



## Novel indices reveal that pollinator exposure to pesticides varies across biological compartments and crop surroundings

Marion LAURENT<sup>a</sup>, Stéphanie BOUGEARD<sup>b</sup>, Lucile CARADEC<sup>c</sup>, Florence GHESTEM<sup>c</sup>, Matthias ALBRECHT<sup>d</sup>, Mark J.F. BROWN<sup>e</sup>, Joachim DE MIRANDA<sup>f</sup>, Reet KARISE<sup>g</sup>, Jessica KNAPP<sup>f,h</sup>, José SERRANO<sup>i</sup>, Simon G. POTTS<sup>j</sup>, Maj RUNDLÖF<sup>f</sup>, Janine SCHWARZ<sup>d</sup>, Eleanor ATTRIDGE<sup>k</sup>, Aurélie BABIN<sup>a</sup>, Irene BOTTERO<sup>h</sup>, Elena CINI<sup>j</sup>, Pilar DE LA RÚA<sup>i</sup>, Gennaro DI PRISCO<sup>l,m</sup>, Christophe DOMINIK<sup>n</sup>, Daniel DZUL<sup>i</sup>, Andrés GARCÍA REINA<sup>i</sup>, Simon HODGE<sup>h</sup>, Alexandra M. KLEIN<sup>o</sup>, Anina KNAUER<sup>d</sup>, Marika MAND<sup>g</sup>, Vicente MARTÍNEZ LÓPEZ<sup>i</sup>, Giorgia SERRA<sup>l</sup>, Helena PEREIRA-PEIXOTO<sup>o</sup>, Risto RAIMETS<sup>g</sup>, Oliver SCHWEIGER<sup>n</sup>, Deepa SENAPATHI<sup>j</sup>, Jane C. STOUT<sup>h</sup>, Giovanni TAMBURINI<sup>o</sup>, Cecilia COSTA<sup>l</sup>, Tomasz KILJANEK<sup>p</sup>, Anne-Claire MARTEL<sup>a</sup>, Sébastien LE<sup>c</sup>, Marie-Pierre CHAUZAT<sup>a,q,\*</sup>

<sup>a</sup> Anses, Sophia Antipolis laboratory, Unit of Honeybee Pathology, France

<sup>b</sup> Anses, Ploufragan-Plouzané-Niort Laboratory, Epidemiology and welfare of pork, France

<sup>c</sup> CNRS, Statistics and Computer Science Department, L'Institut Agro Rennes-Angers, UMR 6625 IRMAR CNRS, 35042 Rennes Cedex, France

<sup>d</sup> Agroscope, Agroecology and Environment, Reckenholzstrasse 191, 8046 Zurich, Switzerland

<sup>e</sup> Department of Biological Sciences, School of Life Sciences and the Environment, Royal Holloway University of London, Egham, UK

<sup>f</sup> Department of Biology, Lund University, Lund, Sweden

<sup>g</sup> Chair of Plant Health, Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences, Fr. R. Kreutzwaldi 1a, 51006 Tartu, Estonia

<sup>h</sup> Department of Botany, School of Natural Sciences, Trinity College Dublin, Dublin 2, Ireland

<sup>i</sup> Department of Zoology and Physical Anthropology, Faculty of Veterinary, University of Murcia, 30100 Murcia, Spain

<sup>j</sup> School of Agriculture, Policy and Development, Reading University, RG6 6AR, UK

<sup>k</sup> Federation of Irish Beekeepers' Associations, Ireland

<sup>l</sup> CREA - Research Centre for Agriculture and Environment, Bologna, Italy

<sup>m</sup> Institute for Sustainable Plant Protection, The Italian National Research Council, Napoli, Italy

<sup>n</sup> Helmholtz Centre for Environmental Research - UFZ, Dep. Community Ecology, Theodor-Lieser-Strasse 4, 06120 Halle, Germany

<sup>o</sup> Nature Conservation and Landscape Ecology, University of Freiburg, Germany

<sup>p</sup> PIWET, Department of Pharmacology and Toxicology, National Veterinary Research Institute, Pulawy, Poland

<sup>q</sup> Paris-Est University, Anses, Laboratory for Animal Health, Maisons-Alfort, France

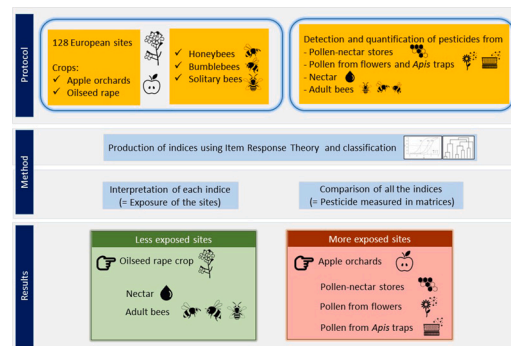
\* Corresponding author at: ANSES, Laboratory for Animal Health, 14 rue Pierre et Marie Curie, F-94701 Maisons-Alfort Cedex, France.

E-mail address: [marie-pierre.chauzat@anses.fr](mailto:marie-pierre.chauzat@anses.fr) (M.-P. CHAUZAT).

## HIGHLIGHTS

- We use new indices to summarise big datasets on pesticide exposure of three species of bees
- Novel indices are calculated using Item Response Theory (IRT) models
- The indices are linked to the number of pesticides rather than the active ingredients
- Matrices collected from apple orchards are exposed to a higher number of pesticides compared to oilseed rape crop sites
- Pollen related matrices contained more pesticides than were found in nectar and on the bees themselves

## GRAPHICAL ABSTRACT



## ARTICLE INFO

Editor: Yolanda Picó

## Keywords:

Item response theory  
Bumblebee  
Osmia  
Apple orchards  
Oilseed rape

## ABSTRACT

Declines in insect pollinators have been linked to a range of causative factors such as disease, loss of habitats, the quality and availability of food, and exposure to pesticides. Here, we analysed an extensive dataset generated from pesticide screening of foraging insects, pollen-nectar stores/beebread, pollen and ingested nectar across three species of bees collected at 128 European sites set in two types of crop. In this paper, we aimed to (i) derive a new index to summarise key aspects of complex pesticide exposure data and (ii) understand the links between pesticide exposures depicted by the different matrices, bee species and apple orchards versus oilseed rape crops. We found that summary indices were highly correlated with the number of pesticides detected in the related matrix but not with which pesticides were present. Matrices collected from apple orchards generally contained a higher number of pesticides (7.6 pesticides per site) than matrices from sites collected from oilseed rape crops (3.5 pesticides), with fungicides being highly represented in apple crops. A greater number of pesticides were found in pollen-nectar stores/beebread and pollen matrices compared with nectar and bee body matrices. Our results show that for a complete assessment of pollinator pesticide exposure, it is necessary to consider several different exposure routes and multiple species of bees across different agricultural systems.

## 1. Introduction

Declines in species of both managed and wild pollinators has been repeatedly documented (Potts *et al.*, 2010) in Europe (IPBES *et al.*, 2016), the US (Kulhanek *et al.*, 2017), Canada (CAPA, 2022), Asia (van der Zee *et al.*, 2012) and to some extent in South-America (Maggi *et al.*, 2016) and Africa (Pirk *et al.*, 2014). Managed bees such as honeybees (*Apis mellifera*) (Moritz and Erler, 2016) and wild bees (Soroye *et al.*, 2020; Powney *et al.*, 2019) are the most important group of pollinators in Europe and other regions of the world (IPBES, 2016). A range of factors have been suggested to explain losses of bees such as diseases (Pritchard *et al.*, 2021; Oddie *et al.*, 2021), loss of habitats (Rollin *et al.*, 2019; Ollerton *et al.*, 2014), the quality and availability of food (Somme *et al.*, 2016; Di Pasquale *et al.*, 2016) and exposure to pesticides (Siviter *et al.*, 2021; Zioga *et al.*, 2020). The way bees are exposed to pesticides is variable and depends mainly on the type of pesticide (Ward *et al.*, 2022; Straw and Brown, 2021), their purpose of use (which is related to the application mode i.e. spray, soil treatment, trunk injection), (Graham *et al.*, 2022) and on the ecology of species (Rundlof *et al.*, 2015; Knapp *et al.*, 2023). Application timing (pre-bloom versus at-bloom) has logically dramatic impacts on exposure levels for pollinators feeding on nectar and pollen from flowers (Zioga *et al.*, 2020). Several techniques have been developed to limit this exposure such as microencapsulated compounds and seed coated insecticides with systemic properties (Wisk *et al.*, 2014). Bees can also be exposed to pesticides through water consumption (Carter *et al.*, 2020; McCune *et al.*, 2021), pesticide contact (Arena and Sgolastra, 2014), air (Ward *et al.*, 2022; Negri *et al.*, 2015; Pochi *et al.*, 2012) and, in the case of managed bees, the use of veterinary products (Mahefarisoa *et al.*, 2021; Kiljanek *et al.*, 2021). However, dietary consumption is the major route of exposure (Zioga *et al.*, 2020).

Honeybees produce large quantities of honey from collected nectar. In addition, for storage purposes, after collection, pollen grains are processed into beebread. This term usually refers to honeybee pollen stores, as beebread is pollen with added nectar and enzymes (Pavlova *et al.*, 2022) and stored in frames made of beeswax. For other bee species, however, any substance consisting predominantly of stored pollen will be referred to as pollen-nectar stores in this paper.

Previously, pesticide residues have been documented in nectar (Zioga *et al.*, 2020), honey (Kavanagh *et al.*, 2021), pollen collected on flowers (Ward *et al.*, 2022), honeybee pollen pellets collected with traps (Favaro *et al.*, 2019), honeybee beebread (Raimets *et al.*, 2020), wax (Mullin *et al.*, 2010) and honeybees themselves (Martinello *et al.*, 2020). However, the majority of exposure studies describe the contamination of one or two matrices at the same time (Demares *et al.*, 2022). To our knowledge, our study is the first to present results across pesticides in pollen collected from flowers and from pollen pellets, in pollen-nectar stores and beebread, in nectar regurgitated from honeybees and from other bee species and from bee bodies, collected at the same time in the same site. In an attempt to better understand the exposure route of three bee species (*Apis mellifera*, *Bombus terrestris* and *Osmia bicornis*), we assessed pesticides in each of these matrices at the same time in 128 sites set in two types of crops (apple orchards, oilseed rape) across Europe. To our knowledge, this dataset is one of the most extensive datasets of bee exposure to pesticides currently available.

As the number of pesticides measured in the different matrices and for each site was very large, it was necessary to synthesise this complex information. The construction of such indices, that are able to summarise information for all pesticides detected at a site, is of paramount interest. Such an index can be used, for instance, for investigating the links between the different matrices under study or in structuring model

equations to explore the role of stresses on bee population dynamics. A classic way to summarise pesticide information is to calculate the richness (i.e., the number of pesticides detected in a given sample), or the abundance (i.e., the total quantity of pesticides detected in a given sample) (Traynor et al., 2021). However, these simple calculations do not capture information on pesticide variability across the samples. In this paper, we propose to apply an original method, namely Item Response Theory (IRT) models to calculate an index that includes as much variability as possible while being easily interpretable.

The IRT models build such indices, each being associated with a matrix (i.e., pollen-nectar stores or beebread, pollen, nectar and foragers from different species and flowers) and a crop (i.e., apple orchards, oilseed rape). We also propose a method to interpret these indices (Section 3.1). In a second step, the links between all these indices are studied (Section 3.2). Results are discussed in the context of the existing literature (Section 4).

## 2. Materials and methods

### 2.1. Samples collection in PoshBee site network

Within the H2020 project ‘PoshBee’ ([www.PoshBee.EU](http://www.PoshBee.EU)), a site network for assessing exposure of bees to chemical, nutritional, and pathogen stressors was established in 2019 (Hodge et al., 2022). Data were collected at 128 sites across eight participating countries (Estonia, Germany, Ireland, Italy, Spain, Sweden, Switzerland and the United Kingdom) situated in either apple orchards or oilseed rape crops. At each site, three honeybee colonies, three trap nests seeded with male and female cocoons of *Osmia bicornis* (solitary bee) and three *Bombus terrestris* (bumblebee) colonies were installed following the PoshBee protocols (Hodge et al., 2022).

At each site, various matrices were collected from all colonies and nests in equal proportions, pooled per species and subsequently sent for pesticide residues analyses in different laboratories (Hodge and Stout, 2019). If field constraints prevented the collection of equal proportions, acceptable differences between colony/nest were limited to a maximum of 30 %. If one colony/nest did not produce the quantity required, the quantities from the remaining two were increased in order to reach the total quantity required. The sampling of each matrix was performed only once for each species at each site generally on the same day. Depending on the matrix, sampling was performed either during or towards the end

of the flowering period to be consistent with biological cycles of bees (Fig. A1 and Fig. A2, in supplementary material). At each site, *A. mellifera* and *B. terrestris* adults were collected alive. Bees were gently pressed at the two first abdominal segments on the crop (honey sack) until a drop of nectar was regurgitated between the bee mandibles. Nectar was collected and pooled for each species to produce one sample per species for each site for pesticide analysis.

The matrices listed in Fig. A1 were sampled and subsequently analysed for determination and quantification of pesticide residues. Due to the behavior and limited success of solitary bees in the wild, it was not possible to obtain sufficient numbers of *O. bicornis* bees or amounts of regurgitated nectar to perform analyses for pesticide residues on these matrices (Table 1).

### 2.2. Analytical methods for pesticide determination and quantification

Four different laboratories analysed the samples to identify and quantify pesticide residues. Each laboratory was in charge of a specific matrix and had specific developed and validated methods with liquid chromatography tandem mass spectrometry (LC-MS/MS) and gas chromatography tandem mass spectrometry (GC-MS/MS). The different analytical methods were detailed for pollen-nectar stores and beebread (Kiljanek et al., 2021), nectar (Martel et al., submitted), bees (Serra et al., 2021) and pollen from flowers and from traps.

Sample preparation for residue analysis of 261 pesticides and their metabolites as well as 6 congeners of non-dioxin like polychlorinated biphenyls (ndl-PCB) in a very low mass beebread or pollen-stores samples was based on modified QuEChERS protocol with all steps miniaturized to enable multiresidue analysis. Sample of beebread (0.3 g) was extracted with 1 mL of acetonitrile containing 5 % formic acid and ammonium formate salt was added for partitioning. Then the supernatant was subjected to clean-up by freezing and two-step dispersive solid phase extraction (dSPE) into a Supel™ QuE Verde mini tube with sorbents (Supelclean™ ENVI-Carb™ Y, 10 mg; Supelclean™ PSA, 50 mg; Z-Sep+, 60 mg; magnesium sulfate, 150 mg). After 1st step dSPE, a portion of extract was analysed by LC-MS/MS for 200 pesticide residues. Remaining extract was subjected to a 2nd step dSPE clean-up by another Supel™ QuE Verde mini tube and then after evaporation to dryness and dissolved in hexane, it was analysed by GC-MS/MS for another 61 pesticide and 6 ndl-PCB residues. Method enabled determination of residues of 101 insecticides, 72 herbicides, 67 fungicides, 10 acaricides,

**Table 1**

Overview of the number of sites sampled and analysed, the number of pesticides screened and detected in each matrix for each species and crop corresponding to the 18 datasets included in the indices calculation. The percentages of sites with analysed samples were compared to the theoretical number of samples according to the protocol (=64 samples for each matrix, i.e., 8 sites × 8 countries). A. m: *Apis mellifera*. B. t: *Bombus terrestris*. O. b: *Osmia bicornis*. APP: apple. OSR: oilseed rape. Apis: pollen collected with pollen traps set up on *A. mellifera* colonies.

Matrix	Type	Crop	Number of sites with samples sent to the laboratories	Number of sites with analysed samples (%) - N	Number of sites with at least one pesticide detected (%)	Number of pesticides detected in at least one site - P	Number of pesticides screened
Beebread	A. m	APP	62	62 (97 %)	62 (100 %)	98 (37 %)	267
		OSR	62	62 (97 %)	62 (100 %)	78 (29 %)	267
Pollen-nectar stores	B. t	APP	56	51 (80 %)	50 (98 %)	97 (36 %)	267
		OSR	60	56 (87 %)	55 (98 %)	84 (31 %)	267
	O. b	APP	42	42 (65 %)	39 (93 %)	79 (30 %)	267
		OSR	47	46 (72 %)	46 (100 %)	73 (27 %)	267
Bees	A. m	APP	64	64 (100 %)	41 (64 %)	22 (6 %)	373
		OSR	64	64 (100 %)	24 (37 %)	14 (4 %)	373
	B. t	APP	61	61 (95 %)	36 (59 %)	19 (5 %)	373
		OSR	64	64 (100 %)	21 (33 %)	11 (3 %)	373
Nectar	A. m	APP	64	64 (100 %)	49 (77 %)	21 (25 %)	85
		OSR	63	63 (98 %)	42 (67 %)	16 (19 %)	85
	B. t	APP	61	60 (94 %)	47 (78 %)	24 (28 %)	85
		OSR	64	64 (100 %)	39 (61 %)	14 (16 %)	85
Pollen	Apis	APP	56	56 (87 %)	49 (88 %)	44 (13 %)	336
		OSR	63	61 (95 %)	44 (72 %)	25 (7 %)	336
	Flowers	APP	51	26 (41 %)	26 (100 %)	58 (19 %)	300
		OSR	53	34 (53 %)	32 (94 %)	57 (19 %)	300

6 growth regulators, 5 veterinary drugs and 6 ndl-PCB's. The limits of quantification (LOQ) values have been established as follows: 0.001 mg/kg for 105, 0.005 mg/kg for 96, 0.01 mg/kg for 31, 0.05 mg/kg for 31 and 0.1 mg/kg for 4 compounds respectively.

For the determination of 85 pesticides in nectar regurgitated by honey bees and bumble bees two multiresidue methods were applied. One method involved an extraction of 40 pesticides in 10 µL of nectar sample and an analysis with LC-MS/MS. The other one for the determination of 45 pesticides in 100 µL of nectar sample involved a liquid/liquid partitioning with an organic solvent and an evaporation to dryness. Then the extract was recovered with appropriate solvent for GC-MS/MS analysis. The LOQ were ranged between 5 and 10 pg/µL for LC-MS/MS and between 10 and 521 pg/µL for GC-MS/MS.

In the bee bodies (honey bees and bumble bees) the total number of molecules that was screened for, inclusive of isomers and metabolites, was 373. A simplified QuEChERS method was used for sample preparation, which consisted of an extraction with water, acetonitrile and salts (MgSO<sub>4</sub> and NaCl). After centrifugation, the supernatant was cleaned-up on PSA (dSPE). The sample was again centrifuged, concentrated and a specific solvent was added for the GC-MS/MS or LC-MS/MS analysis. The LOQ were between 0.002 and 0.05 mg/kg.

Sample preparation of pollen to quantify >336 compounds was based on modified QuEChERS protocol. Water was added to the prepared homogeneous samples (1 g) before extraction with acetonitrile. Samples below 300 mg could not be analysed. Magnesium sulfate, sodium chloride and sodium citrate salts were added to the sample for liquid/liquid partitioning. A portion of the organic phase was subjected to a step of freezing following by a clean-up on a mixture of MgSO<sub>4</sub>, PSA and C18 (dSPE). Then, the extract with 5 % of formic acid in acetonitrile was directly analysed by LC-MS/MS. The GC-MS analysis required the change of solvent to hexane/acetone 4:1 (v/v). Limits of quantification were 0.005 to 0.01 mg/kg.

This resulted in five different lists of pesticides depending on matrices. However, 64 common pesticides were selected at the beginning of PoshBee based on agrochemicals applied on crops at the European level to enable comparison between matrices. The index calculation was not restricted to these 64 pesticides. Indeed, if a pesticide was detected in only one matrix, it contributed to increase the exposure in the site where it was detected. As a consequence, the indices' values increased. At the end, 267 pesticides were screened for in pollen-nectar stores and beebread, 373 pesticides in foragers, 85 pesticides in nectar, 336 pesticides in pollen from *A. mellifera* traps and 300 pesticides in pollen from flowers.

A minimum quantity was required to perform laboratory analysis. This requirement was not always met due to field constraints. Thus, results were missing for some sites or matrices. At the end, 319 pollen-nectar store/beebread samples, 253 forager samples, 251 nectar samples, 117 *A. mellifera* pollen-trap samples and 60 flower pollen samples were analysed (Table 1).

The quality and consistency of all the analytical results was automatically controlled in a database designed for this purpose (named Poshbase) enabling the collection of 18 datasets corresponding to the matrices across the three bee species (Table 1).

The theoretical number of sites under study was 64 for a given matrix and crop (Table 1). However for various reasons (i.e. quantity of sampled matrix not sufficient for subsequent laboratory analysis, difficulty to retrieve matrix from the field due to weather conditions or scarce quantity), the actual number of sites in the statistical analysis was reduced. The largest reduction was observed for the pollen collected directly on flowers in apple orchards ( $N = 26$ ) and oilseed rape ( $N = 34$ ). The number of sites with at least one pesticide detected in a matrix varied from 100 % in beebread from honeybee colonies in apple orchards or oilseed rape and in pollen-nectar stores from solitary bees' nests in oilseed rape crops for instance, to 33 % in bumblebee foragers in oilseed rape crops. Between 11 (in bumblebees in oilseed rape crops) and 98 (in honeybee beebread collected in colonies in apple orchards)

pesticides were detected in any given matrix, representing between 3 % and 37 % of the pesticides screened for.

As the calculation of the indices was intended to give the best discrimination between sites, only pesticides detected in at least one site were taken into account. Thus, each dataset used for the statistical analysis was of dimension  $N \times P$  (Table 1; e.g. for *Beebread.Apis* and for apple orchards,  $P = 98$  pesticides were detected and measured in  $N = 62$  sites) and included the quantification of each pesticide in each site. More precisely for a given site, a given pesticide and a given matrix, the following rules were applied: the LOQ (limit of quantification, the pesticides detected below this value cannot be quantified) was used for values between the LOD (limit of detection; below this value, the pesticides cannot be detected with sufficient confidence) and the LOQ, and quantified values were kept in cases of values higher than LOQ. As the data had many zeros (i.e., non-detected pesticides), the calculation of the indices was based on binary data: 0 was used if the value was inferior to LOD and 1 was used otherwise. However, the index's interpretation was based on raw quantified values.

### 2.3. Statistical analyses

Our aim was to summarise and interpret the large amount of information available in each dataset. For this purpose and in a first step, 18 indices were built, one for each matrix and each crop. The objective was to reduce the dimensionality of the datasets to characterise the site exposure to pesticides in a unidimensional and interpretable index. Subsequently, each index was interpreted according to the pesticides detected. Finally, and for each crop, the links between the nine indices were studied with a Principal Component Analysis as a summary of correlation matrix (Fig. 1).

**Calculation of indices.** Initially developed in the psychology framework, the Item Response Theory (IRT) models aim at building a unidimensional scale (= latent trait = index), from different items that measure this trait (Bock and Aitkin, 1981; Van der Linden and Hambleton, 1997). The IRT concept was translated as to whether a site exhibited a given pesticide or if the pesticide was absent from the site. The more pesticide were recorded the  $i^{\text{th}}$  site was, the higher its index value, denoted  $\theta_i$ .

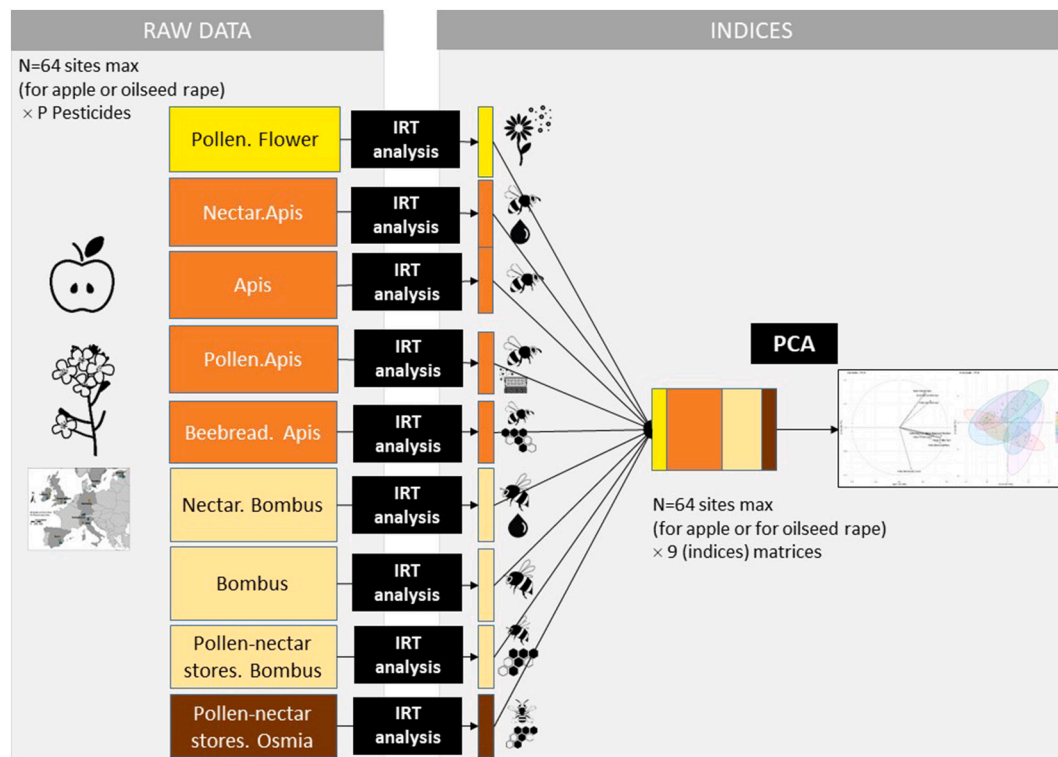
For a given pesticide  $j$ , the two parameters to be estimated in the model were the mean exposure level of a site ( $a_j$ ) and the specific exposure level of a site ( $b_j$ ), fitted with an EM algorithm (Chalmer, 2012). The exposure level (measured here as the number of detected pesticides per site) was the level a site should have, to have 50 % chance to exhibit a pesticide. The specific exposure level represented how well the item (i.e. pesticide) separated sites with high exposure scores from sites with low exposure scores. In theory, most, if not all pesticides, should have a positive specific exposure level: the more exposed a site was, the more likely it was to detect a given pesticide. For this purpose, the following two-parameter logistic model was applied. Let  $P(X_{i,j}|\theta_i)$  be the probability that the site  $i$  exhibited the pesticide  $j$  given its exposure level, such as:

$$P(X_{i,j}|\theta_i) = \frac{1}{1 + e^{-a_j(\theta_i - b_j)}} \text{ for the } j^{\text{th}} \text{ pesticide and the } i^{\text{th}} \text{ site } (i = 1, \dots, 64)$$

With  $a_j$  the exposure level,  $b_j$  the site-discrimination and  $\theta_i$  the level of exposure at site  $i$ .

For several pesticides under study, the previous model was adapted: all the pesticides were included and then selected through a backward selection algorithm applied to filter out non-interpretable pesticides. To maximize the statistical significance of the two parameters ( $a_j$  and  $b_j$ ), a double control on each step of the algorithms was implemented: (i) a stepwise loop stopped if there were no more pesticides with a negative discrimination, or (ii) if the performance criterion of the model (=Akaike information criterion, AIC) stopped decreasing. At the end, only pesticides with a positive discrimination were retained. In addition,





**Fig. 1.** The overall statistical procedure for a given crop (apple orchard or oilseed rape) for the nine matrices across the three bee species (*Pollen.Flower*, *Nectar.Apis*, *Apis*, *Pollen.Apis*, *Beebread.Apis*, *Nectar.Bombus*, *Bombus*, *Pollen-nectar stores.Bombus*, *Pollen-nectar stores.Osmia*). The map is from [Hodge et al., 2022](#). IRT: Item Response Theory. PCA: Principal Component Analysis.

the stability of the selection was tested with a leave-one-out cross validation, both on sites and pesticides. In summary, using the index was relevant when the information on the pesticide detection was fragmented between different pesticides (see the discussion for details).

**Interpretation of indices.** The index was calculated on pesticide presence or absence to have robust calculations and deal with the many zeros. However, as the interpretation was not based on more robust statistical tests, the quantities of pesticides from the raw quantified data were used ([Table 2](#)). For a given matrix and a given crop, the pesticides, as well as countries, that most contributed to the index were highlighted and interpreted. For this purpose, all the available sites were clustered by means of a Hierarchical Clustering Analysis applied to each index value ([Everitt, 1974](#)). Then, the pesticides that were significantly over-represented in a cluster compared to the mean were highlighted ([Husson et al., 2017](#)). Similarly, under-represented pesticides compared to the mean could also be identified; they were detailed only in [Table 2](#) for the example and interpretation. Two supplementary variables (i.e., number of pesticides and country) were also taken into account. Sites of a given country that were over- or under-represented in a cluster compared to the mean were also highlighted. Consequently, the interpretation of presence/absence of sites from a given country compared to sites from other countries was possible (see [Table 2](#)). It is worth noting that the number of sites per country ( $N = 8$  sites) did not allow the extrapolation of results to the whole country. Indeed, the site network was not designed to be representative of countries, but rather to be representative of these crop landscapes in the European territory.

**Links between indices.** For a given crop (apple or oilseed rape), the links between the nine indices - related to the different matrices - were studied with a Principal Component Analysis (PCA) ([Jolliffe, 1986](#)).

All the analyses were implemented in R software (version 4.1.3 <https://www.r-project.org/>). The IRT models were estimated using the *mirt* R package with the 'Rasch' option. The clustering was applied with the *HCPC* function of the *FactoMineR* package ([Lé et al., 2008](#)) and the interpretation of the indices was made with the *catdes* (for categorical

variable such as country) or *condes* (for numeric variable such as the number of pesticides) functions of the *FactoMineR* package. Principal Component Analyses were performed with the PCA function of the *FactoMineR* package.

### 3. Results

#### 3.1. Indices: IRT results and interpretation

##### 3.1.1. Detailed interpretation of indices related to beebread collected in *A. mellifera* colonies in apple orchards

As a proof of principle, we chose to interpret in detail the index of site characterisation for a single dataset: the pesticide residues detected in beebread collected from *A. mellifera* colonies in the 62 apple orchard sites ([Table 2](#)). The complete set of the indices' values for each site and the interpretation of the indices are given in [Tables A.1 to A.4](#) (in supplementary material).

According to their index values, the sites were separated into four clusters. The statistical differences between clusters highlighted the unequal repartition of detected pesticides. In other words, if a pesticide was detected (respectively not detected) in a limited number of clusters, it was qualified as an over-represented (respectively under-represented) pesticide. If a pesticide was present in all the clusters, it was not considered as over-represented. Pesticides were less present in Cluster 1 ( $N = 10$  sites out of the 62) than the mean calculated across all sites. It presented the lowest index value ( $-1.32$ ). Only a few pesticides (mean of 3.90) were detected in samples and none were over-represented compared to the mean. Estonian sites were the most frequent in this cluster. Cluster 2 ( $N = 12$ ) did not contain sites over or under-represented compared to the mean. The index value was negative ( $-0.49$ ) but higher than cluster 1's, meaning that cluster 2's sites were exposed to fewer pesticides than the mean calculated across all sites but exposed to a higher number of pesticides than the sites in the cluster 1. Cluster 3 (index value of 0.16) contained most of the sites ( $N = 21$ )

Table 2

Field site characterisation based on the index calculated on pesticide residues detected in beebread collected in *A. mellifera* colonies in the 62 apple orchards sites. CHE: Swiss sites. EST: Estonian sites. GER: German sites. IRL: Irish sites. SWE: Swedish sites. UK: The United Kingdom sites.

Cluster	Cluster 1	Cluster 2	Cluster 3	Cluster 4
Number of sites	10	12	21	19
Mean index	−1.32	−0.49	0.16	0.83
Mean number of pesticides	3.90	9.67	15.8	23.7
Number of pesticides over-represented compared to the mean with p-value<0.05 (% compared to the total pesticides detected in at least one site)	0	7 (7.1 %)	0	30 (30.6 %)
The most over-represented pesticides compared to the mean (p-value <0.003). I: insecticides, F: fungicides (the mean concentration in µg/kg)	0	0	0	FLONICAMID I (78.2) PYRIMETHANIL F (1 090) FLUXAPYROXAD F (452) PENTHIOPYRAD F (62.8) BOSCALID F (555) DITHIANON F (9 230) GER (36.8 %) UK (31.6 %)
Countries over-represented within clusters compared to the mean with p-value<0.05 (% compared to the total number of sites in the cluster)	EST (50 %)	0	0	
Countries under-represented within clusters compared to the mean with p-value<0.05 (% compared to the total number of sites in the cluster)	0	0	0	CHE (0 %) IRL (0 %) SWE (0 %)

though no pesticide nor country was over- or under- represented. Cluster 4 ( $N = 19$ , index value of 0.83) included the sites exposed to a high number of pesticides with 30 pesticides over-represented compared to the mean. One insecticide (flonicamid) and five herbicides were the most significant pesticides ( $p < 0.005$ ). The concentrations in this cluster ranged from 9230 for the dithianon to 78.2 µg/kg for the flonicamid. The United Kingdom and German sites were over-represented in this cluster and therefore hosted sites with higher number of detected pesticides. Swiss, Irish and Swedish sites were significantly absent from Cluster 4. They were present in Clusters 1, 2 and 3 but not over-represented in any of these clusters.

3.1.2. Overall description of the indices

All 18 indices were highly positively correlated with the number of pesticides detected in the matrices (mean correlation = 0.99; Table A.5, in supplementary material). This meant the higher the value of an index, the more exposed to a high number of pesticides the site was (details in Tables A.3 and A.4). Generally, matrices collected from apple orchards were exposed to a higher number of pesticides than matrices collected from oilseed rape crops, with respectively 7.6 [3.3–11.9] versus 3.5 [0.9–6.1] pesticides on average (details in Tables A.3 and A.4). Fungicides were highly present in the pesticides significant for the discrimination of clusters: 70 % and 43.4 % in apple orchards sites and in oilseed rape crops, respectively (Table A.6). Insecticides (20 % and 33.9 %, respectively) and herbicides (10 % and 16.9 %, respectively) were the other pesticide families the most represented. The quantities of these

pesticides ranged from a minimum of 1.04 (insecticides) to a maximum of 9230 µg/kg (fungicides) in apple orchard sites; and from 0.47 (for insecticides and herbicides) to 2880 µg/kg (fungicides) in oilseed rape crop sites. Irrespective of the crop, pollen-nectar stores/bee bread and pollen matrices contained a higher number of pesticides than nectar and forager matrices (Tables A.7 and A.8, in supplementary material). For apple orchards for instance, 15.1 and 10.4 pesticides were found respectively in bee bread collected from *Apis* foragers and pollen from flowers whereas only 2.2 and 1.3 were found in nectar regurgitated from *Apis* foragers and in *Apis* foragers respectively. For oilseed rape, 14.9 and 7.7 pesticides were found in pollen-nectar stores from *Bombus* foragers and pollen from flowers respectively, whereas only 1.2 were found in nectar regurgitated from *Bombus* foragers and 0.4 in *Bombus* foragers themselves. It is worth noting that only 85 pesticides were screened for in nectar whereas hundreds were screened in pollen-nectar stores/bee bread, pollen and foragers. However, despite the high number of pesticides screened for in foragers, only a few were found.

The pesticide residue presence in **pollen-nectar stores/bee bread** collected from bees in apple orchards was high in sites located in Italy for *Bombus* and *Osmia* species and in Germany and the United Kingdom for *Apis* species. It was low in Estonian sites, irrespective of bee species (Fig. 3, Table A.3 and A.7). When looking at the pesticide residue presence in pollen-nectar stores/bee bread collected from bees in oilseed rape, the least exposed sites were in Estonia for *Apis* and *Osmia* species and in Switzerland for *Bombus* species (Fig. 3 and Table A.4). In addition, sites located in Germany and Spain for *Apis* species and in Italy for *Osmia* species were the most exposed according to the indices for pollen-nectar stores/bee bread. No country was over-represented in the exposed oilseed rape sites for *Bombus* species. Pesticides that characterised the indices were different between the two crops. For a given crop, different pesticides characterised the indices related to pollen-nectar stores/bee bread from the different bee species. In other words, pollen-nectar stores/bee bread collected by the three species did not contain the same type of pesticides irrespective of whether sampling sites were in apple orchards or in oilseed rape crops. However, the characterisation of the sites with a higher number of pesticides surrounded by oilseed rape included DMF (one metabolite of the acaricide amitraz) for pollen-nectar stores/bee bread collected from *Apis* (3.49 µg/kg) and *Bombus* species (7.9 µg/kg) and the herbicide S-metolachlor for pollen-nectar stores/bee bread collected from *Apis* (3.93 µg/kg) and *Osmia* species (122.1 µg/kg).

Irrespective of the focal crop, the pesticide residue presence in **pollen collected from flowers** was low in Spanish sites (Fig. 3, Tables A.3 and A.4). The insecticide diflubenzuron (17.7 and 80 µg/kg, respectively) and the fungicide dimetomorph (15.6 and 58.3 µg/kg, respectively) characterised the sites with a higher number of pesticides for pollen collected from apple orchard and oilseed rape flowers (Tables A.3 and A.4).

Looking at **pollen loads** collected from honeybee colonies in apple orchards, pesticide residue presence was high in sites located in Germany and low in sites located in Spain (Fig. 3 and Table A.3). For honeybee pollen loads collected in oilseed rape sites, no sites were over-represented in the highest cluster but Italian sites were over-represented in the lowest cluster (Fig. 3 and Table A.4). Different pesticides characterised the indices related to pollen loads in the two crops. In other words, pollen loads collected from honeybee colonies did not contain the same type of pesticides in apple orchards or in oilseed rape crops.

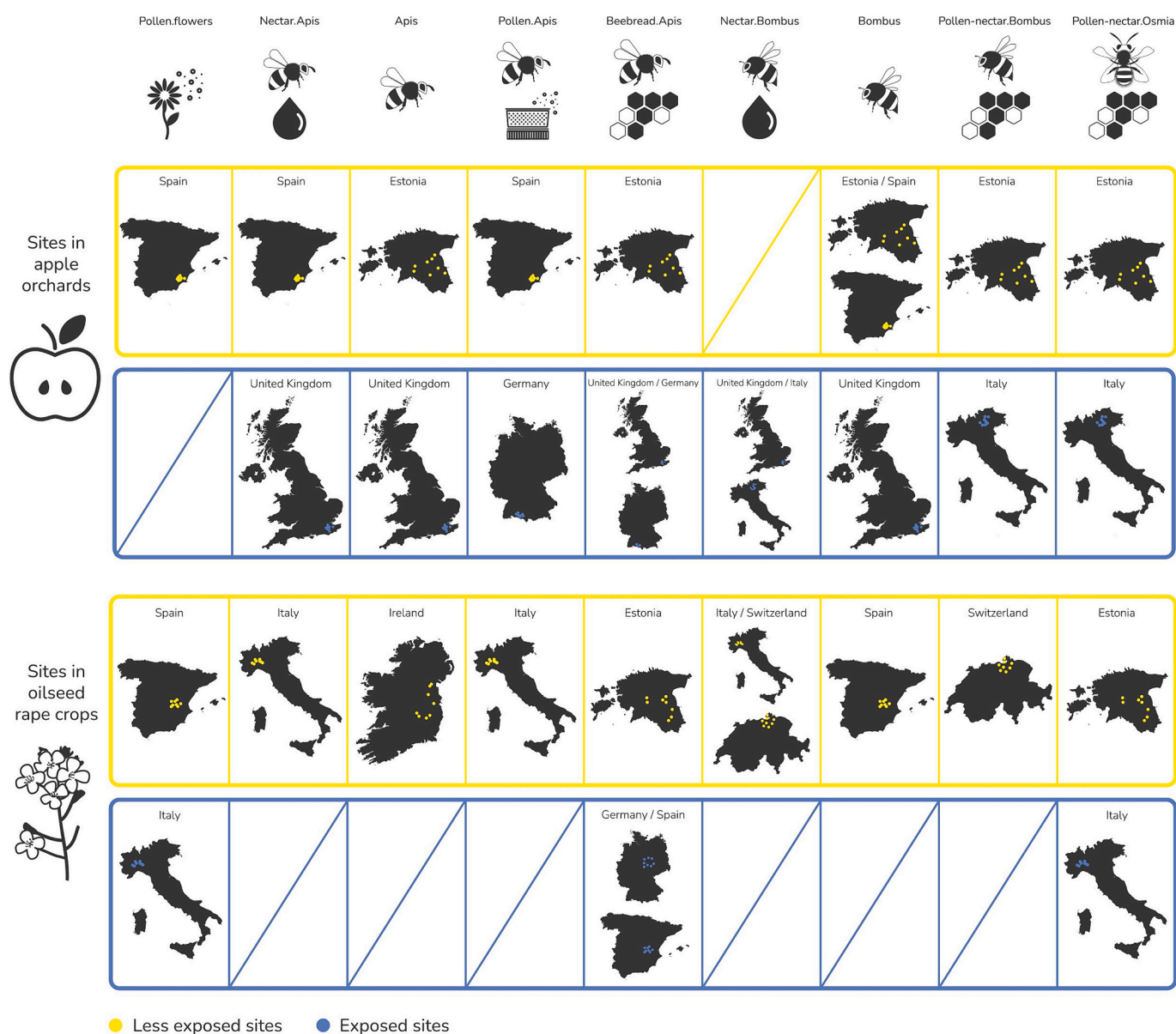
According to the indices, the **nectar** samples contained a higher number of pesticides when collected in the United Kingdom sites in apple orchard, and fewer pesticides in Italian sites in oilseed rape irrespective of the bee species (Fig. 3, Tables A.3 and A.4). The characterisation of the sites with a higher number of pesticides in apple orchards included the fungicide epoxyconazole (2.43 µg/kg in nectar collected by honeybees). It was also present in nectar (2.7 µg/kg) regurgitated from bumblebees collected in by oilseed rape sites and characterised the sites with a higher number of pesticides.

When looking at pesticides present in **bees** collected from apple orchards sites, the indices indicated that sites located in the United Kingdom had the highest number of pesticides and those located in Estonia had the lowest, irrespective of the bee species (Fig. 3, Tables A.3 and A.4). The pesticide residue presence in bees in oilseed rape crops was low in Irish sites for *Apis* species and in Spanish sites for *Bombus* species (Fig. 3, Tables A.3, A.4, A.7 and A.8). No country was over-represented with respect to oilseed rape in the most exposed (in terms of number of detected pesticides) sites. The characterisation of the most exposed sites in apple orchards included the pesticide 1,2,3,6 tetrahydrophthalimide (metabolite of a foliar fungicide Captan) for bees collected from both species (700.2 µg/kg in honeybees and 2170 µg/kg in bumblebees). It was also present in bumblebees collected in the most exposed sites in oilseed rape crops (197 µg/kg). The insecticide tau-fluvalinate characterised the most exposed sites in oilseed rape crops independently of the bee species. The fungicide boscalid characterised the most exposed sites in both crops for bees collected from *Apis* species (176 µg/kg in apple site and 275.2 µg/kg in oilseed rape sites).

For indices related to the matrices collected in apple orchards, the clusters of sites with the highest rank of exposure included sites from either Germany, Italy or the United Kingdom (Fig. 2). The clusters with the lowest rank of exposure included sites from either Estonia or Spain. Irish and Swiss sites were never over-represented in clusters for these indices. For the indices related to the matrices from sites in by oilseed rape crops, the clusters of sites with the highest rank of exposure included sites from either Germany, Italy or Spain. The clusters with the lowest rank of exposure included sites from either Estonia, Ireland, Italy, Spain or Switzerland. The United Kingdom and Swedish sites were never over-represented in clusters for these indices.

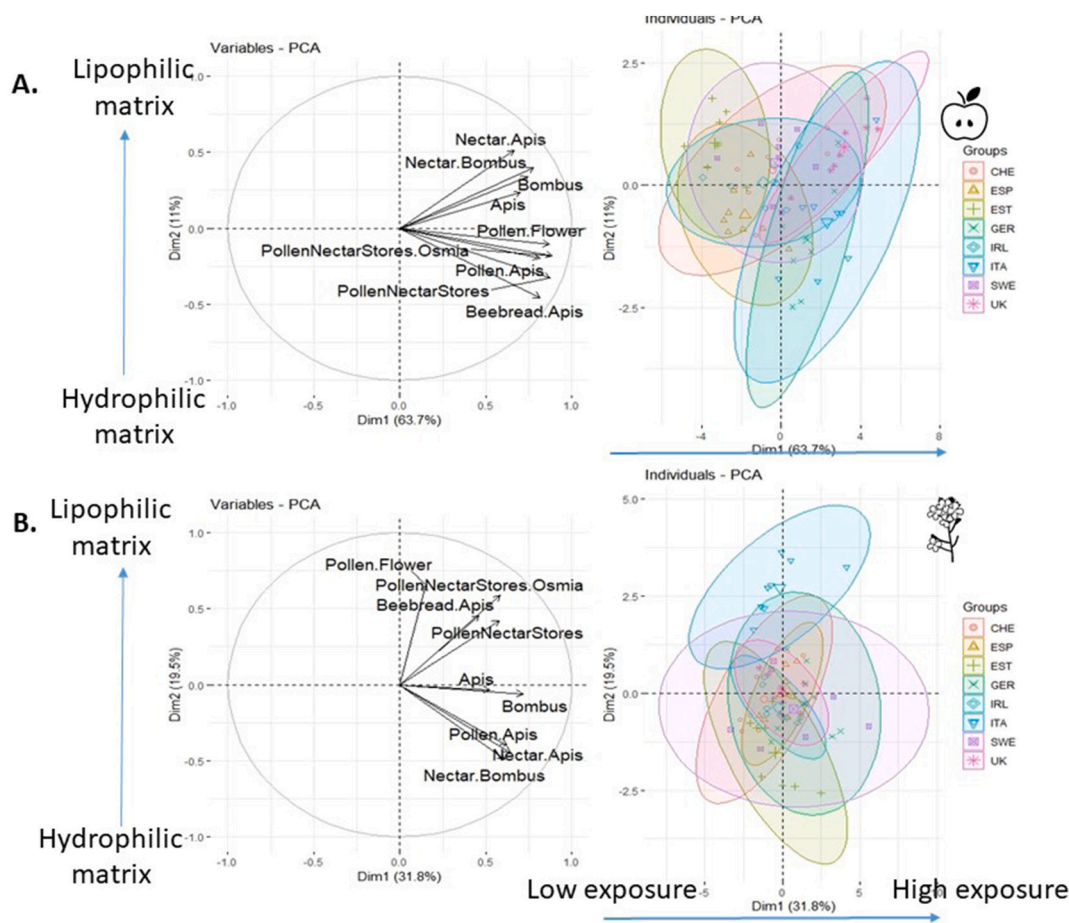
### 3.2. Links between the indices

The links between indices were illustrated by means of a PCA for matrices collected in apple orchards and in oilseed rape crops (Fig. 3). The PCA correlation circles of variables (left plots) represented the link between the nine indices related to each matrix for a given crop. The



**Fig. 2.** Summary of the sites that were most over-represented compared to the mean ( $p$ -value  $< 0.05$ ) in the clusters with low (yellow) and high (blue) number of pesticides based on IRT index values for the nine matrices. Sites in apple orchards are at the top of the figure, whereas those in oilseed rape are below. The bars mean that no sites were over-represented compared to the mean in a cluster.





**Fig. 3.** Graphical display of the first two components of the Principal Component Analysis of the nine indices (left) from the 64 sites (right) in **apple orchards** (A) or **oilseed rape crops** (B), the country being considered as a supplementary information. The interpretation arrows indicate the nature of the matrices regarding their content of fat (lipophilic, they attract molecules that dissolve in fats) and water (hydrophilic, they attract molecules soluble in water – see discussion for details) and their level pesticide content (low or high number of pesticides – details are given in the text).

plots on the right represent the 64 sites, the country being considered as a supplementary information. In data from apple orchard sites, 74.8 % of the overall inertia was explained. Inertia is the overall information contained in the data. The remaining 15.6 % of missing values were imputed. In data from oilseed rape sites, 51.3 % of the overall inertia was explained. The remaining 10.8 % of missing values were imputed.

Irrespective of the crop (Fig. 3), the positive correlations between the nine indices meant that the number of pesticides measured in the various matrices varied in the same way. As indices and number of pesticides were highly correlated (Section 3.1.2), the more detected pesticides there were in any given matrix, the more there were in related matrices. However detected pesticides were hardly the same.

In the apple orchard sites (Fig. 3A left), two bundles of variables were highlighted: on one hand, indices related to nectar regurgitated from *Apis* and *Bombus* foragers and to *Apis* and *Bombus* foragers themselves, and on the other hand, indices related to pollen-nectar stores/beebread collected from colonies and nests, pollen collected from flowers and pollen loads from *Apis* traps. The indices related to nectar were highly correlated with each other ( $\text{cor} = 0.69$ ) as well as with bumblebees ( $\text{cor} = 0.47$  for *Nectar.Apis/Bombus* and  $\text{cor} = 0.60$  for *Nectar.Bombus/Bombus*). The indices related to pollen-nectar stores/beebread collected in honeybee or in bumblebee colonies were highly correlated with each other ( $\text{cor} = 0.83$ ) and, to a lesser extent, to the one collected in solitary bee nests ( $\text{cor} = 0.79$  for *Pollen-nectar stores.Osmia/Beebread.Apis* and  $\text{cor} = 0.83$  for *Pollen-nectar stores.Osmia/Pollen-nectar stores.Bombus*). These three indices related to pollen-nectar stores/beebread were also linked with the pollen collected from flowers ( $\text{cor} = 0.72$  to  $0.75$ ) and

with the pollen loads collected from *Apis* traps ( $\text{cor} = 0.65$  to  $0.72$ ).

Some Italian apple orchard sites were the most exposed for pollen collected from flowers and from *Apis* traps, pollen-nectar stores/ beebread collected in colonies and nests from the three bee species and honeybee foragers, whereas some the United Kingdom sites were the most exposed for nectar regurgitated from both bee species and bumblebee foragers (Fig. 3A right). In Estonian, Spanish and Swedish sites, pesticide were less found in the matrices in general. In some countries (Ireland, Italy and Sweden), the levels of exposure were highly variable, whereas in others (Estonia, Spain) the levels were homogeneous.

In the oilseed rape sites (Fig. 3B left), three bundles of variables were highlighted: (i) indices related to pollen-nectar stores/beebread and pollen from flowers, (ii) indices related to *Apis* and *Bombus* foragers, and (iii) indices related to nectar regurgitated from foragers and pollen from *Apis* traps. The indices were less correlated than indices from the apple orchard sites. In the oilseed rape sites, the indices related to nectar were correlated with each other ( $\text{cor} = 0.63$  for *Nectar.Apis* and *Nectar.Bombus*). The indices related to pollen-nectar stores/beebread (*Beebread.Apis*, *Pollen-nectar stores.Bombus* and *Pollen-nectar stores.Osmia*) were moderately correlated with each other ( $\text{cor} = 0.31$  to  $0.45$ ). These three indices related to pollen-nectar stores/beebread were also slightly correlated to the pollen collected from flowers ( $\text{cor} = 0.11$  with *Beebread.Apis*,  $\text{cor} = 0.23$  with *Pollen-nectar stores.Bombus* and  $\text{cor} = 0.41$  with *Pollen-nectar stores.Osmia*).

Italian sites, and to a lesser extent, the German, Spanish and Swiss sites contained the highest number of pesticides for pollen from flowers



and pollen-nectar stores/beebread. In Estonian and Irish sites the matrices contained the lowest number of pesticides in general (Fig. 3B right). In some countries (Germany and Sweden) the number of detected pesticides was highly variable whereas in some others (Italy and Spain), it was rather homogeneous.

#### 4. Discussion and conclusions

While several surveys have explored the presence of pesticides at the same time in different matrices (Ward *et al.*, 2022; Favaro *et al.*, 2019; Wen *et al.*, 2021), none proposed an index to characterise the exposure to pesticides. In this paper, we presented a highly novel statistical method using the IRT models to summarise complex information on pesticide presence into a single, yet interpretable, index.

##### 4.1. Indices from IRT models: strengths, adaptation and limits

This index illustrated the exposure to pesticides. It was more informative than a classic assessment of richness or abundance because it took into account the overall repartition of pesticides between samples together with quantities of pesticides. This index made possible the calculation of clusters based on similarity or dissimilarity of samples in terms of pesticide detection. As a consequence, comparison between sites (based on pesticide detection in the different samples collected in a given site) was possible.

Before choosing IRT models, different statistical methods were considered to reduce the complexity of the 18 datasets that originated from bee exposure to apple orchards and oilseed rape crops including the Multiple Correspondence Analysis (MCA) (Greenacre, 1984) applied on the overall distance matrix (Legendre and Legendre, 1998). Contrary to the indices summarising the exposure to infectious and parasitic agents (IPAs) (Huyen Ton *et al.*, 2023), the MCA was not adapted to deal with the multidimensionality of our data, as there was a very slow decay of eigenvalues due to the strong association between sites and pesticides. The proposed indices revealed a structure related to the number of pesticides detected on the sites, illustrated by the linear link between the number of pesticides detected and the exposure level of the sites (the index). The clustering of the sites based on the indices showed a clear separation between the clusters (Tables A.3 and A.4).

##### 4.2. Links between matrices and species

When designing the site network, one goal was to explore land-use management across countries and across agroecosystems, resulting in a gradient of exposure to pesticides (Hodge *et al.*, 2022). The land-use management data will be used in forthcoming statistical analyses. Eight countries from four biogeographic zones and two crops were included in the site network. The country of origin was not considered for the index calculation. However, this additional information was very useful to explain the different exposure levels at the sites. Applied to our dataset, the indices showed that in general, matrices collected in apple orchards contained a higher number of pesticides than matrices collected in oilseed rape crops. For a given matrix and a given country, different pesticides characterised the exposure at the sites according to crop exposure. These differences resulted from the crop treatments that were also different from country to country, most probably because of weather constraints and the blooming stage when sampling was performed. However, other factors may explain the diversity of pesticide uses across European countries such as the type of soils, the cultural habits and the commercial strategies from the pesticide industry.

In all cases, further statistical analysis is needed to compare the pesticide residue results to the real use of pesticides in the different countries. In other words, it would be worth investigating if, in the example of bees, the 1,2,3,6 tetrahydrophthalimide was more applied on apple orchards in the United Kingdom sites than in Estonian sites. Statistical analysis could focus on field treatments recorded during

PoshBee; and on the theoretical number of formulations with a market authorisation in these countries. To our knowledge, such comparison has never been made.

In general, the same countries had the most exposed (Germany and Italy) or the least exposed sites (Estonia, Spain) irrespective of the analysed matrix and the crop. However, there was some variation in pesticide detection between matrices for example between beebread collected in *Apis* bees and nectar regurgitated from *Apis* bees in oilseed rape sites located in Italy and Spain. These results show the difference of use and application of pesticides between European countries. This could be further explored with analyses including additional data on pesticide availability in the European countries. Our results also give first insights in the pathway of the contamination chain to understand the source and effect of pesticide residues on bees as aimed at by the site network (Hodge *et al.*, 2022). For a given site, all matrices contained similar number of pesticides but not necessarily by the same pesticides.

At apple orchard sites, the PCA highlighted the discrimination between pollen-nectar stores/beebread and pollen indices from nectar and bee indices. This separation was expected due to the high fat content of pollen-nectar stores/beebread and pollen and the high water content of nectar. This matrix discrimination was independent of country. To our surprise, the indices from the bee matrices (honeybees and bumblebees) were associated with the hydrophilic matrix (regurgitated nectars) rather than lipophilic matrix. It should be noted that this discrimination is based on pesticide numbers, as mentioned before. To further understand the matrix partition, it would be worth looking at the type of pesticides found in the sites, and checking if their chemical characteristics (lipophilicity, use of pKa) are in accordance with the discrimination of the matrices.

Consistently across bee species, sites were exposed at the same level for a given matrix. Some pesticides were in common, but in general the detected pesticides were different between the bee species. The three focal bee species selected in this study differ in foraging distances from <1 km for solitary bees (Zurbuchen *et al.*, 2010) up to 6 km for honeybees (Beekman and Ratnieks, 2000) and foraging preferences. Thus, they probably foraged to different extents on the two focal crops, other flowering crops and wild plants, contributing to different detected pesticide exposure levels. This question will be further explored with the palynological data analysis of pollen-nectar stores/beebread and published in future papers.

The number of samples collected from *Osmia* bees were either reduced (for the pollen-nectar stores) or absent (for the regurgitated nectar and for the bee bodies). This was an unfortunate side-effect of the ecology and biology of this species. If the difficulty to retrieve this matrix could be overcome, it would be worth examining the characteristics of pesticides (family, active ingredients and quantities) found in *Osmia* pollen-nectar stores compared to the ones found in pollen-nectar stores/beebread from the other two bee species.

Although there was a tendency for the UK, German, and Italian sites to be the most exposed and the Spanish and Estonian sites the least exposed, there were exceptions according to matrices. For example, sites located in Italy were the least exposed when looking at the pesticide residue presence in nectar regurgitated from *Apis* and *Bombus* foragers and pollen loads collected from *Apis* traps following oilseed rape exposure (Tables A.1 to A.4).

##### 4.3. Chemicals analysis as a key point to compare results on pesticide detection

The four laboratories involved in the analyses used different methods with large variation of screened pesticides depending on the extraction procedures and the analytical devices used (Kiljanek *et al.*, 2021; Martel *et al.*, 2023). Ring tests between the different analytical laboratories could be implemented to produce comparable results. This preliminary work should be taken into consideration in future surveys. Usually, stock standard solutions are used to calibrate the analytical devices, with

ready-to-use solutions containing several active ingredients. The non-availability of these stock standard solutions depending on the countries was a key point, preventing from having a common list of active ingredients screened for across the four laboratories. However, the list of 64 common pesticides to be screened in all the matrices defined before analyses enabled statistical comparisons when looking at analytical results. Many pesticides were included in the lists of screened pesticides and of those relatively few were found in the matrices – maximum 37 % in beebread collected from honeybee colonies (Table 1). These results show that more reflection should be made on targeting analyses to reduce the number of screened pesticides without impairing analytical relevancy. Indeed chemical analyses have potentially important economic and ecological costs.

#### 4.4. Risk posed by pesticide residue presence in various matrices

The IRT-based indices focused on bee exposure, not on risk assessment. However, considering the toxicity of detected pesticides is key for the assessment of pesticide risks for different bee species (Storck *et al.*, 2016) and is linked to the quantities of pesticides in the different matrices. The pesticides significant for discrimination (Table A.6) were mainly fungicides (70 % in matrices collected in apple orchard sites, and 43.4 % in those surrounded by apple). The proportion was the other way around for insecticides, more frequently found in apple orchard sites compared to oilseed rape. Being more toxic to bees, the exposure to insecticides puts bees more at risk than fungicide exposure. However, quantities and exposure scenarios are also important and should be integrated in the calculation of risk indicators. It would be interesting to explore whether the sites would be similarly clustered for pesticide risk, e.g., assessment based on hazard quotients (Favaro *et al.*, 2019; Wen *et al.*, 2021; Thomson, 2010; Rortais *et al.*, 2017) as regards to exposure, and if correlation between matrices would be similar. In other words, would the risk posed by pollen-nectar stores consumption to bumblebees be positively correlated to the risk posed by beebread consumption to honeybees? Such statistical work should be further explored. Another way to look at these data would be to explore the correlation between the cumulative concentrations of pesticides and the IRT-based indices for each site. If there was a correlation, we could discuss the notion of toxicity. It would be very interesting to have a comparison between cumulative concentrations and added toxic units such as toxicity-weighted concentration (Rundlöf *et al.*, 2022; Scientific Committee *et al.*, 2019).

Future studies could further assess whether pesticide residue exposure was related to bee population traits recorded in the field (Hodge *et al.*, 2022) along with further potential stressors of bee health (Breda *et al.*, 2022). In a previous study, we proposed an index calculation to summarise the exposure to IPAs (Huyen Ton *et al.*, 2023). The two kinds of indices (IPA and pesticide exposure) could be related to each other or used in structural modeling equations to understand the drivers of bee health. PoshBee data from the site network made it possible to assess pollinator development under field conditions, which is likely more informative for real world scenarios than tests conducted in laboratory conditions (Stanley and Raine, 2017). Comparing the pesticides found in the different matrices is also of importance and should be conducted in future statistical works.

To conclude, the index calculation based on the IRT methodology presented in this paper is reliable and offers many applications. The characterisation of sampling sites based on the number of detected pesticides across different matrices enabled us to summarise information from complex samples into a single and interpretable index. Our results show that although pesticide numbers were similar in matrices from any given country irrespective of bee species, some important variations could be observed. Therefore, for a complete assessment of pollinator pesticide exposure, it is necessary to consider several different exposure routes and multiple species of bees across different agricultural systems. Other parameters should be considered such as bee population traits,

different pesticide and application use between countries, other potential stressors of bee health. However all these information are usually lacking in field studies.

These results highlight the variation in the use and application of pesticides across European countries. This could be further explored with analyses including additional data on pesticide availability in the European countries. Our results also give first insights in the pathway of the contamination chain to understand the source and effect of pesticide residues on bees as aimed at by the site network (Hodge *et al.*, 2022). For a given site, all matrices experienced similar number of pesticides but not by the same pesticides or in comparable quantities.

Beyond such summarisation of complex data, the indices can be used in many ways, e.g. to compare and explore the correlation between matrices. Our datasets and matrices offer important opportunities for statistical analyses to examine relationships of the presented IRT indices with risks posed by pesticides to pollinators or their influence on bee health.

#### Funding

This research was funded by the European Horizon 2020 research and innovation program under grant agreement no.773921 and by the Anses.



A CC-BY public copyright licence

has been applied by the authors to the present document and will be applied to all subsequent versions up to the Author Accepted Manuscript arising from this submission, in accordance with the grant's open access conditions.

#### CRediT authorship contribution statement

**Marion LAURENT:** Writing – review & editing, Writing – original draft, Supervision, Funding acquisition, Data curation, Conceptualization. **Stéphanie BOUGEARD:** Writing – review & editing, Writing – original draft, Supervision, Resources, Methodology, Formal analysis, Data curation, Conceptualization. **Lucile CARADEC:** Writing – original draft, Methodology, Formal analysis, Data curation, Conceptualization. **Florence GHESTEM:** Writing – original draft, Methodology, Formal analysis, Data curation, Conceptualization. **Matthias ALBRECHT:** Writing – review & editing, Data curation. **Mark J.F. BROWN:** Writing – review & editing. **Joachim DE MIRANDA:** Data curation. **Reet KARISE:** Writing – review & editing, Data curation. **Jessica KNAPP:** Writing – review & editing, Data curation. **José SERRANO:** Writing – review & editing, Data curation. **Simon G. POTTS:** Writing – review & editing, Data curation. **Maj RUNDLÖF:** Data curation. **Janine SCHWARZ:** Data curation. **Eleanor ATTRIDGE:** Data curation. **Aurélié BABIN:** Writing – review & editing, Data curation. **Irene BOTTERO:** Data curation. **Elena CINI:** Data curation. **Pilar DE LA RÚA:** Data curation. **Gennaro DI PRISCO:** Data curation. **Christophe DOMINIK:** Data curation. **Andrés GARCÍA REINA:** Data curation. **Daniel DZUL:** Data curation. **Simon HODGE:** Writing – review & editing, Data curation. **Alexandra M. KLEIN:** Data curation. **Anina KNAUER:** Data curation. **Marika MAND:** Data curation. **Vicente MARTÍNEZ LÓPEZ:** Data curation. **Giorgia SERRA:** Methodology. **Helena PEREIRA-PEIXOTO:** Data curation. **Risto RAIMETS:** Data curation. **Oliver SCHWEIGER:** Data curation. **Deepa SENAPATHI:** Data curation. **Jane C. STOUT:** Data curation. **Giovanni TAMBURINI:** Data curation. **Cecilia COSTA:** Data curation. **Tomasz KILJANEK:** Writing – review & editing, Data curation. **Anne-Claire MARTEL:** Writing – review & editing, Data curation. **Sébastien LE:** Writing – review & editing, Writing – original draft, Visualization, Supervision, Methodology, Formal analysis, Data curation, Conceptualization. **Marie-Pierre CHAUZAT:** Writing – review & editing, Writing

– original draft, Validation, Supervision, Funding acquisition, Conceptualization.

## Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: Marie-Pierre CHAUZAT reports financial support was provided by European Commission. Marion LAURENT reports financial support was provided by European Commission. Matthias ALBRECHT reports financial support was provided by European Commission. Mark JF BROWN reports financial support was provided by European Commission. Joachim DE MIRANDA reports financial support was provided by European Commission. Reet KARISE reports financial support was provided by European Commission. Jessica KNAPP reports financial support was provided by European Commission. Jose SERRANO MARINO reports financial support was provided by European Commission. Simon G. POTTS reports financial support was provided by European Commission. Maj RUNDLOF reports financial support was provided by European Commission. Janine SCHWARZ reports financial support was provided by European Commission. Eleanor ATTRIDGE reports financial support was provided by European Commission. Irene BOTTERO reports financial support was provided by European Commission. Elena CINI reports financial support was provided by European Commission. Maria Pilar DE LA RUA reports financial support was provided by European Commission. Gennaro DI PRISCO reports financial support was provided by European Commission. Christophe DOMINIK reports financial support was provided by European Commission. Daniel DZUL reports financial support was provided by European Commission. Andres GARCIA REINA reports financial support was provided by European Commission. Simon HODGE reports financial support was provided by European Commission. Alexandra M KLEIN reports financial support was provided by European Commission. Anina KNAUER reports financial support was provided by European Commission. Marika MAND reports financial support was provided by European Commission. Vicente MARTINEZ LOPEZ reports financial support was provided by European Commission. Giorgia SERRA reports financial support was provided by European Commission. Helena PEREIRA-PEIXOTO reports financial support was provided by European Commission. Risto RAIMETS reports financial support was provided by European Commission. Oliver SCHWEIGER reports financial support was provided by European Commission. Deepa SENAPATHI reports financial support was provided by European Commission. Jane C. STOUT reports financial support was provided by European Commission. Giovanni TAMBURINI reports financial support was provided by European Commission. Cecilia COSTA reports financial support was provided by European Commission. Tomasz KILJANEK reports financial support was provided by European Commission. Anne-Claire MARTEL reports financial support was provided by European Commission. If there are other authors, they declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

we provided detailed indices in the supplementary material file

## Acknowledgements

This work would not have been possible without the tremendous work of the numerous persons involved at setting up the sampling sites network and at collecting, sending and analyzing all the samples. The authors would like to thank each of these collaborators. We thank Pensoft for help in designing the Fig. 3. This work has been performed in the framework of the European Reference Laboratory for Bee health.

## Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.scitotenv.2024.172118>.

## References

- Arena, M., Sgolastra, F., 2014. A meta-analysis comparing the sensitivity of bees to pesticides. *Ecotoxicology* 23, 324–334. <https://doi.org/10.1007/s10646-014-1190-1>.
- Beekman, M., Ratnieks, F.L.W., 2000. 2000 Long-range Foraging by the Honey-bee, *Apis mellifera* L. *Functional Ecology*.
- Bock, R.D., Aitkin, M., 1981. Marginal maximum likelihood estimation of item parameters: application of an EM algorithm. *Psychometrika* 46, 443–459.
- Breda, D., Frizzera, D., Giordano, G., Seffin, E., Zanni, V., Annoscia, D., Topping, C.J., Blanchini, F., Nazzi, F., 2022. A deeper understanding of system interactions can explain contradictory field results on pesticide impact on honey bees. *Nat. Commun.* 13, 5720. <https://doi.org/10.1038/s41467-022-33405-7>.
- CAPA, 2022. CAPA statement on Colony Losses in Canada, 24 pages. <https://capabees.com/capa-statement-on-honey-bees/>.
- Carter, L.J., Agatz, A., Kumar, A., Williams, M., 2020. Translocation of pharmaceuticals from wastewater into beehives. *Environ. Int.* 134, 105248 <https://doi.org/10.1016/j.envint.2019.105248>.
- Chalmer, R.P., 2012. Mirt: A multidimensional item response theory package for the R environment. *J. Stat. Softw.* 48, 1–29. <https://doi.org/10.18637/jss.v048.i06>.
- Demares, F.J., Schmehl, D., Bloomquist, J.R., Cabrera, A.R., Huang, Z.Y., Lau, P., Rangel, J., Sullivan, J., Xie, X., Ellis, J.D., 2022. Honey bee (*Apis mellifera*) exposure to pesticide residues in nectar and pollen in urban and suburban environments from four regions of the United States. *Environ. Toxicol. Chem.* 41, 991–1003. <https://doi.org/10.1002/etc.5298>.
- Di Pasquale, G., Alaux, C., Le Conte, Y., Odoux, J.F., Pioz, M., Vaissiere, B.E., Belzunces, L.P., Decourtaye, A., 2016. Variations in the availability of pollen resources affect honey bee health. *PLoS One* 11, e0162818. <https://doi.org/10.1371/journal.pone.0162818>.
- Everitt, B., 1974. *Cluster Analysis*. Heinemann Educ. Books, London.
- Favaro, R., Bauer, L.M., Rossi, M., D'Ambrosio, L., Bucher, E., Angeli, S., 2019. Botanical origin of pesticide residues in pollen loads collected by honeybees during and after apple bloom. *Front. Physiol.* 10, 1069. <https://doi.org/10.3389/fphys.2019.01069>.
- Graham, K.K., Milbrath, M.O., Zhang, Y., Baert, N., McArt, S., Isaacs, R., 2022. Pesticide risk to managed bees during blueberry pollination is primarily driven by off-farm exposures. *Sci. Rep.* 12, 7189. <https://doi.org/10.1038/s41598-022-11156-1>.
- Greenacre, M.J., 1984. *Theory and Applications of Correspondence Analysis*. Academic Press, London.
- Hodge, S., Stout, J., 2019. *Protocols for Methods of Field Sampling Deliverable D1.1*.
- Hodge, S., Schweiger, O., Klein, A.M., Potts, S.G., Costa, C., Albrecht, M., de Miranda, J. R., Mand, M., De la Rúa, P., Rundlöf, M., et al., 2022. Design and planning of a transdisciplinary investigation into farmland pollinators: rationale, co-design, and lessons learned. *Sustainability* 14, 10549. <https://doi.org/10.3390/su141710549>.
- Husson, F., Lê, S., Pages, J., 2017. *Exploratory Multivariate Analysis by Example Using R*. CRC press.
- Huyen Ton, Nu, Nguyen, M., Bougeard, S., Babin, A., Dubois, E., Druesne, C., Rivière, M. P., Laurent, M., Chauzat, M.P., 2023. Building composite indices in the age of big data – application to honey bee exposure to infectious and parasitic agents. *Heliyon* e15244. <https://doi.org/10.1016/j.heliyon.2023.e15244>.
- IPBES, Potts, S.G., Imperatriz-Fonseca, V.L., Ngo, H.T., Biesmeijer, J.C., Breeze, T.D., Dicks, L.V., Garibaldi, L.A., Hill, R., Settele, J., et al., 2016. Summary for Policymakers of the Assessment Report of the Intergovernmental Science-policy Platform on Biodiversity and Ecosystem Services on Pollinators, Pollination and Food Production. Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, Bonn, Germany.
- Jolliffe, I.T., 1986. *Principal Component Analysis*. Springer Verlag, New-York.
- Kavanagh, S., Henry, M., Stout, J.C., White, B., 2021. Neonicotinoid residues in honey from urban and rural environments. *Environ. Sci. Pollut. Res.* <https://doi.org/10.1007/s11356-021-12564-y>.
- Kiljanek, T., Niewiadomska, A., Malysiak, M., Posyniak, A., 2021. Miniaturized multiresidue method for determination of 267 pesticides, their metabolites and polychlorinated biphenyls in low mass beebread samples by liquid and gas chromatography coupled with tandem mass spectrometry. *Talanta* 235, 122721. <https://doi.org/10.1016/j.talanta.2021.122721>.
- Knapp, J.L., Nicholson, C.C., Jonsson, O., de Miranda, J.R., Rundlöf, M., 2023. Ecological traits interact with landscape context to determine bees' pesticide risk. *Nat. Ecol. Evol.* 7, 547–556. <https://doi.org/10.1038/s41559-023-01990-5>.
- Kulhanek, K., Steinhauer, N., Rennich, K., Caron, D.M., Sagili, R.R., Pettis, J.S., Ellis, J.D., Wilson, M.E., Wilkes, J.T., Tarpy, D.R., et al., 2017. A national survey of managed honey bee 2015–2016 annual colony losses in the USA. *J. Apic. Res.* 56, 328–340. <https://doi.org/10.1080/00218839.2017.1344496>.
- Lê, S., Josse, J., Husson, F., 2008. FactoMineR: an R package for multivariate analysis. *J. Stat. Softw.* 25, 1–18. <https://doi.org/10.18637/jss.v025.i01>.
- Legendre, P., Legendre, L., 1998. *Numerical Ecology*. Second ed ed. Elsevier, Amsterdam.
- Maggi, M., Antunez, K., Invernizzi, C., Aldea, P., Vargas, M., Negri, P., Brascato, C., de Jong, D., Message, D., Teixeira, E.W., et al., 2016. Honeybee health in South America. *Apidologie*. <https://doi.org/10.1007/s13592-016-0445-7>.
- Mahefarisoa, K.L., Simon Delso, N., Zaninotto, V., Colin, M.E., Bonmatin, J.M., 2021. The threat of veterinary medicinal products and biocides on pollinators: a one health



- perspective. *One Health*. 12, 100237 <https://doi.org/10.1016/j.onehlt.2021.100237>.
- Martel, A.C., Pierotti, N., Bray, E., 2023. Development and Validation of Two Multiresidue Methods for the Determination of Pesticides in Nectar Collected by Honey Bees and Bumble Bees (Submitted).
- Martinello, M., Manzinello, C., Borin, A., Avram, L.E., Dainese, N., Giuliano, I., Gallina, A., Mutinelli, F., 2020. A survey from 2015 to 2019 to investigate the occurrence of pesticide residues in dead honeybees and other matrices related to honeybee mortality incidents in Italy. *Diversity* 12. <https://doi.org/10.3390/d12010015>.
- McCune, F., Samson-Robert, O., Rondeau, S., Chagnon, M., Fournier, V., 2021. Supplying honey bees with waterers: a precautionary measure to reduce exposure to pesticides. *Environ. Sci. Pollut. Res.* 28, 17573–17586. <https://doi.org/10.1007/s11356-020-12147-3>.
- Moritz, R.F.A., Erler, S., 2016. Lost colonies found in a data mine: global honey trade but not pests or pesticides as a major cause of regional honeybee colony declines. *Agric. Ecosyst. Environ.* 216, 44–50. <https://doi.org/10.1016/j.agee.2015.09.027>.
- Mullin, C.A., Frazier, M., Frazier, J.L., Ashcraft, S., Simonds, R., Vanengelsdorp, D., Pettis, J.S., 2010. High levels of miticides and agrochemicals in North American apiaries: implications for honey bee health. *PLoS One* 5, e9754.
- Negri, I., Mavris, C., Di Prisco, G., Caprio, E., Pellicchia, M., 2015. Honey bees (*Apis mellifera*, L.) as active samplers of airborne particulate matter. *PLoS One* 10, e0132491. <https://doi.org/10.1371/journal.pone.0132491>.
- Oddie, M.A.Y., Burke, A., Dahle, B., Le Conte, Y., Mondet, F., Locke, B., 2021. Reproductive success of the parasitic mite (*Varroa destructor*) is lower in honeybee colonies that target infested cells with recapping. *Sci. Rep.* 11 <https://doi.org/10.1038/s41598-021-88592-y>.
- Ollerton, J., Erenler, H., Edwards, M., Crockett, R., 2014. Pollinator declines. *Extinctions of aculeate pollinators in Britain and the role of large-scale agricultural changes. Science* 346, 1360–1362 (doi:346/6215/1360).
- Pavlova, D., Atanassova, J., Karadjova, I., Bani, A., 2022. Pollen and chemical content of Beebreads from serpentine areas in Albania and Bulgaria. *Biol. Trace Elem. Res.* 200, 413–425. <https://doi.org/10.1007/s12011-021-02638-w>.
- Pirk, C.W., Human, H., Crewe, R.M., Vanengelsdorp, D., 2014. A survey of managed honey bee colony losses in the Republic of South Africa–2009 to 2011. *J. Apic. Res.* 53, 35–42. <https://doi.org/10.3896/IBRA.1.53.1.03>.
- Pochi, D., Biocca, M., Fanigliulo, R., Pulcini, P., Conte, E., 2012. Potential exposure of bees, *Apis mellifera* L., to particulate matter and pesticides derived from seed dressing during maize sowing. *Bull. Environ. Contam. Toxicol.* 89, 354–361.
- Potts, S.G., Biesmeijer, J.C., Kremen, C., Neumann, P., Schweiger, O., Kunin, W.E., 2010. Global pollinator declines: trends, impacts and drivers. *Trends Ecol. Evol.* 25, 345–353. <https://doi.org/10.1016/j.tree.2010.01.007>.
- Powney, G.D., Carvell, C., Edwards, M., Morris, R.K.A., Roy, H.E., Woodcock, B.A., Isaac, N.J.B., 2019. Widespread losses of pollinating insects in Britain. *Nat. Commun.* 10, 1018. <https://doi.org/10.1038/s41467-019-08974-9>.
- Pritchard, Z.A., Hendriksma, H.P., St Clair, A.L., Stein, D.S., Dolezal, A.G., O'Neal, M.E., Toth, A.L., 2021. Do viruses from managed honey bees (Hymenoptera: Apidae) endanger wild bees in native prairies? *Environ. Entomol.* <https://doi.org/10.1093/ee/nvaa181>.
- Raimets, R., Bontsutsnaja, A., Bartkevics, V., Pugajeva, I., Kaart, T., Puusepp, L., Pihlik, P., Keres, I., Viinalass, H., Mand, M., et al., 2020. Pesticide residues in beehive matrices are dependent on collection time and matrix type but independent of proportion of foraged oilseed rape and agricultural land in foraging territory. *Chemosphere* 238, 124555. <https://doi.org/10.1016/j.chemosphere.2019.124555>.
- Rollin, O., Pérez-Méndez, N., Bretagnolle, V., Henry, M., 2019. Preserving habitat quality at local and landscape scales increases wild bee diversity in intensive farming systems. *Agric. Ecosyst. Environ.* 275, 73–80. <https://doi.org/10.1016/j.agee.2019.01.012>.
- Rortais, A., Arnold, G., Dorne, J.L., More, S.J., Sperandio, G., Streissl, F., Szentes, C., Verdonck, F., 2017. Risk assessment of pesticides and other stressors in bees: principles, data gaps and perspectives from the European Food Safety Authority. *Sci. Total Environ.* <https://doi.org/10.1016/j.scitotenv.2016.09.127>.
- Rundlöf, M., Andersson, G.K., Bommarco, R., Fries, I., Hederstrom, V., Herbertsson, L., Jonsson, O., Klatt, B.K., Pedersen, T.R., Yourstone, J., et al., 2015. Seed coating with a neonicotinoid insecticide negatively affects wild bees. *Nature* 521, 77–80. <https://doi.org/10.1038/nature14420>.
- Rundlöf, M., Stuligross, C., Lindh, A., Malfi, R.L., Burns, K., Mola, J.M., Cibotti, S., Williams, N.M., 2022. Flower plantings support wild bee reproduction and may also mitigate pesticide exposure effects. *J. Appl. Ecol.* 59, 2117–2127. <https://doi.org/10.1111/1365-2664.14223>.
- Scientific Committee, E.F.S.A., More, S.J., Bampidis, V., Benford, D., Bennekou, S.H., Bragard, C., Halldorsson, T.I., Hernández-Jerez, A.F., Koutsoumanis, K., Naegeli, H., et al., 2019. Guidance on harmonised methodologies for human health, animal health and ecological risk assessment of combined exposure to multiple chemicals. *EFSA J.* 17, e05634 <https://doi.org/10.2903/j.efsa.2019.5634>.
- Serra, G., Costa, C., Cardaio, I., Colombo, R., 2021. Report on exposure of bees to agrochemicals. Deliverable D2, 2.
- Siviter, H., Richman, S.K., Muth, F., 2021. Field-realistic neonicotinoid exposure has sub-lethal effects on non-Apis bees: a meta-analysis. *Ecol. Lett.* 24, 2586–2597. <https://doi.org/10.1111/ele.13873>.
- Somme, L., Moquet, L., Quinet, M., Vanderplanck, M., Michez, D., Lognay, G., Jacquemart, A.L., 2016. Food in a row: urban trees offer valuable floral resources to pollinating insects. *Urban Ecosyst.* <https://doi.org/10.1007/s11252-016-0555-z>.
- Soroye, P., Newbold, T., Kerr, J., 2020. Climate change contributes to widespread declines among bumble bees across continents. *Science* 367, 685–688. <https://doi.org/10.1126/science.aax8591>.
- Stanley, D.A., Raine, N.E., 2017. Bumblebee colony development following chronic exposure to field-realistic levels of the neonicotinoid pesticide thiamethoxam under laboratory conditions. *Sci. Rep.* 7, 8005. <https://doi.org/10.1038/s41598-017-08752-x>.
- Storck, V., Karpouzias, D.G., Martin-Laurent, F., 2016. Towards a better pesticide policy for the European Union. *Sci. Total Environ.* <https://doi.org/10.1016/j.scitotenv.2016.09.167>.
- Straw, E.A., Brown, M.J.F., 2021. Co-formulant in a commercial fungicide product causes lethal and sub-lethal effects in bumble bees. *Sci. Rep.* 11, 21653 <https://doi.org/10.1038/s41598-021-00919-x>.
- Thomson, H.M., 2010. Risk assessment for honey bees and pesticides - recent developments and 'new issues'. *Pest Manag. Sci.* 66, 1157–1162.
- Traynor, K.S., Tosi, S., Rennich, K., Steinhauer, N., Forsgren, E., Rose, R., Kunkel, G., Madella, S., Lopez, D., Eversole, H., et al., 2021. Pesticides in honey bee colonies: establishing a baseline for real world exposure over seven years in the USA. *Environ. Pollut.* 279, 116566 <https://doi.org/10.1016/j.envpol.2021.116566>.
- Van der Linden, W.J., Hambleton, R.K., 1997. *Handbook of Modern Item Response Theory*. Springer, New-York, NY.
- van der Zee, R., Pisa, L., Andonov, S., Brodschneider, R., Charrière, J.-D., Chlebo, R., Coffey, M.F., Crailsheim, K., Dahle, B., Gajda, A., et al., 2012. Managed honey bee colony losses in Canada, China, Europe, Israel and Turkey, for the winters of 2008–9 and 2009–10. *J. Apic. Res.* 51, 100–114. <https://doi.org/10.3896/IBRA.1.51.1.12>.
- Ward, L.T., Hladik, M.L., Guzman, A., Winsemius, S., Bautista, A., Kremen, C., Mills, N.J., 2022. Pesticide exposure of wild bees and honey bees foraging from field border flowers in intensively managed agriculture areas. *Sci. Total Environ.* 831, 154697 <https://doi.org/10.1016/j.scitotenv.2022.154697>.
- Wen, X., Ma, C., Sun, M., Wang, Y., Xue, X., Chen, J., Song, W., Li-Byarlay, H., Luo, S., 2021. Pesticide residues in the pollen and nectar of oilseed rape (*Brassica napus* L.) and their potential risks to honey bees. *Sci. Total Environ.* 786, 147443 <https://doi.org/10.1016/j.scitotenv.2021.147443>.
- Wisk, J.D., Pistorius, J., Beevers, M., Bireley, R., Browning, Z., Chauzat, M.P., Nikolakis, A., Overmyer, J.P., Rose, R., Sebastien, R., et al., 2014. Assessing the exposure of pesticides to bees. In: *Pesticide Risk Assessment for Pollinators*. (ed. eds. D. Fischer, T. Moriarty). Wiley Blackwell, pp. 45–74.
- Zioga, E., Kelly, R., White, B., Stout, J.C., 2020. Plant protection product residues in plant pollen and nectar: a review of current knowledge. *Environ. Res.* 189, 109873 <https://doi.org/10.1016/j.envres.2020.109873>.
- Zurbuchen, A., Landert, L., Klaiber, J., Müller, A., Hein, S., Dorn, S., 2010. Maximum foraging ranges in solitary bees: only few individuals have the capability to cover long foraging distances. *Biol. Conserv.* 143, 669–676. <https://doi.org/10.1016/j.biocon.2009.12.003>.