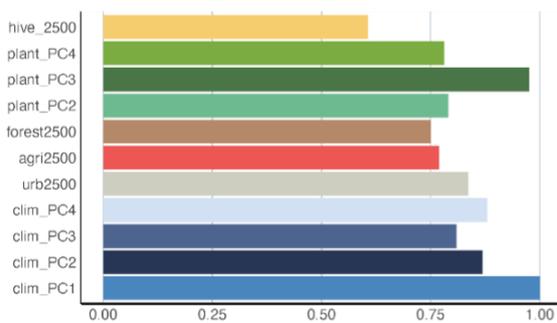
(e) Phenology duration



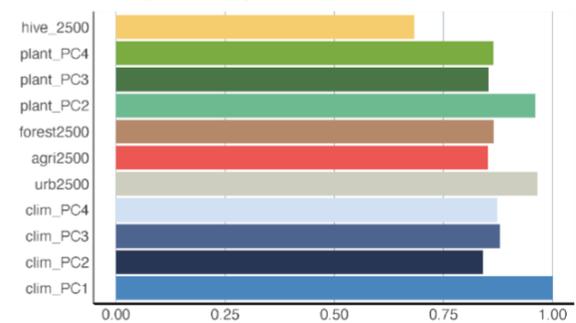
(f) Phenology start



(g) Proportion solitary

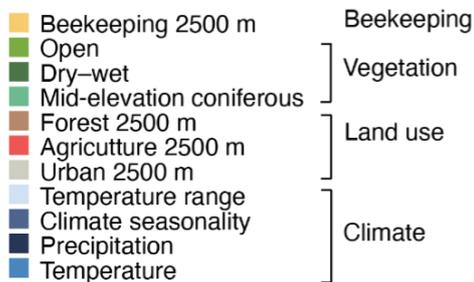


(h) Tongue length

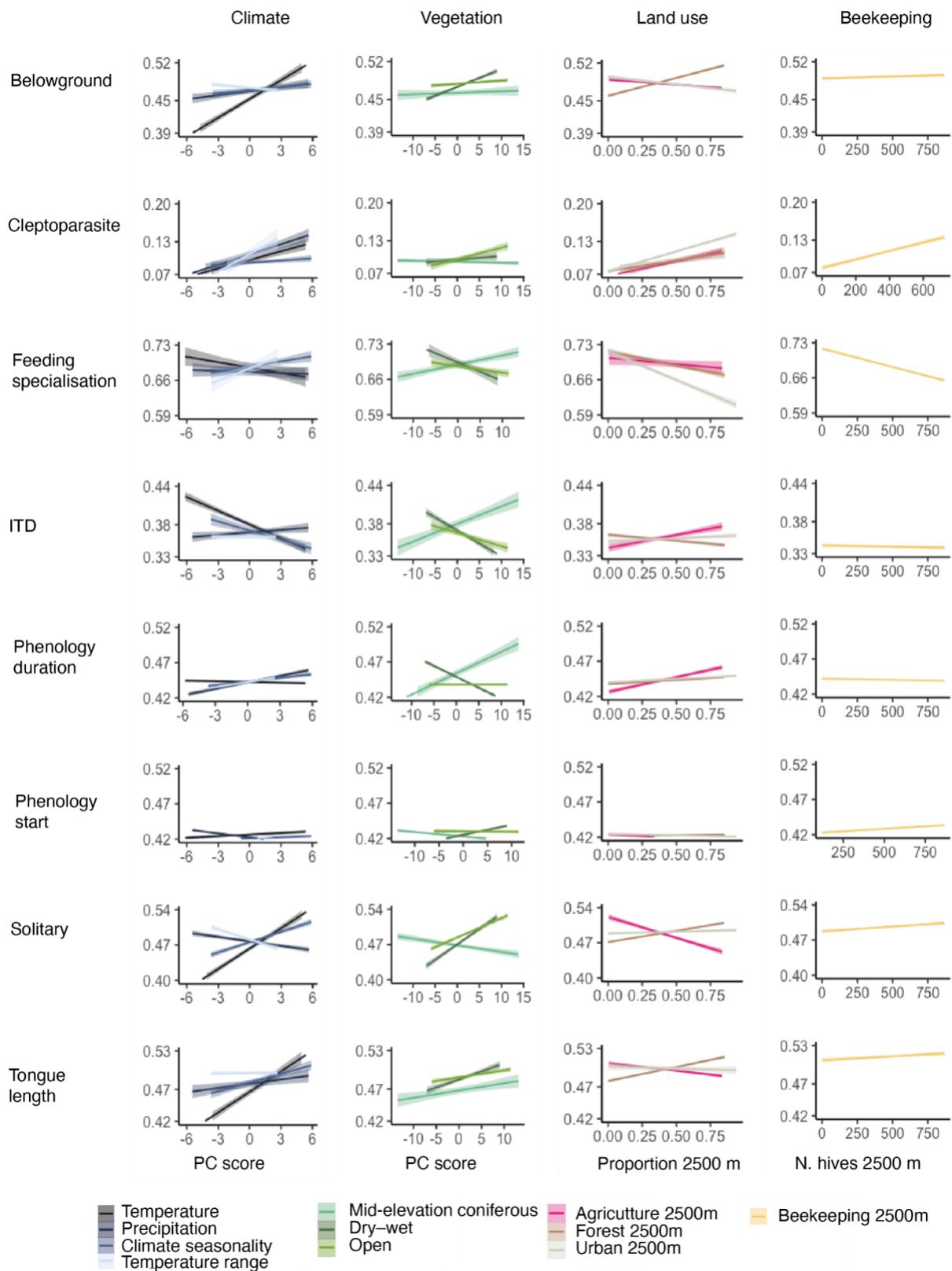


Variable importance

Variable importance

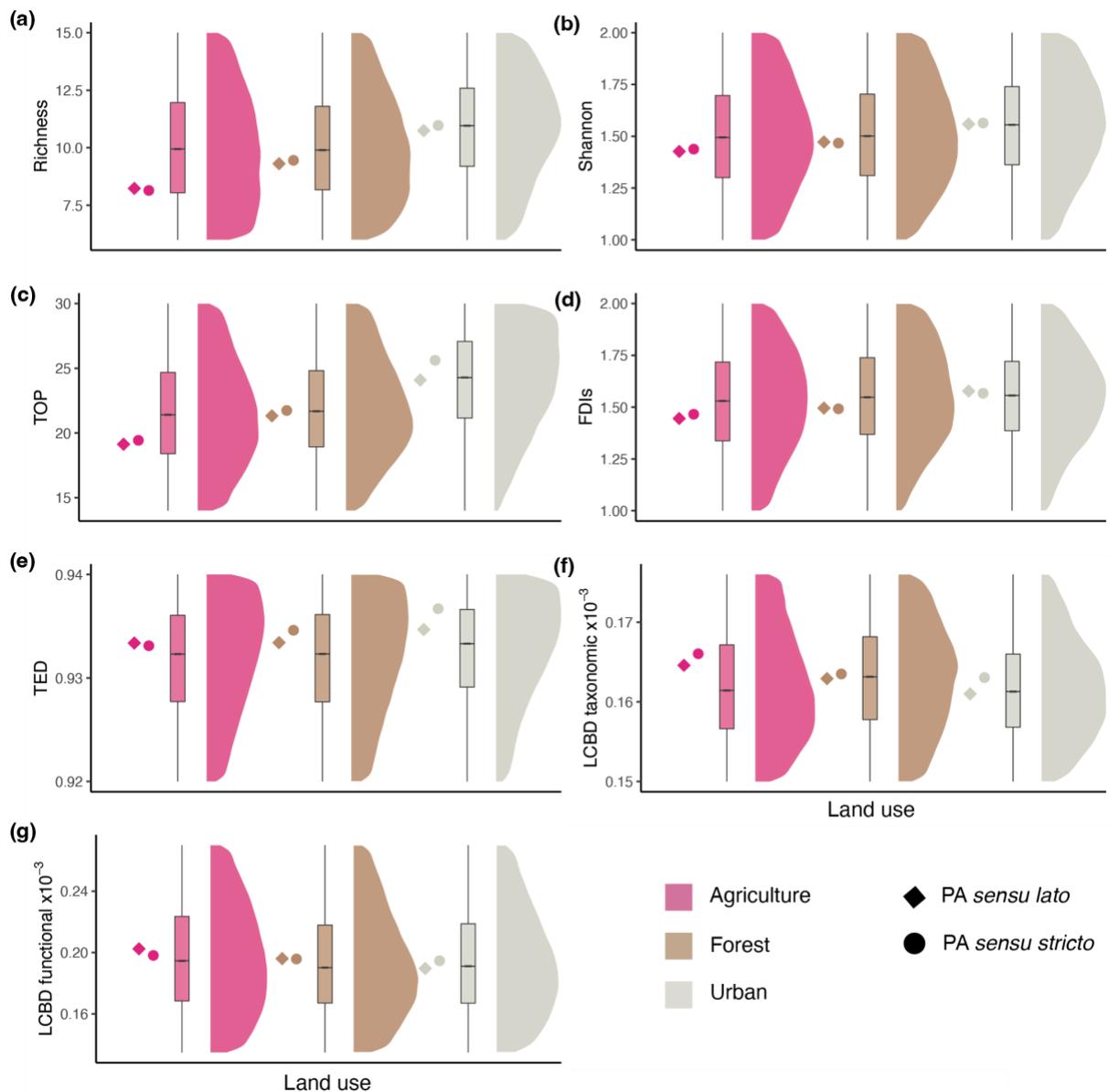


Appendix S26. Importance of the environmental variables as predictors of the community-weighted means of the eight studied wild bee traits in Switzerland. Predictors were classified into four main categories: climate, vegetation, land use and beekeeping. Variable importance was estimated using the residual sum of squares from random forest models (Breiman, 2001). Longer bars indicate variables that are better predictors of community attributes. Note that all importance values were divided by the maximum value to obtain a comparable range from 0 to 1. Climate and vegetation variables represent the principal components analysis (PCA) axis (PCA 1–4 for climate, PCA 2–4 for vegetation, representing 17% of the variation; for details see Methods, Tables S3–S4 and Figs S4–S5).

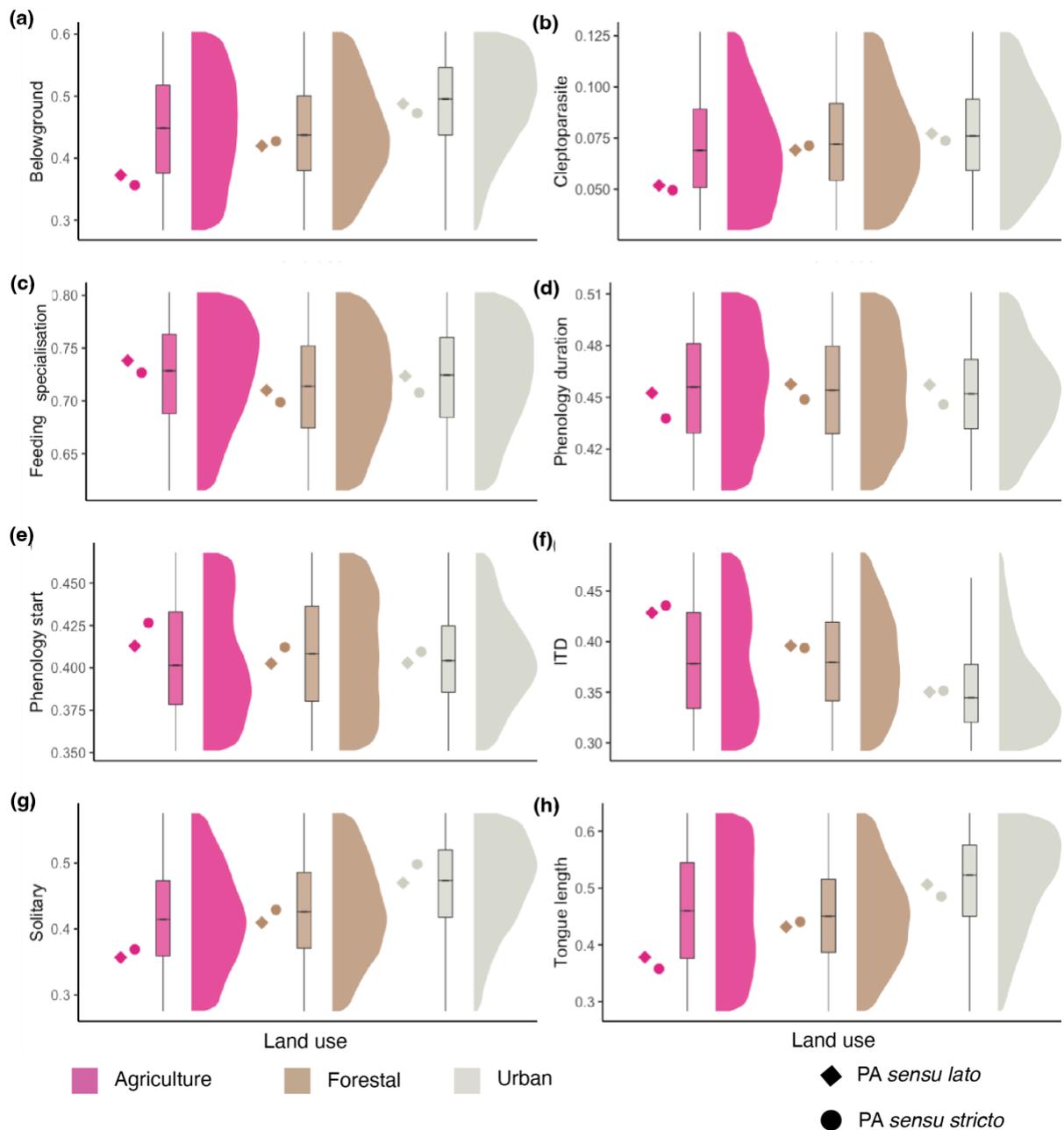


Appendix S27. Predicted changes in the community-weighted means (CWMs) of the eight studied wild bee traits along multiple environmental gradients. Partial dependence plots showing the shifts induced

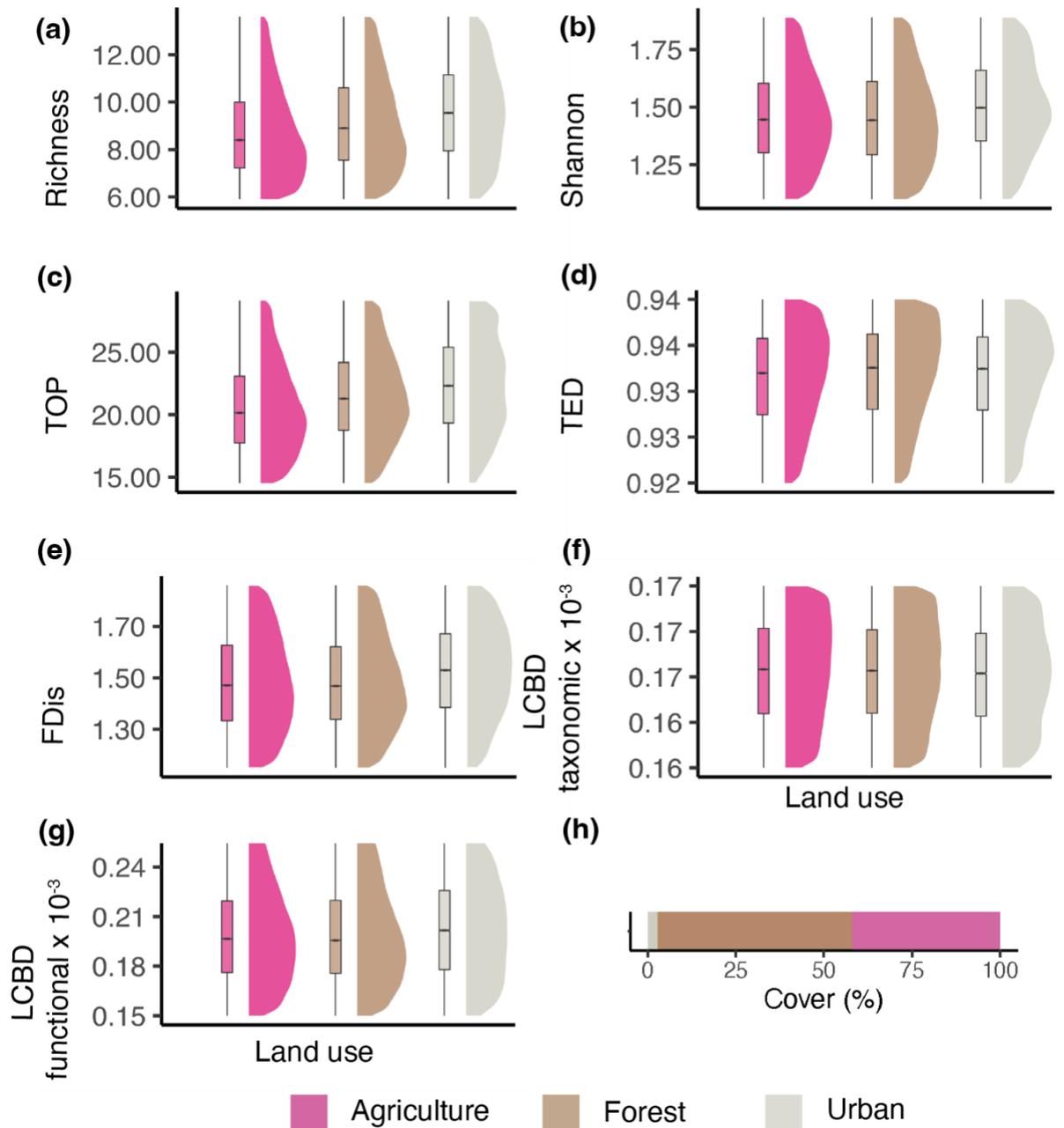
by selected variables representing climate, vegetation, land-use and beekeeping gradients, indicated by the CWM of the proportion of belowground species (belowground), proportion of cleptoparasitic species (cleptoparasite), feeding specialisation, intertegular distance (ITD), start of phenology (phenology start), duration of phenology (phenology duration), proportion of solitary species (solitary), and tongue length. Shaded bands indicate 95% confidence intervals. Climate and vegetation variables represent the principal components analysis (PCA) axis (PCA 1–4 for climate, PCA 2–4 for vegetation). For details see Methods, Tables S3–S4 and Figs S4–S5.



Appendix S28. Flat violin plots and boxplots showing the differences in the α and β taxonomic diversity and functional diversity metrics among the three main land-use types agricultural, urban and forest. Notches in the boxplots indicate the 95% confidence interval of the median. Dots and diamonds indicate the median value of the metric in protected areas (PA) *sensu stricto* and *sensu lato*, respectively. (a) Species richness, (b) Shannon diversity, (c) functional richness (trait onion peeling TOP), (d) functional evenness (trait evenness distribution TED), (e) functional dispersion (FDis), (f) local community contributions to taxonomic β -diversity (LCBD taxonomic), and (g) local community contributions to functional β -diversity (LCBD functional).

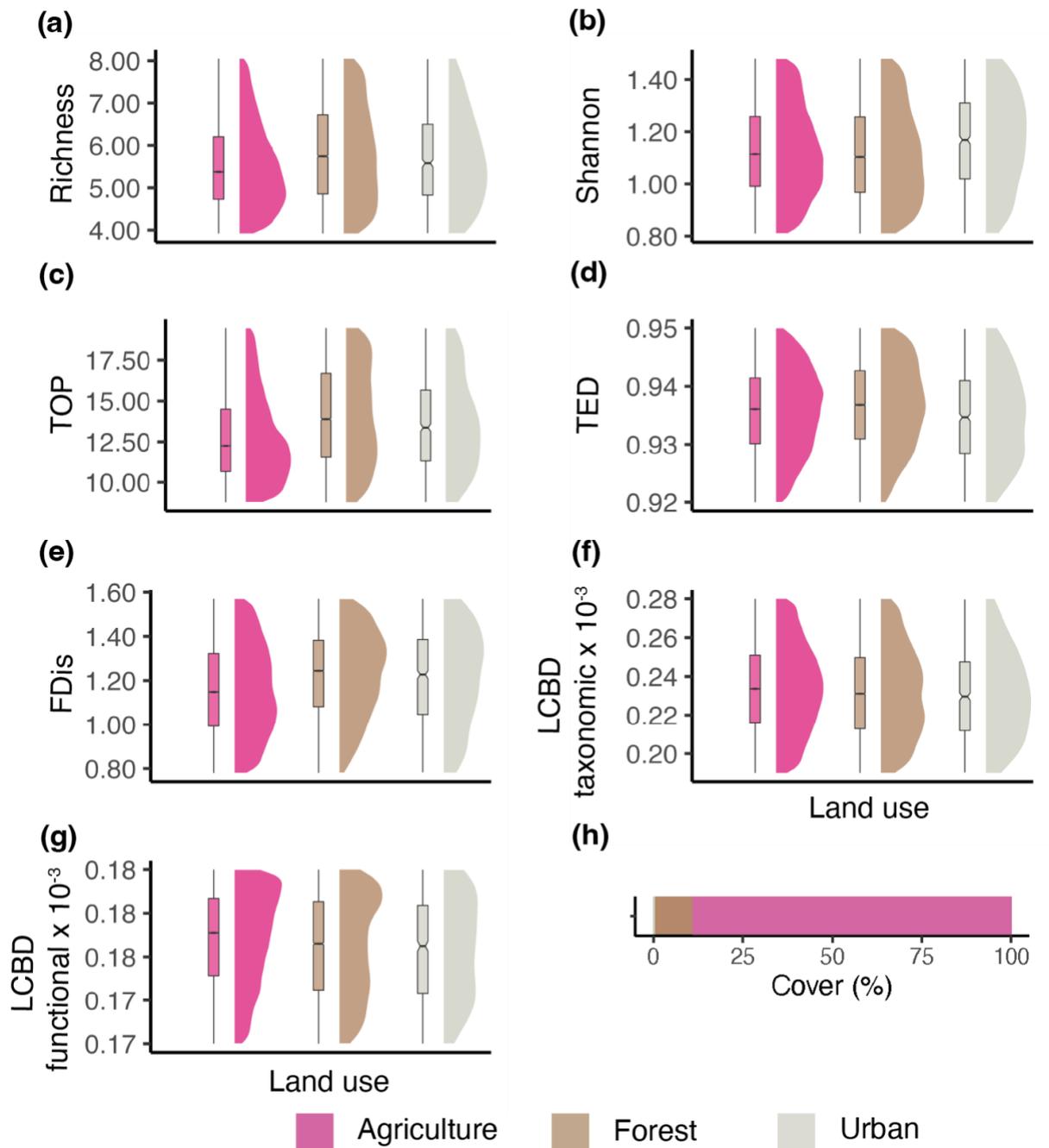


Appendix S29. Flat violin plots and boxplots showing the differences in the community-weighted means of the eight studied wild bee traits among the three main land-use types agricultural, urban and forest. Notches in the boxplots indicate the 95% confidence interval of the median. Dots and diamonds indicate the median value of the metric in protected areas (PA) *sensu stricto* and *sensu lato*, respectively. (a) Belowground nesters, (b) cleptoparasites, (c) feeding specialisation, (d) duration of phenology, (e) start of phenology, (f) intertegular distance (ITD), (g) solitary, and (h) tongue length. Note that trait values have been standardised.



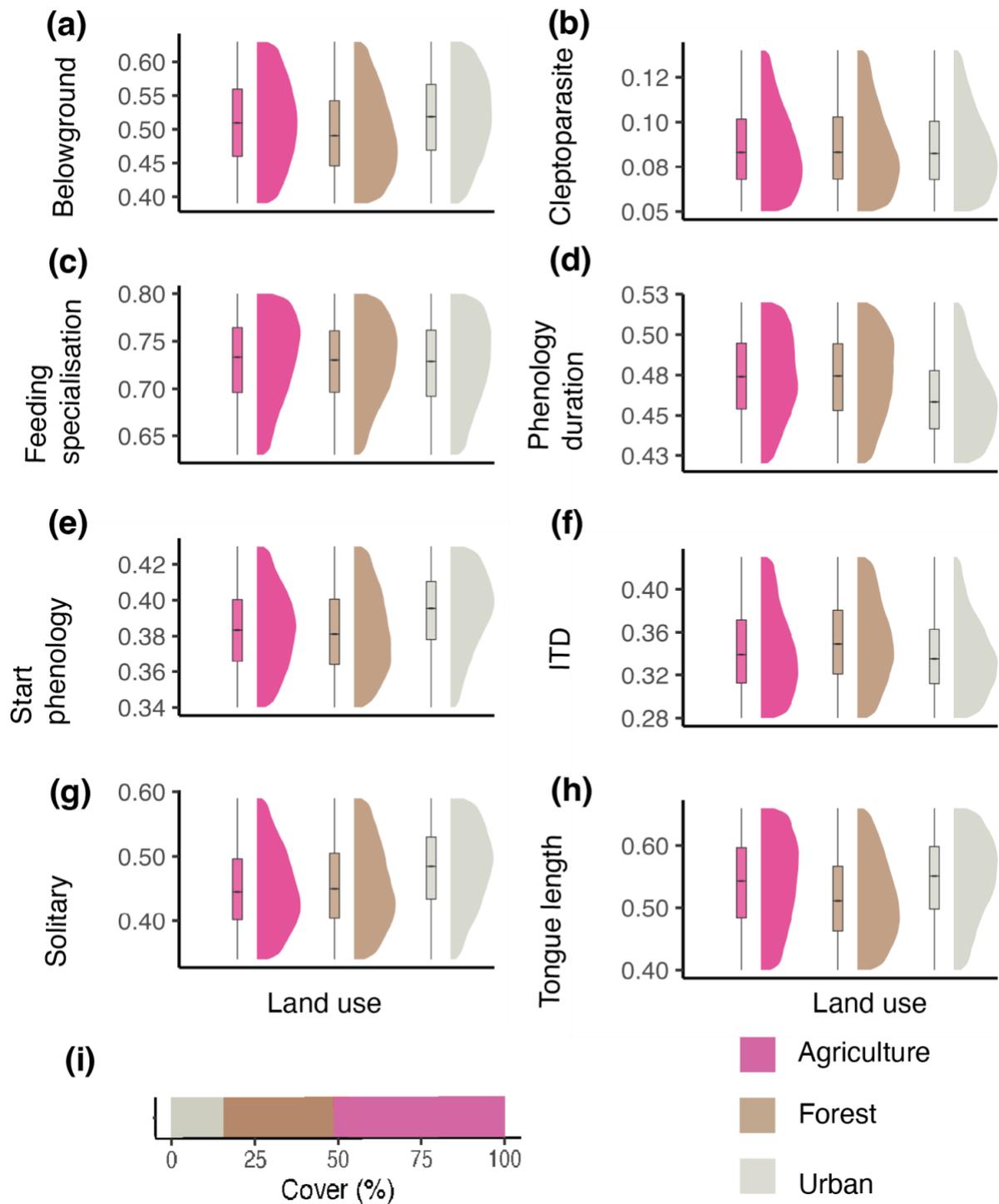
Appendix S30. Flat violin plots and boxplots showing the differences in the α and β taxonomic and functional diversity metrics among the three main land-use types, agricultural, urban and forest, at mid elevation (1000–2000 m a.s.l.). Notches in the boxplots indicate the 95% confidence interval of the median. (a) Species richness, (b) Shannon diversity, (c) functional richness (trait onion peeling TOP), (d) functional evenness (trait evenness distribution TED), (e) functional dispersion (FDis), (f) local community contributions to taxonomic β -diversity (LCBD taxonomic), and (g) local community contributions to functional β -diversity (LCBD functional). Note that to facilitate the comparison of

the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles. Note that to facilitate the comparison of the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles. (h) Land-use composition (% cover) at mid elevations.



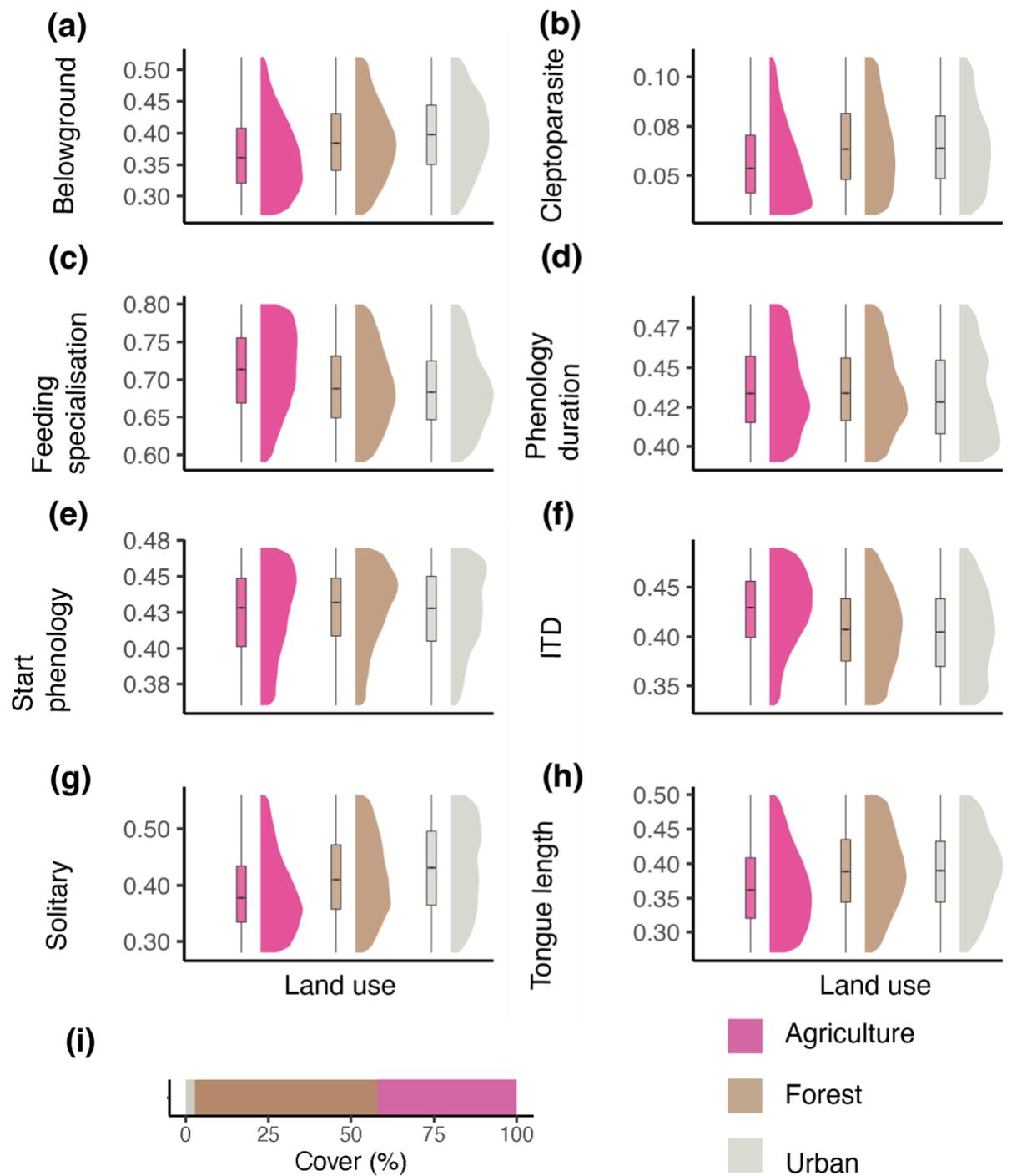
Appendix S31. Flat violin plots and boxplots showing the differences in the α and β taxonomic and functional metrics among the three main land-use types, agricultural, urban and forest, at high elevation (> 2000 m a.s.l.). Notches in the boxplots indicate the 95% confidence interval of the median. (a) Species richness, (b) Shannon diversity, (c) functional richness (trait onion peeling TOP), (d) functional evenness (trait evenness distribution TED), (e) functional dispersion (FDis), (f) local community contributions to taxonomic β -diversity (LCBD taxonomic), and (g) local community contributions to

functional β -diversity (LCBD functional). (h) Land-use composition (% cover) at high elevations. Note that to facilitate the comparison of the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles. Furthermore, urban land-use at high elevation refers mostly to sparse infrastructure (e.g. small roads, dams, avalanche protection areas, small resorts, sport surfaces) rather than densely build-up surfaces.



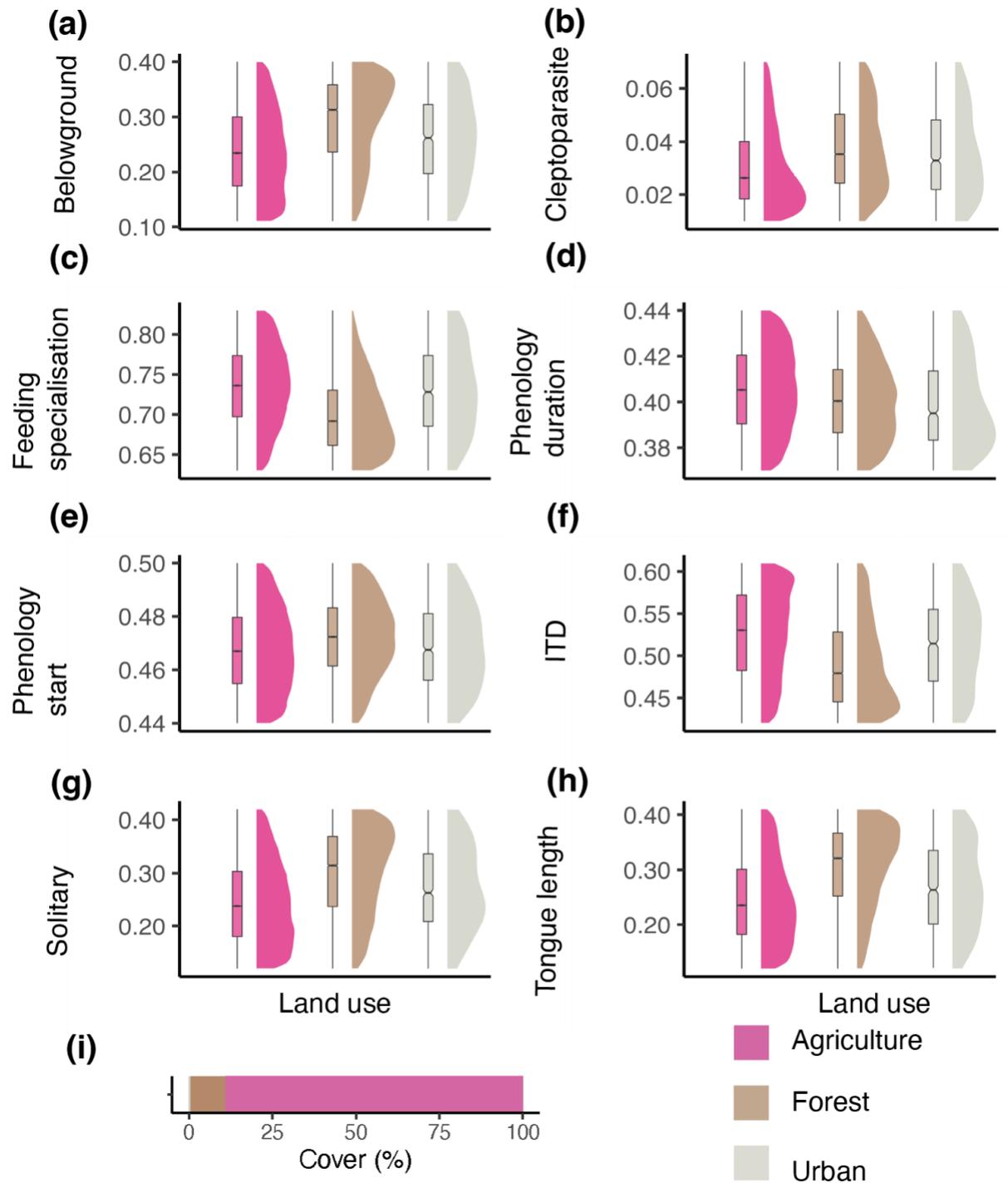
Appendix S32. Flat violin plots and boxplots showing the differences in the community-weighted means of the eight studied wild bee traits among the three main land-use types, agricultural, urban and forest, at low elevation (197–1000 m a.s.l.). Notches in the boxplots indicate the 95% confidence interval of the median. (a) Belowground nesters, (b) cleptoparasites, (c) feeding specialisation, (d)

duration of phenology, (e) start of phenology, (f) intertegrular distance (ITD), (g) solitary, and (h) tongue length. (i) Land-use composition (% cover) at low elevations. Note that trait values have been standardised. Note that to facilitate the comparison of the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles.



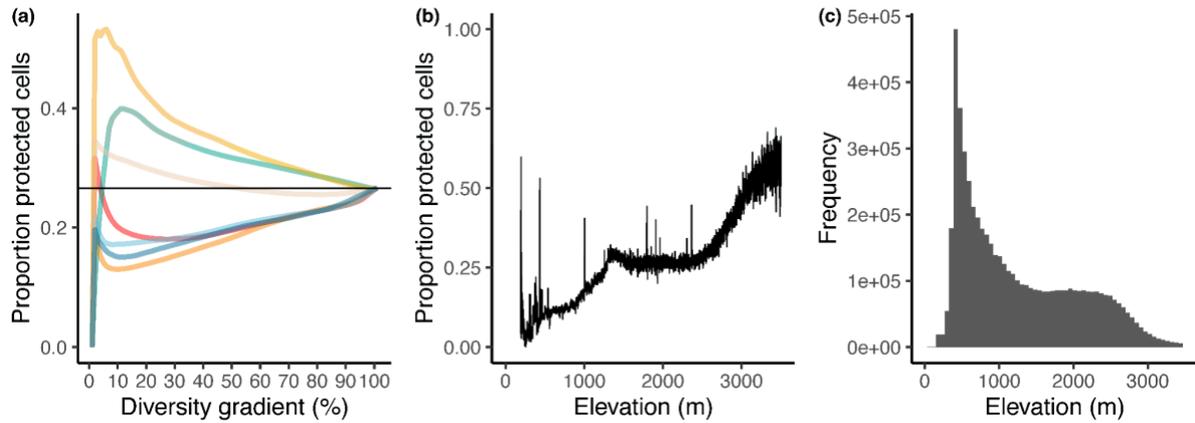
Appendix S33. Flat violin plots and boxplots showing the differences in the community-weighted means of the eight studied wild bee traits among the three main land-use types, agricultural, urban and forest, mid elevation (1000–2000 m a.s.l.). Notches in the boxplots indicate the 95% confidence interval of the median. (a) Belowground nesters, (b) cleptoparasites, (c) feeding specialisation, (d) duration of phenology, (e) start of phenology, (f) intertegular distance (ITD), (g) solitary, and (h) tongue length. (i)

Land-use composition (% cover) at mid elevations. Note that trait values have been standardised. Note that to facilitate the comparison of the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles.

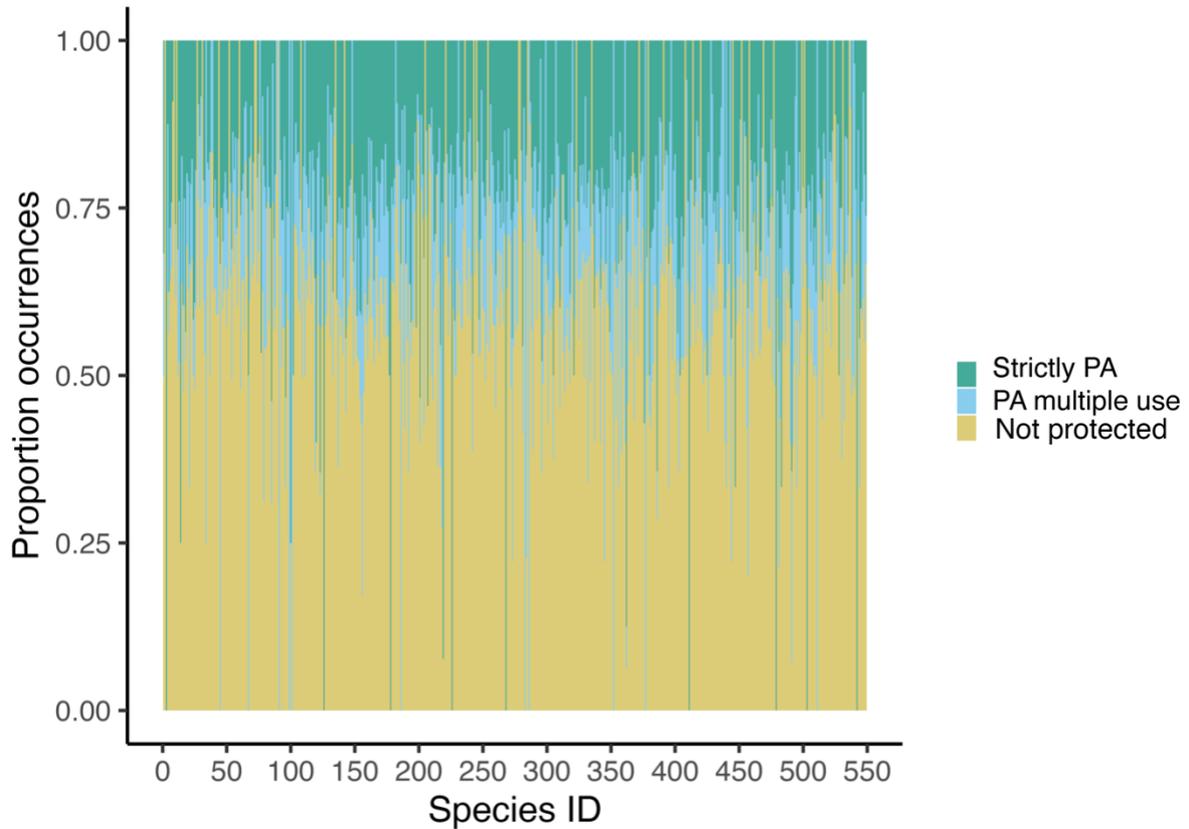


Appendix S34. Flat violin plots and boxplots showing the differences in the community-weighted means of the eight studied wild bee traits among the three main land-use types, agricultural, urban and

forest, at high elevation (> 2000 m a.s.l.). Notches in the boxplots indicate the 95% confidence interval of the median. (a) Belowground nesters, (b) cleptoparasites, (c) feeding specialisation, (d) duration of phenology, (e) start of phenology, (f) intertegular distance (ITD), (g) solitary, and (h) tongue length. (i) Land-use composition (% cover) at high elevations. Note that trait values have been standardised. Note that to facilitate the comparison of the boxplots, the Appendix Shows the data between the 10th and the 90th percentiles. Furthermore, urban land-use at high elevation refers mostly to sparse infrastructure (e.g. small roads, dams, avalanche protection areas, small resorts, sport surfaces) rather than densely build-up surfaces.



Appendix S35. Proportion of taxonomic and functional diversities included in all the protected areas (protected areas *sensu lato* + protected areas *sensu stricto*). For each diversity metric, we ranked each cell from the most to the least diverse using quantile values. The x-axis (diversity gradient in percentage) depicts a decreasing diversity gradient inversely related to the quantiles of the diversity metrics, with lower x-axis values indicating the most diverse cells. The y-axis depicts the cumulative proportion of cells belonging to protected areas. The horizontal black line indicates the proportion of protected cells in all of Switzerland (ca. 27 % of the surface). (a) is based on Devictor et al. (2010). Taxonomic α -diversity metrics = species richness and Shannon diversity; functional α -diversity metrics = functional richness (trait onion peeling TOP), functional evenness (trait evenness distribution TED), and functional dispersion (FDis). Taxonomic β -diversity metrics = local community contributions to taxonomic diversity (LCBD taxonomic); functional β -diversity metrics = local community contributions to functional diversity (LCBD functional). Proportion of protected cells along an elevation gradient considering protected areas (b). The upper maps show the distribution of the protected areas in Switzerland (in grey).



Appendix S36. Overlap between species occurrence and protected areas. The X-axis shows the 550 species sampled in the unfiltered 6200 community plots (each bar is a species) and the Y-axis depicts the proportion of occurrences in protected areas *sensu lato* (PA SL), protected areas *sensu stricto* (PA ss) and in unprotected areas (Not protected). Seven species only occurred in protected areas s.s., that is, *Andrena saxonica*, *Andrena suerinensis*, *Andrena synadelpha*, *Hylaeus pilosulus*, *Lasioglossum quadrisignatum*, *Megachile genalis*, *Sphecodes marginatus*. Nine species only occurred in protected areas s.l., that is, *Ammobates punctatus*, *Anthophora dispar*, *Ceratina nigrolabiata*, *Eucera nigrifacies*, *Hylaeus crassanus*, *Nomada discrepans*, *Osmia steinmanni*, *Sphecodes dusmeti*, *Thyreus histrionicus*.

Supplementary references

- Allen, M., Poggiali, D., Whitaker, K., Marshall, T.R. & Kievit, R. (2019) Raincloud plots: a multi-platform tool for robust data visualization. *Wellcome Open Research*, **4**.
- Altwegg, D. & Weibel, F. (2015) *L'utilisation du sol en Suisse. Exploitation et analyse*, Neuchatel.
- Brosi, B.J., Daily, G.C., Shih, T.M., Oviedo, F. & Durán, G. (2007) The effects of forest fragmentation on bee communities in tropical countryside. *Journal of Applied Ecology*, **45**, 773–783.
- Buchholz, S., Gathof, A.K., Grossmann, A.J., Kowarik, I. & Fischer, L.K. (2020) Wild bees in urban grasslands: Urbanisation, functional diversity and species traits. *Landscape and Urban Planning*, **196**, 103731.
- Cane J.H. & Sipes S. (2006). Characterizing floral specialization by bees: analytical methods and a revised lexicon for oligolecty. *Plant-pollinator interactions: from specialization to generalization*, 99-122.
- Cariveau, D. P., Nayak, G. K., Bartomeus, I., Zientek, J., Ascher, J. S., Gibbs, J., & Winfree, R. (2016). The Allometry of Bee Proboscis Length and Its Uses in Ecology. *PLOS ONE*, 11(3), e0151482. <https://doi.org/10.1371/journal.pone.0151482>
- Evans, E., Smart, M., Cariveau, D. & Spivak, M. (2018) Wild, native bees and managed honey bees benefit from similar agricultural land uses. *Agriculture, Ecosystems and Environment*, **268**, 162–170.
- Federal Office for the Environment FOEN, 2002. Modification de l'ordonnance sur les réserves d'oiseaux d'eau et de migrants d'importance internationale et nationale (OROEM ; RS 922.32)
- Federal Office for the Environment FOEN, 2003. Smaragd-Netz in der Schweiz.
- Federal Office for the Environment FOEN, 2012a. Biotopes d'importance nationale Stratégie de restauration écologique des biotopes d'importance nationale.

- Federal Office for the Environment FOEN, 2012b. Descriptifs des sites EMERAUDE suisses.
- Federal Office for the Environment FOEN, 2016. Patrimoine mondial de l'UNESCO. Plan d'action de la Suisse 2016 à 2023.
- Federal Office for the Environment FOEN, 2018a. Manuel de création et de gestion de parcs d'importance nationale.
- Federal Office for the Environment FOEN, 2018b. Manuel sur les conventions programmes 2020-2024 dans le domaine de l'environnement. Communication de l'OFEV en tant qu'autorité d'exécution.
- Federal Office for the Environment FOEN, 2020. Forest Policy 2020. Visions, objectives, and measures for the sustainable management of forests in Switzerland.
- Federal Office for the Environment FOEN, 2021. Forest Policy: objectives and measures 2021–2024.
- Federal Office for the Environment FOEN, 2022. État de la mise en œuvre des inventaires de biotopes d'importance nationale. Enquête auprès des cantons en 2021.
- Federal Statistical Office FSO, 2013. Land use in Switzerland: Results of the Swiss land use statistics. Gujarati, D. (1970) Use of Dummy Variables in Testing for Equality between Sets of Coefficients in Linear Regressions: A Generalization, *The American Statistician*, 24:5, 18-22, DOI: [10.1080/00031305.1970.10477220](https://doi.org/10.1080/00031305.1970.10477220)
- Karger, D. N., Conrad, O., Böhrner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W., Zimmermann, N. E., Linder, H. P. & Kessler, M. (2017) Climatologies at high resolution for the earth's land surface areas. *Scientific Data* **4**, 170122.
- Meier, E.S., Indermaur, A., Ginzler, C. & Psomas, A. (2020) An effective way to map land-use intensity with a high spatial resolution based on habitat type and environmental data. *Remote Sensing*, **12**, 1–21.
- Müller, A., & Kuhlmann, M. (2008). Pollen hosts of western palaeartic bees of the genus *Colletes* (Hymenoptera: Colletidae): The Asteraceae paradox. *Biological Journal of the Linnean Society*, 95(4), 719–733. <https://doi.org/10.1111/j.1095-8312.2008.01113.x>
- Pronatura, 2022. <https://www.pronatura.ch/en> [accessed 11.2022]

- Ramsar, 2022. <https://www.ramsar.org/> [accessed 11.2022]
- Robertson C. (1925). Heterotropic bees. *Ecology*, 6, 412-436.
- Swiss National Park, 2022. <https://www.nationalpark.ch/en/?changelang> [accessed 11.2022]
- UNESCO, 2022. UNESCO biosphere reserves. <https://en.unesco.org/biosphere> [accessed 11.2022]