

SUPPORTING INFORMATION

A Photo-Crosslinking Approach to Identify Class II SUMO-1 Binders

Kira Brüninghoff¹, Stephanie Wulff¹, Wolfgang Dörner¹, Ruth Geiss-Friedlander², Henning D. Mootz^{1*}

¹University of Münster, Institute of Biochemistry, Münster, Germany

²University of Freiburg, Institute of Molecular Medicine and Cell Research, Freiburg, Germany

Table of contents

Supporting Figures	page S2
Supporting Tables	page S7
Supporting References	page S11

SUPPORTING FIGURES

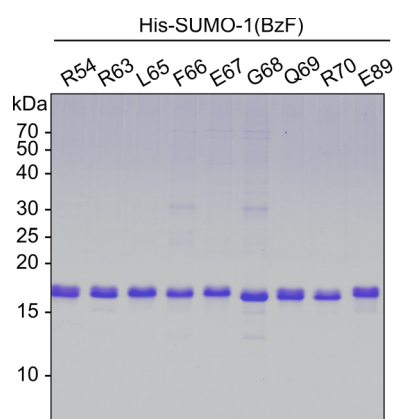
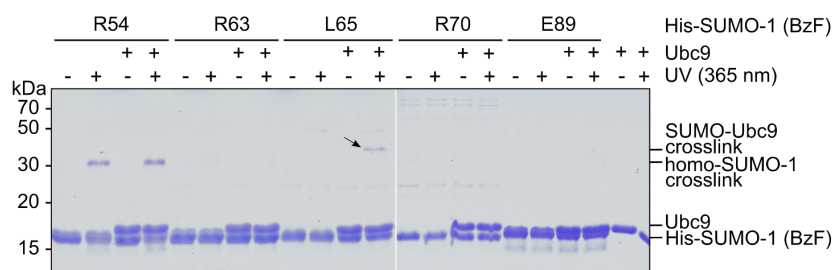


Figure S1. SUMO-1 mutants. SUMO-1 mutants with BzF at different positions were recombinantly expressed in *E.coli* and purified by N-terminal His₆-tag using Ni-NTA and size exclusion chromatography.

A



B

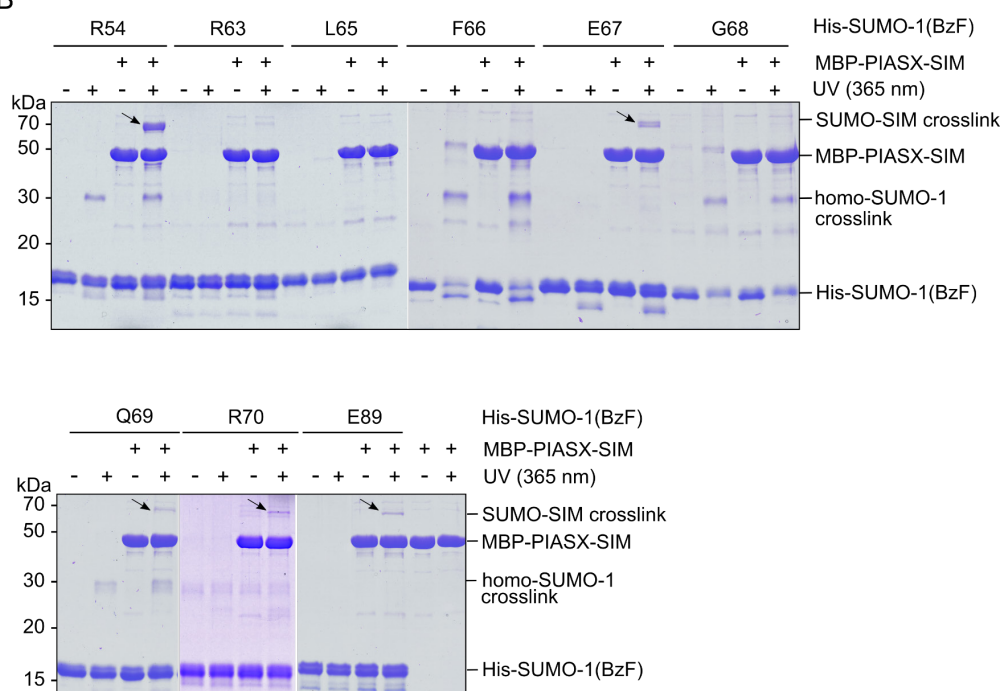
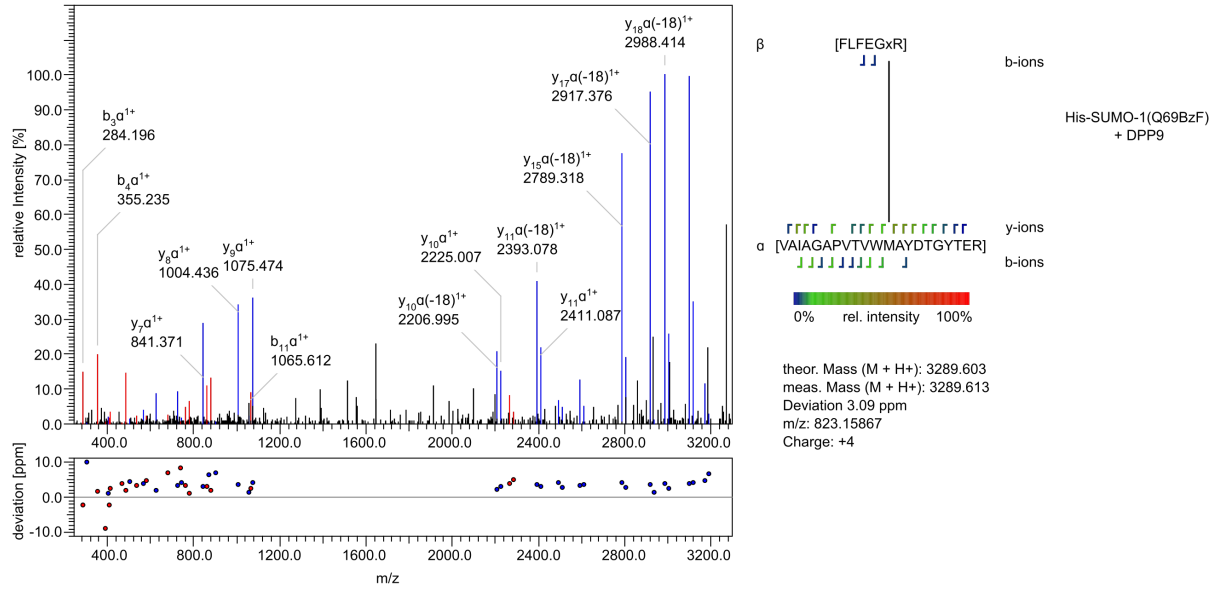


Figure S2. Photo-crosslinking of SUMO-1 mutants with Ubc9 (A) and MBP-PIASX-SIM (B). Proteins were incubated followed by UV-irradiated for 60 min. Samples before and after irradiation were analyzed by SDS-PAGE analysis and Coomassie staining.

A



B

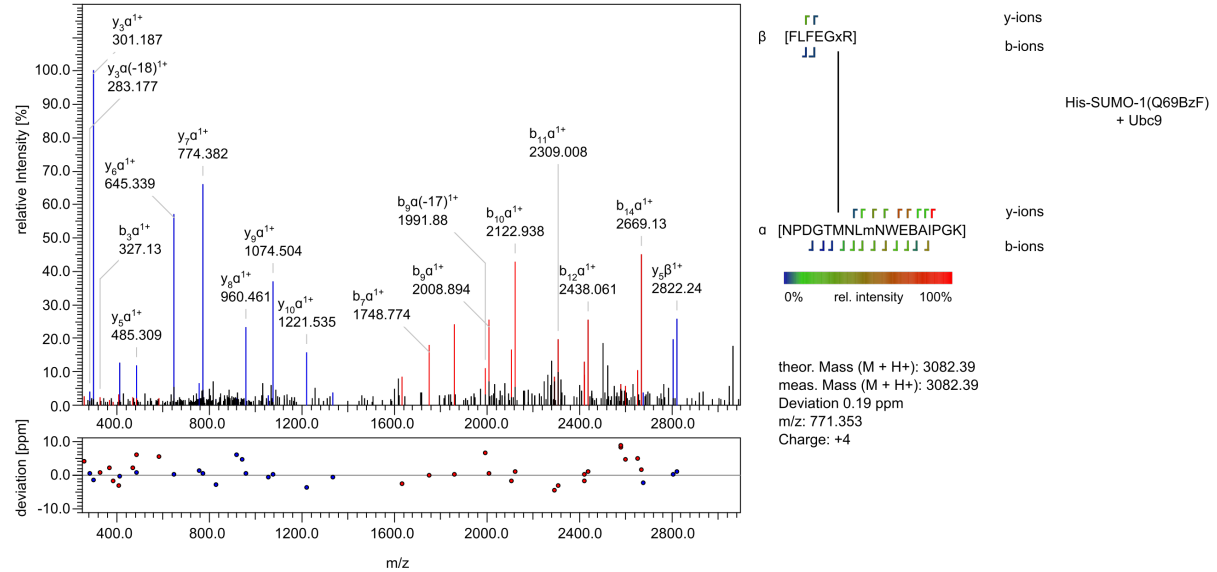
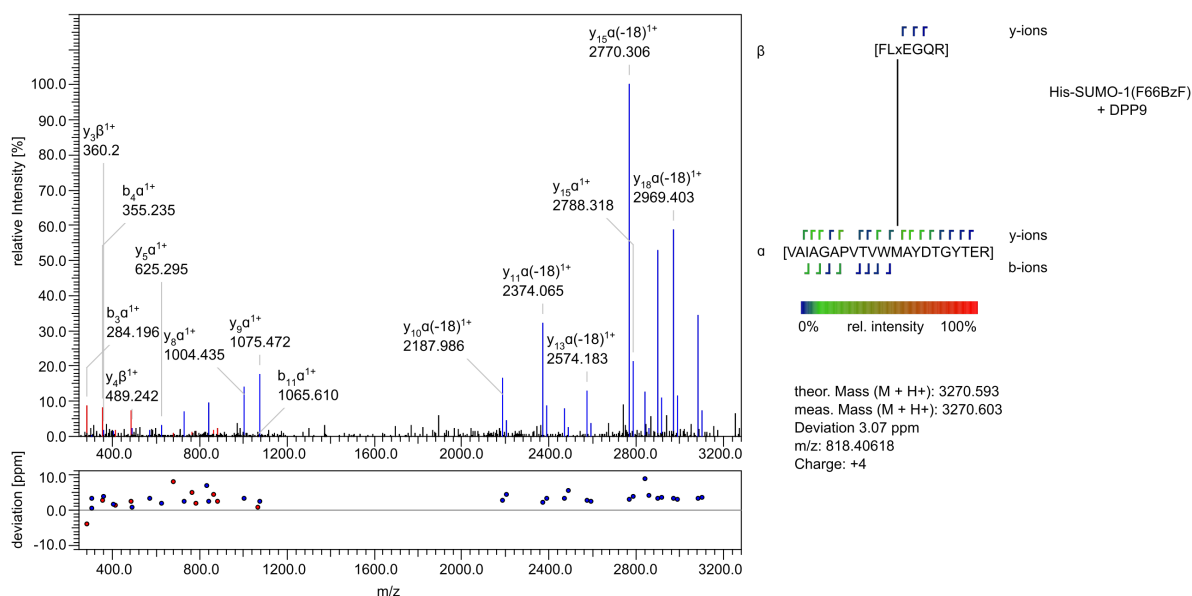


Figure S3. Analyses of photo-crosslinking products of His-SUMO-1(Q69BzF) with class II SUMO-1 binding partners DPP9 and Ubc9. (A) MS/MS spectra and fragmentation representation of the α - β crosslinked product of His-SUMO-1(Q69BzF) (β) and DPP9 (α) peptides. The precursor ion with a charge of +4 was measured at m/z 823.15867. The deviation of the singly charged precursor mass ($[M+H^+]$: 3289.613) from the theoretical singly charged crosslinked product ($[M+H^+]$: 3289.603) is 3.09 ppm. (B) MS/MS spectra and fragmentation representation of the α - β crosslinked product of His-SUMO-1(Q69BzF) (β) and Ubc9 (α) peptides. The precursor ion with a charge of +4 was measured at m/z 771.353. The deviation of the singly charged precursor mass ($[M+H^+]$: 3082.39) from the theoretical singly charged crosslinked product ($[M+H^+]$: 3082.39) is 0.19 ppm. The b- and y-type fragment ions of the peptide backbone are shown as red and blue peaks in the respective MS/MS spectra. Each of the depicted crosslink positions is one of the most likely positions hit by the photo-crosslinker, as can be determined from the accuracy of the data. ‘x’ represents photo-crosslinker BzF. ‘m’ indicates oxidized methionine.

A



B

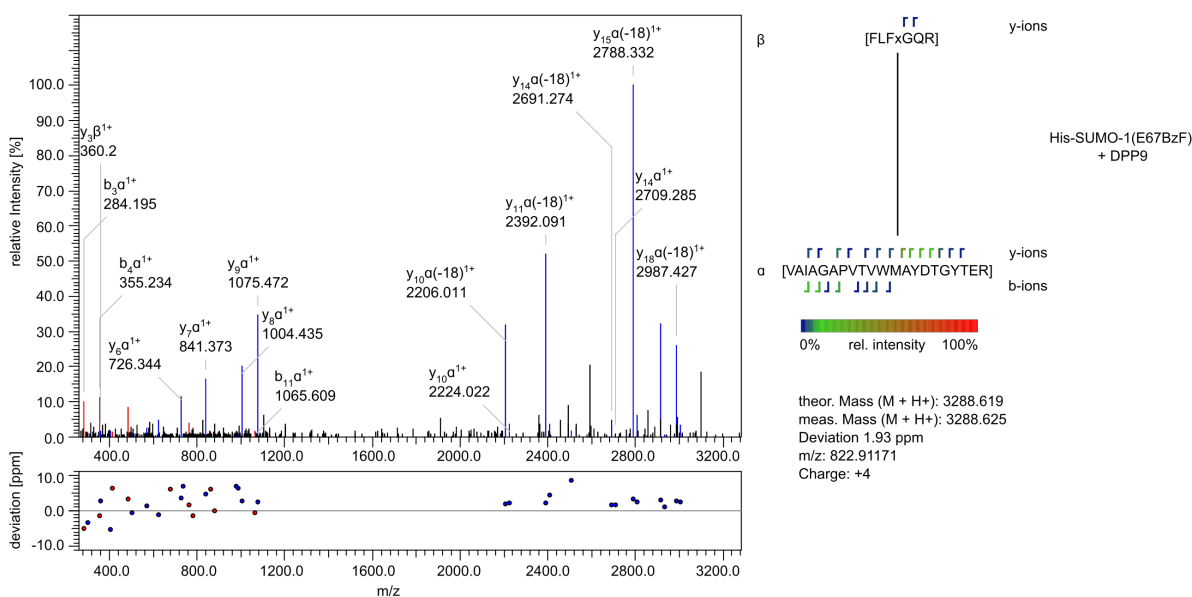


Figure S4. Analyses of photo-crosslinking products of His-SUMO-1(F66BzF) and His-SUMO-1(E67BzF) with DPP9. (A) MS/MS spectra and fragmentation representation of the α - β crosslinked product of His-SUMO-1(F66BzF) (β) and DPP9 (α) peptides. The precursor ion with a charge of +4 was measured at m/z 818.40618. The deviation of the singly charged precursor mass ($[M+H^+]$: 3270.603) from the theoretical singly charged crosslinked product ($[M+H^+]$: 3270.593) is 3.07 ppm. (B) MS/MS spectra and fragmentation representation of the α - β crosslinked product of His-SUMO-1(E67BzF) (β) and DPP9 (α) peptides. The precursor ion with a charge of +4 was measured at m/z 822.91171. The deviation of the singly charged precursor mass ($[M+H^+]$: 3288.625) from the theoretical singly charged crosslinked product ($[M+H^+]$: 3288.619) is 1.93 ppm. Each of the depicted crosslink positions is one of the most likely positions hit by the photo-crosslinker, as can be determined from the accuracy of the data. 'x' represents photo-crosslinker BzF.

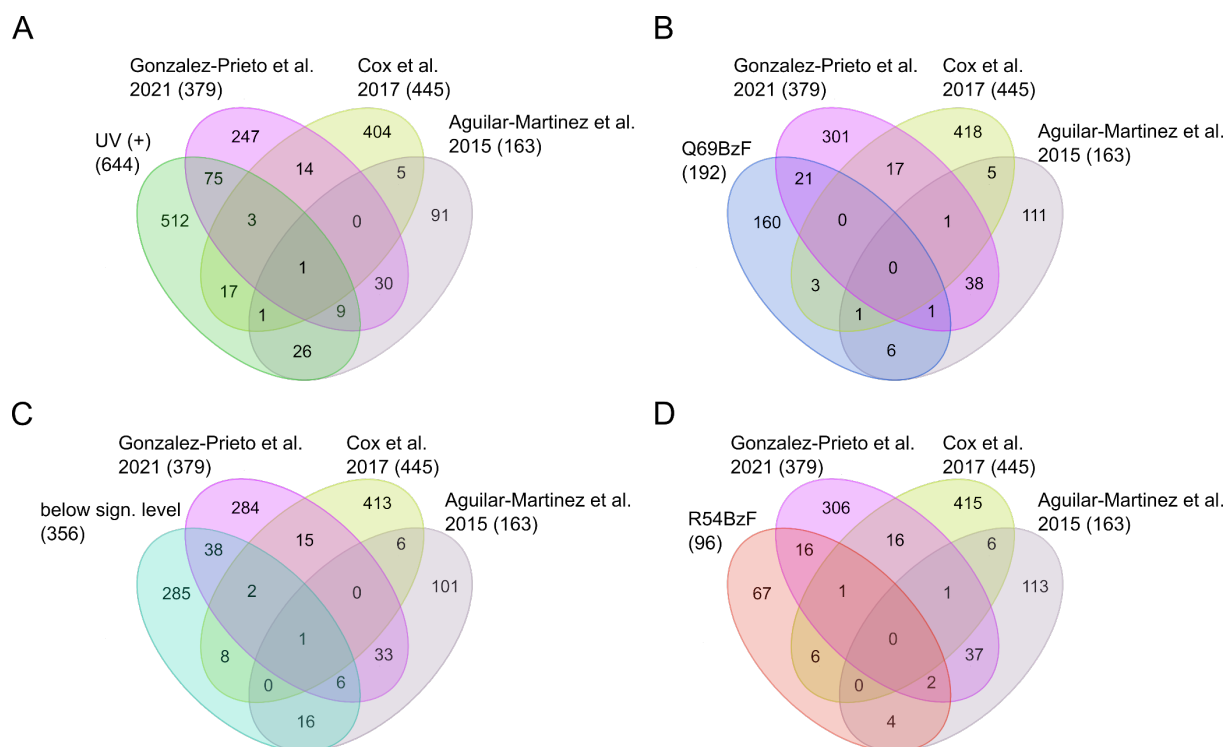


Figure S5. Comparison of all SUMO-binders identified in this study (A), proteins enriched by SUMO-1(Q69BzF) (B), proteins below the significance level (C) and proteins enriched by SUMO-1(R54BzF) (D) with SUMO interactors identified in the recent three major proteomic studies of (González-Prieto et al., 2021), (Cox et al., 2017) and (Aguilar-Martinez et al., 2015). For the comparison analyses, only proteins that could be unambiguously identified with Uniprot were considered.

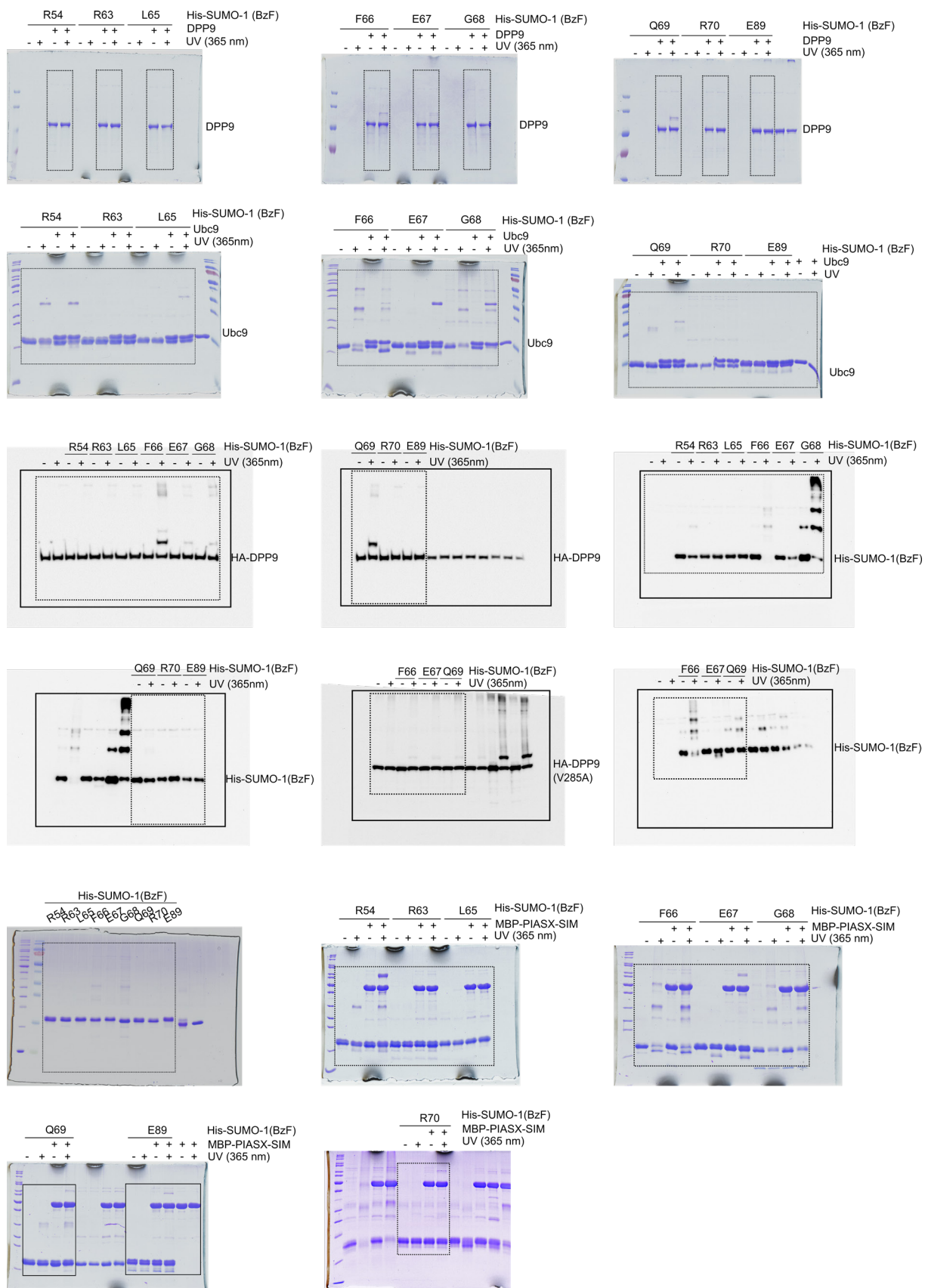


Figure S6. Uncropped images of SDS-gels and blots in Fig. 2, Fig. 3, Fig. S1, Fig. S2.

SUPPORTING TABLES

Tables S1-S5 (Excel sheets) are provided as separate Supporting Information file.

Table S6: List of proteins recombinantly expressed in *E.coli* and their expression plasmids.

Name of construct	Plasmid name	Vector backbone
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R54BzF)	pKBR42 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R63BzF)	pKBR105 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, L65BzF)	pKBR106 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, F66BzF)	pKBR111 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, E67BzF)	pKBR112 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, G68BzF)	pKBR113 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, Q69BzF)	pKBR110 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R70BzF)	pKBR107 + pEVOL(BzF)	pET28a
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, E89BzF)	pKBR108 + pEVOL(BzF)	pET28a
Ubc9	pUbc9	pET23a (provided by (Flotho et al., 2012))
MBP-PIASX-SIM-His ₆	pKT57	pMal-c2X

Table S7: Sequences of proteins used in this study.

Protein	Sequence
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R54BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQ[BzF]QGV MNSLRFLFEGQRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R63BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS L[BzF]FLFEGQRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, L65BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRF[BzF]FEGQRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, F66BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFL[BzF]EGQRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, E67BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFL[BzF]GQRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, G68BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFLFE[BzF]QRIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, Q69BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFLFEG[BzF]RIADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, R70BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFLFEGQ[BzF]IADNHTPKELGMEEEDVIEVYQEQ TGG
Met-Gly-Cys-Ser-His ₆ -SUMO-1(C52A, E89BzF)	MGCSHHHHHHGSDQEAKPSTEDLGDKKEGEYIKLK VIGQDSSEIHFKVKMTTHLKKLKESYAQRQGVPMNS LRFLFEGQRIADNHTPKELGMEEEDVI[BzF]VYQEQ TGG
Ubc9	MSGIALSRLAQRKAWRKDHPFGFVAVPTKNPDGT MNL MNWECAIPGKKGTPWEGGLFKLRMLFKDDYP SSPPKCKFEPPLFHPNVYPSGTVCLSILEEDKDWRPAI TIKQILLGIQELLNEPNIQDPAQAEAYTIYCQNRVEYE KRVRAQAKKFAPS
MBP-PIASX-SIM-His ₆	MKTEEGKLVIWINGDKGYNGLAEVGKKFEKDTGIK VTVEHPDKLEEKFPQVAATGDGPDIIIFWAHDRFGGY AQSGLLAEITPDKAFQDKLYPFTWDAVRYNGKLIAY PIAVEALSLIYNKDLLPNPPKTWEEIPALDKELKAKG KSALMFNLQEPYFTWPLIAADGGYAFKYENGKYDI KDVGVNAGAKAGLTFLVDLIK NKM NADTDYSIA

	EAAFNKGETAMTINGPWAWSNIDTSKVNYGVTVLP TFKGQPSKPFVGVLSAGINAASPNKELAKEFLENYLL TDEGLEAVNKDKPLGAVALKSYEEELAKDPRIAAT MENAQKGEIMPNIQMSAFWYAVRTAVINAASGRQ TVDEALKDAQTNSSSNNNNNNNNNNLGIEGRISEFL VPRGSKKVDVIDLTIESSSDEKVDHHHHHH
His ₆ -TEV-DPP9-S	MSYYHHHHHHHDYDIPTTENLYFQGAMGSATTGTPT ADRGDAAATDDPAARFQVQKHSWDGLRSIIHGSRK YSGLIVNKAPHDFQFVQKTDESGLPHSHRLYYLGMPY GSRENSLLYSEIPKKVRKEALLLSWKQMLDHFQAT PHHGVYSREEELLRERKRLGVFGITSYDFHSEGLFL FQASNSLFHCRDGGKNGFMVSPMKPLEIKTQCSGPR MDPKICPADPAFFSFINNSDLWVANIETGEERRLTFC HQGLSNVLDDPKSAGVATFVIQEEFDRFTGYWWCP TASWEGSEGLKTLRILYEEVDESEVEVIHVPSPALEE RKTDSYRYPRGTGSKNPKIALKLAEFQTDSQGKIVSTQ EKELVQPFSSLPKVEYIARAGWTRDGKYAWAMFL DRPQQWLQLVLLPPALFIPSTENEEQRLASARAVPR NVQPYVVYEEVTNVWINVHDIFYPPFQSEGEDELCE LRANECKTGFCCHLYKVTAVLKSQGYDWSEPFSPGE DEFKCPIKEEIALTSGEWEVLARHGSKIWVNEETKL VYFQGTKDTPLHHLYVVSYEAAAGEIVRLTTPGFSH SCSMSQNFDMMFVSHYSSVSTPPCVHVYKLSGPDDDP LHKQPRFWASMMEAASCPPDYVPPEIFHFHTRSDVR LYGMIYKPHALQPGKKHPTVLFVYGGPQVQLVNNS FKGIKYLRLNTLASLGYAVVVIDGRGSCQRGLRFEG ALKNQMGQVEIEDQVEGLQFVAEKYGFIDLSRVAIH GWAYGGFLSLMGLIHKPQVFKVAIAGAPVTVWMA YDTGYTERYMDVPENNQHGYEAGSVALHVEKLPN EPNRLILHGFLDENVHFFHTNFLVSQIRAGKPYQL QIYPNERHSIRCPESEGEHYEVTLLHFLQEYL
HA-DPP9	MASYPYDVPDYASLGSEFATTGTPTADRGDAAATD DPAARFQVQKHSWDGLRSIIHGSRKYSGLIVNKAPH DFQFVQKTDESGLPHSHRLYYLGMPYGSRENSLLYSE IPKKVRKEALLLSWKQMLDHFQATPHHGVYSREE ELLRERKRLGVFGITSYDFHSEGLFLFQASNSLFHC RDGGKNGFMVSPMKPLEIKTQCSGPRMDPKICPADP AFFSFINNSDLWVANIETGEERRLTFC HQGLSNVLDD PKSAGVATFVIQEEFDRFTGYWWCPTASWEGSEGL KTLRILYEEVDESEVEVIHVPSPALEERKTDSYRYPR TGSKNPKIALKLAEFQTDSQGKIVSTQEKELVQPFSS LFPKVEYIARAGWTRDGKYAWAMFLDRPQQWLQL VLLPPALFIPSTENEEQRLASARAVPRNVQPYVVYEE VTNVWINVHDIFYPPFQSEGEDELCELRANECKTGFC CHLYKVTAVLKSQGYDWSEPFSPGEDEFKCPIKEEIA LTSGEWEVLARHGSKIWVNEETKL VYFQGTKDTPL EHHL YVVS YEAAAGEIVRLTTPGFSHSCSMSQNFDMMF VSHYSSVSTPPCVHVYKLSGPDDDP LHKQPRFWAS MMEAASCPPDYVPPEIFHFHTRSDVR LYGMIYKPHA LQPGKKHPTVLFVYGGPQVQLVNNSFKGIKYLRLNT LASLGYAVVVIDGRGSCQRGLRFEGALKNQMGQVE

	IEDQVEGLQFVAEKYGFIDLSRVAIHGWSYGGFLSL MGLIHKPQVFKVAIAGAPVTVWMAYDTGYTERYM DVPENNQHGYESVALHVEKLPNEPNRLLILHGFL DENVHFFHTNFLVSQLIRAGKPYQLQIYPNERHSIRC PESGEHYEVTLHFLQEYL
HA-DPP9(V285A)	MASYPYDVPDYASLGSEFATTGTPTADRGDAAATD DPAARFQVQKHSWDGLRSIIHGSRKYSGLIVNKAPH DFQFVQKTDESGPHSHRLYYLGMPYGSRENSLLYSE IPKKVRKEALLLSWKQMLDHFQATPHHGVYSREE ELLRERKRLGVFGITSYDFHSESGFLFQASNSLFHC RDGGKNGFMVSPMKPLEIKTQCSGPRMDPKICPADP AFFSFINNSDLWVANETGEERLTFCHQGLSNVLLDD PKSAGVATFVIQEEFDRFTGYWWCPTASWEGSEGL KTLRILYEEVDESEAEVIHVPSPALEERKTDSYRYPR TGSKNPKIALKLAEFQTD SQGKIVSTQEKELVQPFSS LFPKVEYIARAGWTRDGKYAWAMFLDRPQQWLQL VLLPPALFIPSTENEEQRLASARAVPRNVQPYVVYEE VTNVWINVHDIFYPPQSEGEDEL CFLRANECKTGF CHLYKVTAVLKSQGYDWSEPFSPGEDEFKCPIKEEIA LTSGEWEVLARHGSKIWNNEETKL VYFQGT KDTP EHLHYVVS YEAAAGEIVRLTTPGFSHSCSMSQNFDMF VSHYSSVSTPPCVHVYKLSGPDDDPLHKQPRFWAS MMEAASCPDYVPPEIFHFHTRSDVRLYGM IYKPHA LQPGKKHPTVLFVYGGPQVQLVNNSFKGIKYLRLNT LASLGYAVVVIDGRGSCQRGLRFEGALKNQMGQVE IEDQVEGLQFVAEKYGFIDLSRVAIHGWSYGGFLSL MGLIHKPQVFKVAIAGAPVTVWMAYDTGYTERYM DVPENNQHGYESVALHVEKLPNEPNRLLILHGFL DENVHFFHTNFLVSQLIRAGKPYQLQIYPNERHSIRC PESGEHYEVTLHFLQEYL

Table S8: DPP9 plasmid information for expression in SF9 insect cells.

Name of construct	Plasmid name	Vector backbone
His ₆ -TEV-DPP9-short	Human DPP9-short	pFASTBacHT

Table S9: List of plasmids used for transfection.

Name of construct	Plasmid name	Vector backbone
HA-DPP9	pDPP9short	pcDNA3.1 (provided by (Pilla et al., 2012))
HA-DPP9(V285A)	pKBR119	pcDNA3.1

Supporting References

- Aguilar-Martinez, E., Chen, X., Webber, A., Mould, A. P., Seifert, A., Hay, R. T., et al. (2015). Screen for multi-SUMO-binding proteins reveals a multi-SIM-binding mechanism for recruitment of the transcriptional regulator ZMYM2 to chromatin. *Prot. Natl. Acad. Sci. U. S. A.* 112, E4854-63. doi: 10.1073/pnas.1509716112
- Cox, E., Hwang, W., Uzoma, I., Hu, J., Guzzo, C. M., Jeong, J., et al. (2017). Global Analysis of SUMO-Binding Proteins Identifies SUMOylation as a Key Regulator of the INO80 Chromatin Remodeling Complex. *Mol. Cell Proteomics* 16, 812–823. doi: 10.1074/mcp.M116.063719
- Flotho, A., Werner, A., Winter, T., Frank, A. S., Ehret, H., and Melchior, F. (2012). Recombinant reconstitution of sumoylation reactions in vitro. *Methods Mol. Biol.* 832, 93–110. doi: 10.1007/978-1-61779-474-2_5
- González-Prieto, R., Eifler-Olivi, K., Claessens, L. A., Willemstein, E., Xiao, Z., Talavera Ormeno, C. M. P., et al. (2021). Global non-covalent SUMO interaction networks reveal SUMO-dependent stabilization of the non-homologous end joining complex. *Cell Rep.* 34, 108691. doi: 10.1016/j.celrep.2021.108691
- Pilla, E., Möller, U., Sauer, G., Mattioli, F., Melchior, F., and Geiss-Friedlander, R. (2012). A novel SUMO1-specific interacting motif in dipeptidyl peptidase 9 (DPP9) that is important for enzymatic regulation. *J. Biol. Chem.* 287, 44320–44329. doi: 10.1074/jbc.M112.397224