

Supplemental information

**CD4⁺ T-cell-derived IL-10 promotes CNS inflammation
in mice by sustaining effector T cell survival**

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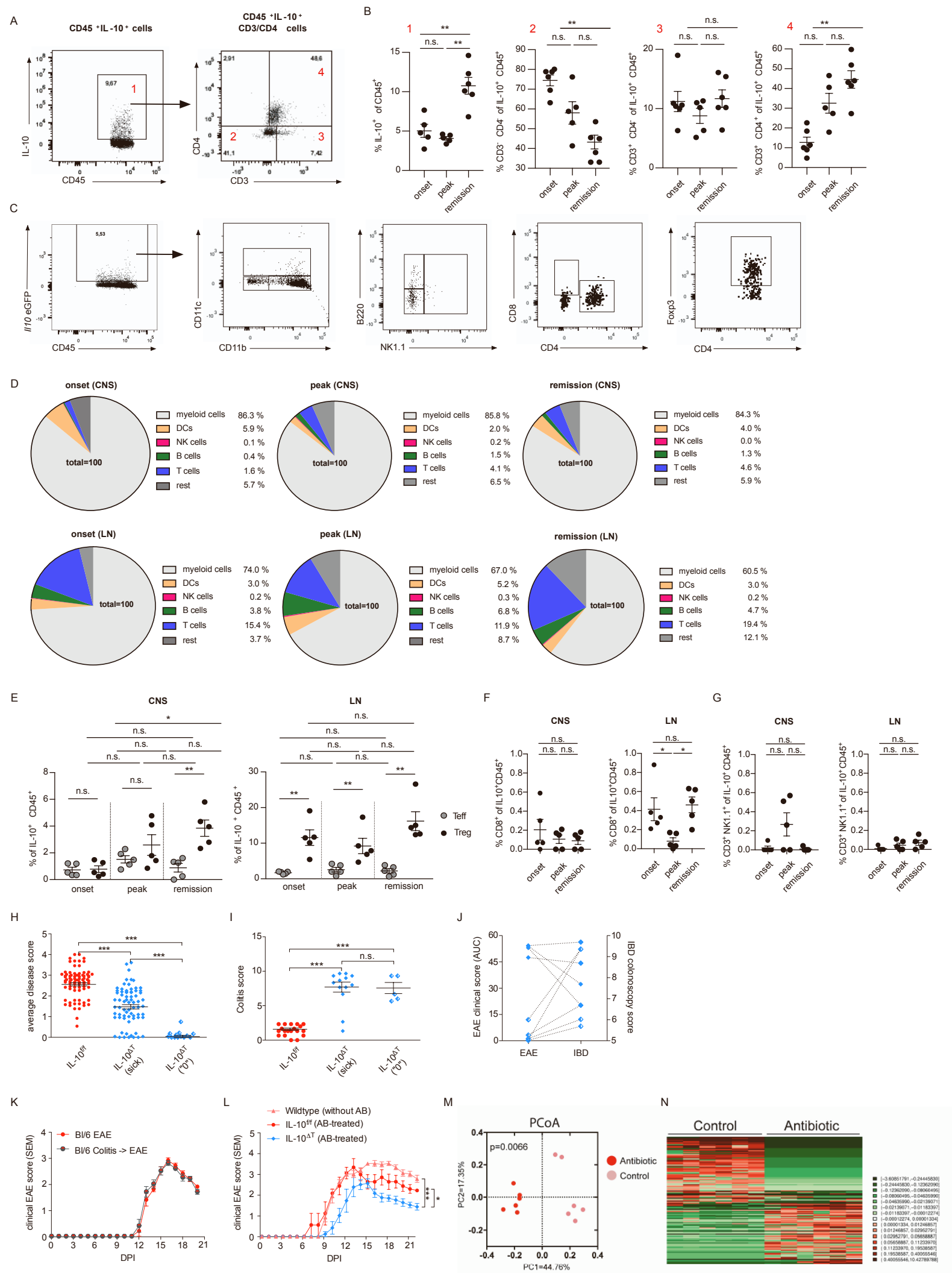


Figure S1 related to Figure 1: EAE mediated immune cell-derived IL-10 production and extra-intestinal inflammation.

(A, B) Lymphocytes were isolated from the CNS of wild-type mice with an EAE score of 1.5 (onset, d11), 3 (peak, d15), and 2.5 after (remission, d23) and restimulated with PMA/Ionomycin in the presence of Monensin for 5 hours prior antibody staining for CD45, CD3, CD4, and IL-10. (A) Representative Dotplots of IL-10⁺CD45⁺ cells (left) and IL-10⁺CD3⁺CD4⁺ (right) at disease remission. (B) Graphs summarize the percentage of CNS-infiltrating IL-10⁺CD45⁺ leukocytes (1) IL-10⁺CD3⁻CD4⁻ (2) IL-10⁺CD3⁺CD4⁻ (3), and IL-10⁺CD3⁺CD4⁺ (4) cells isolated from 5 mice per time-point. (C) Analysis of *Il10*eGFP expression by CD45⁺ leukocyte populations using the following gating strategy: CD45⁺IL-10⁺ cells (left) were subdivided into myeloid cells (CD11b⁺CD11c⁻); DCs (CD11c⁺CD11b^{+/+}); B cells (NK1.1⁻B220⁺); NK cells (B220⁻NK1.1⁺); CD8 T cells (CD4⁻CD8⁺); CD4⁺Foxp3⁺ (Treg), and CD4⁺Foxp3⁻ (Teff). (D-G) Graphs summarize the percentage of the analyzed IL-10⁺ cell populations isolated from the CNS and LN at the indicated time points. Data represent mean \pm SEM of $n = 5$ per condition obtained from 2 independent experiments. For statistical analysis Kruskal-Wallis was used. (H) EAE disease score in both *CD4^{Cre.neg}Il10^{fl/fl}* control (IL-10^{fl/fl}) and *CD4^{Cre.pos}Il10^{fl/f}* (IL-10 ^{Δ T}) mice. Both IL-10^{fl/fl} and IL-10 ^{Δ T} (sick) mice developed EAE score of ≥ 2 that lasted for two days or longer. IL-10 ^{Δ T} ("0") mice on the other hand, developed EAE score of ≤ 1 that lasted for two days or less. (I) Colitis disease score of individual IL-10^{fl/fl} and IL-10 ^{Δ T} animals. Representative animals were randomly selected among those analyzed for EAE development (shown in H). (J) EAE/Colitis correlative incidence analysis among representative individual IL-10 ^{Δ T} animals. (K) EAE disease curve of either C57BL/6 (BI/6 EAE) or C57BL/6 mice that were previously subjected to DSS-induced colitis (BI/6 Colitis \rightarrow EAE). (L) EAE disease curve in IL-10^{fl/fl} and IL-10 ^{Δ T} mice that have been maintained on broad-antibiotic treatment starting from the gestational stage and throughout their entire life span as well as control mice which has not received any antibiotic treatment. (K-L) Representative disease curve of ≥ 2 experiments, shown as mean \pm SEM of $n \geq 8$ per group. (M, N) Mice treated for three consecutive weeks with sulfadoxin-trimethoprim in the drinking water. As control, another group of mice did not receive

antibiotics. Fecal samples were collected three weeks after treatment initiation. The microbiota was analysed through 16s rRNA high-throughput sequencing. (M) Principal coordinate of analysis based on Bray-curtis distances at the level of amplicon sequence variants (ASV). The p value indicates that both groups have a significantly different microbiota composition ($p < 0.05$, PERMANOVA test). (N) Heatmap showing the amplicon sequence variants whose abundance significantly differs between both groups of mice (ANCOM2 & FDR, $q < 0.05$). For each ASV, the mean relative abundance was calculated. Then, for each sample, the original relative abundance was normalized as the difference respect to the mean of the corresponding ASV.

Supplementary Table 1 related to Figure 1: 16S rRNA sequence analysis of the gut microbiota.

ID (a)	p.value	q.value	mean.antibiotic	mean.control	log2FC	Taxonomy (b)
ASV0649	7,57E-05	0,000609696	0,012406567	0	3,291375294	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0637	0,002480471	0,011287939	0,015292912	0	3,565051029	p_Firmicutes;c_Clostridia;o_Clostridia_UCG-014;f_NA;g_NA
ASV0575	0,001003473	0,005626618	0,017464156	0	3,741349914	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0561	0,001889856	0,009484803	0,019237538	0	3,870898532	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Roseburia
ASV0533	0,000718979	0,004180729	0,025454047	0	4,250583315	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_UCG-005
ASV0482	0,004457614	0,015660086	0,029686752	0	4,461660538	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Marvinbryantia
ASV0461	0,004330967	0,015631305	0,031528034	0	4,54464753	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0429	0,004490906	0,015660086	0,037299171	0	4,777559555	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0422	0,001988225	0,00975473	0,05112576	0	5,218170506	p_Firmicutes;c_Bacilli;o_Erysipelotrichales;f_Erysipelotrichaceae;g_Erysipelotrichaceae
ASV0417	0,000234494	0,001636245	0,043646455	0	4,996610791	p_Actinobacteriota;c_Coribacteriales;o_Coribacteriales;f_Eggerthellaceae;g_DNF00809
ASV0391	0,006814125	0,021612478	0,061080381	0	5,468498889	p_Firmicutes;c_Clostridia;o_Clostridia_UCG-014;f_NA;g_NA
ASV0362	1,02E-06	9,66E-06	0,05686926	0	5,367849594	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0339	0,007768574	0,023541258	0,057557688	0	5,384791258	p_Firmicutes;c_Clostridia;o_Clostridia_vadinBB60_group;f_NA;g_NA
ASV0310	1,63E-07	1,60E-06	0,090830032	0	6,030247566	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0294	1,44E-06	1,33E-05	0,087732912	0	5,980975394	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Eisenbergiella
ASV0253	0,00754512	0,02325489	0,111220345	0	6,318374347	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0164	0,003732914	0,013754479	0,215015	0	7,260637523	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0160	0,004338433	0,015660086	0,035515939	0	4,709521237	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Tannerellaceae;g_Parabacteroides
ASV0119	0,002301862	0,010845437	0,330748852	0	7,878640552	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0113	0,00372137	0,01442605	0,31252184	0	7,797218891	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Blausia
ASV0111	8,37E-05	0,000654703	3,048870572	0,043093317	6,098841561	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NK4A136_group
ASV0335	2,86E-05	0,00023612	0,069020164	0,001142272	4,785593833	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0342	8,55E-05	0,000654703	0,072388365	0,001313444	4,785593833	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0177	0,000452542	0,003023368	0,221150123	0,004686677	5,189240182	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0247	0,006472642	0,020738873	0,124541921	0,003264645	4,751465764	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0293	0,000342706	0,002339342	0,1857243	0,008163817	4,288640243	p_Firmicutes;c_Clostridia;o_Peptococcales;f_Peptococcaceae;g_NA
ASV0206	0,005756858	0,018829723	0,077737892	0,003829217	3,916775925	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_GCA-900066575
ASV0279	0,010566415	0,030720874	0,08908268	0,004396106	3,961833658	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Ruminococcaceae;g_Incertae_Sedis
ASV0154	0,00066571	0,004019865	0,210838067	0,011509976	4,03793259	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0246	0,011449	0,032985124	0,111757041	0,00652929	3,833068722	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0350	0,000199169	0,001421345	0,052563055	0,003330251	5,508824142	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_A2
ASV0057	0,003046402	0,012926625	0,597396234	0,043473852	3,737780368	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0281	0,000613285	0,003775914	0,087703025	0,007515044	3,319506012	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0471	0,015873759	0,042601371	0,026713547	0,002689076	2,777996585	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0304	0,004207423	0,015542717	0,090591607	0,009517865	3,073488297	p_Firmicutes;c_Clostridia;o_Clostridia_UCG-014;f_NA;g_NA
ASV0260	0,001240795	0,006603554	0,096727477	0,011357192	2,942228428	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Roseburia
ASV0127	0,00162347	0,008414686	0,293225645	0,034946761	3,018588803	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0448	0,00216292	0,010448567	0,030210524	0,003935155	2,564250702	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Butyricicoccaceae;g_Butyricoccus
ASV0104	0,006337416	0,020514933	0,30602504	0,040193751	2,88545507	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Blausia
ASV0462	0,000770894	0,004180729	0,028509899	0,003897632	2,494659145	p_Firmicutes;c_Bacilli;o_RF39;f_NA;g_NA
ASV0142	0,007797105	0,023541258	0,239293075	0,034888042	2,79244759	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0151	0,004666372	0,015755278	0,21695407	0,037538951	2,48703747	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0244	0,001903002	0,009484803	0,097818689	0,020008981	2,211795149	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0123	0,015025635	0,010426516	0,244374897	0,052143188	2,198323067	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_UCG-003
ASV0303	0,002440659	0,011270103	0,064306145	0,013754917	2,115410141	p_Firmicutes;c_Clostridia;o_Peptostreptococcales-Tissierellales;f_Anaerovaccaceae;g_Anaerovaccaceae
ASV0117	0,000873283	0,004895652	0,267487384	0,057951394	2,179432918	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0148	0,002987451	0,012850132	0,218434791	0,049732437	2,103862919	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0137	0,000608576	0,003775914	0,20230646	0,051443368	1,946467576	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Tannerellaceae;g_Parabacteroides
ASV0456	0,004038044	0,015094591	0,026170262	0,006681437	1,768995992	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Tannerellaceae;g_Parabacteroides
ASV0157	0,012496853	0,035351457	0,193180481	0,053519437	1,824763915	p_Firmicutes;c_Bacilli;o_Lactobacillales;f_Lactobacillaceae;g_Lactobacillus
ASV0037	0,013308835	0,036982072	0,859907125	0,263941528	1,698634511	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_NA
ASV0091	0,00402165	0,015094591	0,309401988	0,10069269	1,606008282	p_Desulfobacterota;c_Desulfobacteriales;o_Desulfobacteriales;f_Desulfobacteriales
ASV0080	0,002314154	0,010845437	0,340540412	0,112126364	1,590621063	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0068	0,010292845	0,030205172	0,413386939	0,143062099	1,521605438	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0005	0,0036528	0,01442605	6,378196003	2,27773613	1,484973343	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Rikenellaceae;g_Alistipes
ASV0042	0,003308357	0,01366874	0,657882641	0,24813828	1,401620841	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_Oscillibacter
ASV0128	0,009921734	0,029390798	0,214393442	0,081513714	1,379846572	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Oscillospiraceae;g_Colidextribacter
ASV0009	0,018101709	0,047764172	3,086078087	1,726080655	0,83775516	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0102	0,004646881	0,016183295	0,119942673	0,249296331	-1,046784144	p_Firmicutes;c_Clostridia;o_Oscillospirales;f_Ruminococcaceae;g_Incertae_Sedis
ASV0035	0,004607727	0,015726273	0,346577473	0,807778858	-1,217437848	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0507	0,01625037	0,035395193	0,191387621	0,564850014	-1,554383397	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Marinifilaceae;g_Odoribacter
ASV0192	0,01156511	0,030313132	0,041457496	0,13378653	-1,673960872	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0044	0,001473394	0,00771076	0,178654404	0,634645363	-1,82063147	p_Firmicutes;c_Bacilli;o_Erysipelotrichales;f_Erysipelotrichaceae;g_Turicibacter
ASV0010	0,008463878	0,025311026	0,843027998	3,017416554	-1,837922782	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Prevotellaceae;g_Prevotellaceae
ASV0322	0,016872291	0,048497541	0,014718203	0,056514376	-1,844496609	p_Cyanobacteria;c_Vampirivibrionia;o_Gastranaerophilales;f_NA;g_NA
ASV0132	0,003195773	0,013379635	0,049675498	0,233042767	-2,198281376	p_Firmicutes;c_Bacilli;o_Lactobacillales;f_Lactobacillaceae;g_Lactobacillus
ASV0141	0,003914722	0,014990521	0,041506977	0,20614122	-2,273810548	p_Firmicutes;c_Bacilli;o_Lactobacillales;f_Lactobacillaceae;g_Lactobacillus
ASV0287	0,015593196	0,042209168	0,004107804	0,076756673	-3,824055034	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0282	0,013672699	0,037659889	0,001680465	0,075727335	-4,640920193	p_Firmicutes;c_Clostridia;o_Clostridia_vadinBB60_group;f_NA;g_NA
ASV0677	0,000561041	0,003959241	0	0,010309926	-3,053965004	p_Firmicutes;c_Clostridia;o_Clostridiales;f_Clostridiaceae;g_Clostridium_sensu_stricto_1
ASV0650	0,002623888	0,01177001	0	0,012512342	-3,032376904	p_Firmicutes;c_Bacilli;o_Erysipelotrichales;f_Erysipelotrichaceae;g_NA
ASV0640	0,002669594	0,018103544	0	0,0121331	-3,262537001	p_Firmicutes;c_Clostridia;o_Clostridia_vadinBB60_group;f_NA;g_NA
ASV0576	0,001148821	0,006219477	0	0,01684567	-3,693285945	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0564	0,002906146	0,012674026	0	0,018081169	-3,787755058	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0554	0,007086522	0,022512565	0	0,02012565	-3,914450662	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Bacteroidaceae;g_Bacteroides
ASV0505	0,004308264	0,015631305	0	0,023667947	-4,151331138	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_Lachnospiraceae
ASV0493	3,85E-09	4,64E-08	0	0,024987079	-4,225286242	p_Firmicutes;c_Clostridia;o_Peptostreptococcales-Tissierellales;f_Anaerovaccaceae;g_NA
ASV0467	0,003694768	0,01442605	0	0,029275768	-4,44267202	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_A2
ASV0453	0,003430602	0,013810371	0	0,030101097	-4,480755791	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0385	9,51E-05	0,000711013	0	0,042781413	-4,968643907	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0272	5,72E-11	1,38E-09	0	0,092839397	-6,062173013	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0270	0,000158915	2,56E-08	0	0,092719804	-6,05055718	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0269	2,75E-14	4,32E-12	0	0,094002049	-6,079023034	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0267	0,007554136	0,02325489	0	0,097877592	-6,136443219	p_Firmicutes;c_Clostridia;o_Lachnospirales;f_Lachnospiraceae;g_NA
ASV0232	4,77E-09	5,54E-08	0	0,118716211	-6,411329854	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0216	4,19E-12	1,88E-10	0	0,13749952	-6,620920937	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Bacteroidaceae;g_Bacteroides
ASV0205	1,75E-09	2,29E-08	0	0,146218593	-6,70874679	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0202	0,001120937	0,006174984	0	0,1612099	-6,848277197	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0201	2,73E-12	1,43E-10	0	0,157006559	-6,810496855	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0189	5,92E-10	9,78E-09	0	0,16423091	-6,874832201	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0187	2,20E-08	2,30E-07	0	0,177603955	-6,986844332	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Sutterellaceae;g_Parasutterella
ASV0178	6,74E-10	1,06E-08	0	0,182803516	-7,028150879	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0176	5,25E-10	9,16E-09	0	0,191678679	-7,096035051	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0174	2,51E-06	2,25E-05	0	0,105404758	-6,241890712	p_Proteobacteria;c_Gammaproteobacteria;o_Burkholderiales;f_Sutterellaceae;g_Parasutterella
ASV0173	1,67E-10	3,08E-09	0	0,194248103	-7,101623618	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0167	0,00054269	0,003550095	0	0,215681766	-7,265075348	p_Firmicutes;c_Clostridia;o_Clostridia_UCG-014;f_NA;g_NA
ASV0161	1,08E-09	1,55E-08	0	0,204765428	-7,190643225	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0159	1,41E-08	1,53E-07	0	0,215499172	-7,263861404	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0150	2,07E-05	0,000175632	0	0,233153449	-7,376747881	p_Bacteroidota;c_Bacteroidia;o_Bacteroidales;f_Muribaculaceae;g_NA
ASV0139	7,92E-11	1,78E-09	0	0,2566		

Supplementary Table 1 related to Figure 1: 16S rRNA sequence analysis of the gut microbiota.

Taxonomy and q values obtained for each amplicon sequence variants (ASV). For each ASV, the mean relative abundance was calculated. Then, for each sample, the original relative abundance was normalized as the difference respect to the mean of the corresponding ASV. The obtained value represented with a colour as depicted. The taxonomy and q values obtained for each ASV are shown.

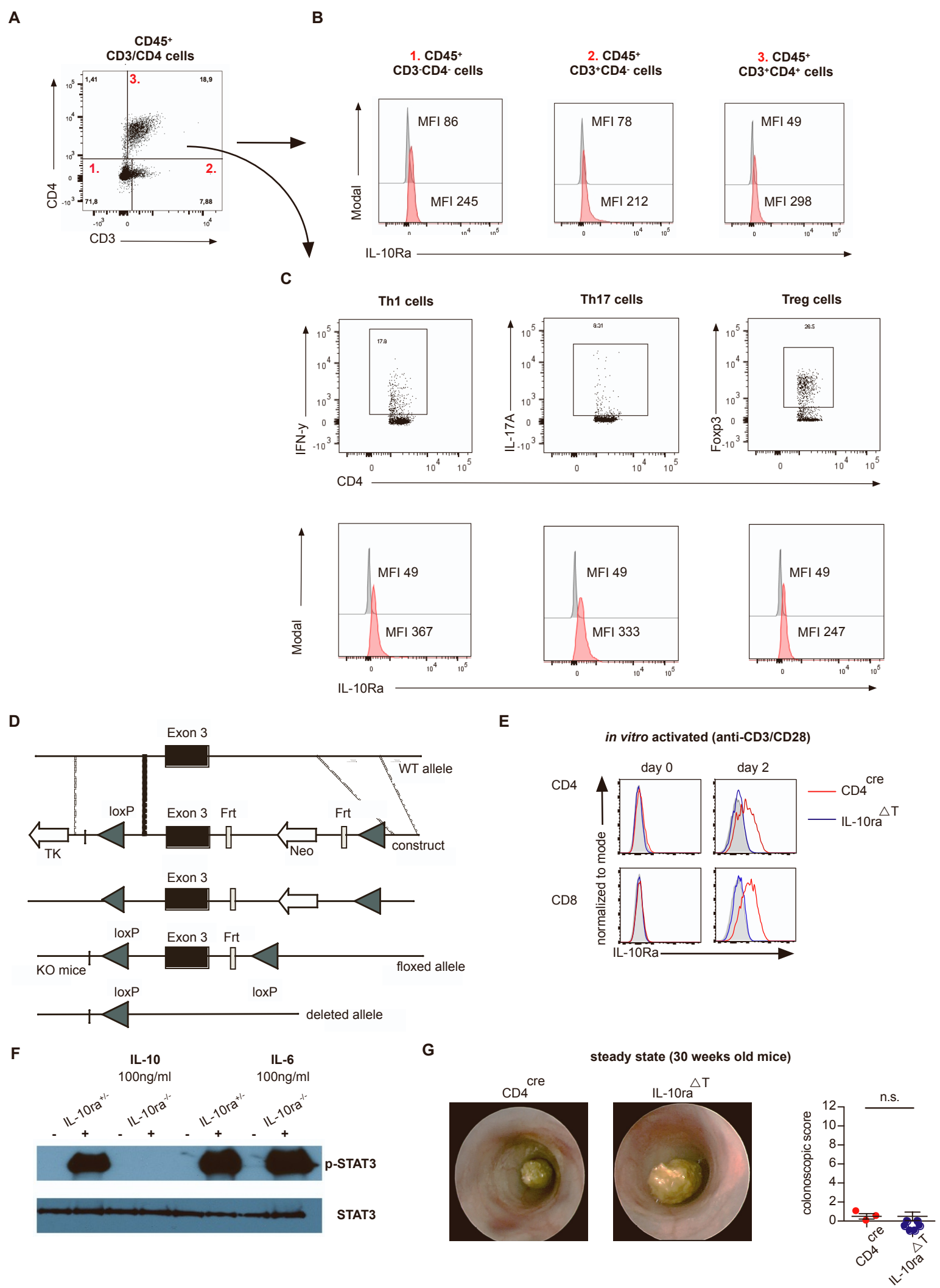


Figure S2

Figure S2 related to Figure 2: Gating strategy for the identification of IL10Ra expressing immune cells and characterization of $CD4^{Cre}Il10ra^{fl/fl}$ mice.

(A) Gating strategy of IL10 receptor expression: $CD45^{+}$ cells (not shown) were first subdivided into $CD3^{-}CD4^{-}$, $CD3^{+}CD4^{-}$, and $CD3^{+}CD4^{+}$ cell subsets. (B) Histograms show FMO for IL-10R staining (grey) of the indicated subset overlaid with IL-10R staining (red) of the same subset. (C) Gating of *Ifng* FP635⁺ Th1 cells, *Il17a* eGFP⁺ Th17 cells and *Foxp3* RFP⁺ Treg cells from $CD3^{+}CD4^{+}$ (as shown in A). lower Panel: Histograms show FMO for IL-10R staining (grey) of $CD4^{+}$ T cells overlaid with IL-10R staining (red) of the indicated subset. (D) *Il10ra* construct used for the generation of $CD4^{Cre}Il10ra^{fl/fl}$ mice. (E) IL-10R α expression on $CD4^{+}$ T cells and $CD8^{+}$ T cells directly after isolation from spleens of untreated $CD4^{Cre}Il10ra^{wt/wt}$ ($CD4^{Cre}$) and $CD4^{Cre}Il10ra^{fl/fl}$ (IL-10R $\alpha^{\Delta T}$) mice and after 2 days of *in vitro* activation in the presence of 1 μ g/ml soluble anti-CD3 and anti-CD28 mAb. (F) Western blot of STAT3 phosphorylation of $CD4^{+}$ T cells isolated from heterozygous $CD4^{Cre}Il10ra^{fl/wt}$ and $CD4^{Cre}Il10ra^{fl/fl}$ (IL-10R $\alpha^{\Delta T}$) mice after *in vitro* stimulation with 100 ng/ml IL-10 or 100 ng/ml IL-6 as a positive control, respectively. (G) Endoscopy of 30 weeks old untreated $CD4^{Cre}$ and IL-10R $\alpha^{\Delta T}$ mice as described in material and methods.

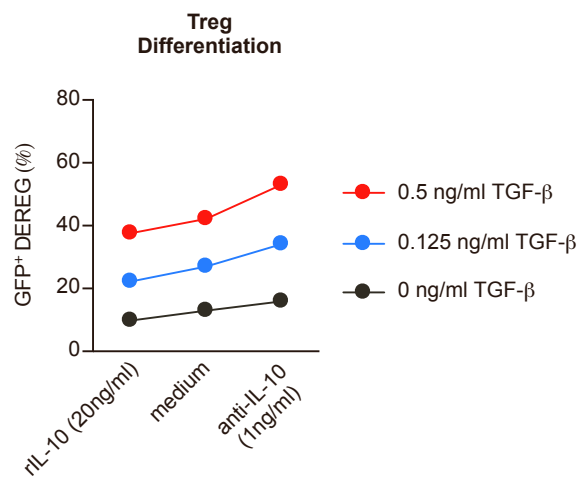
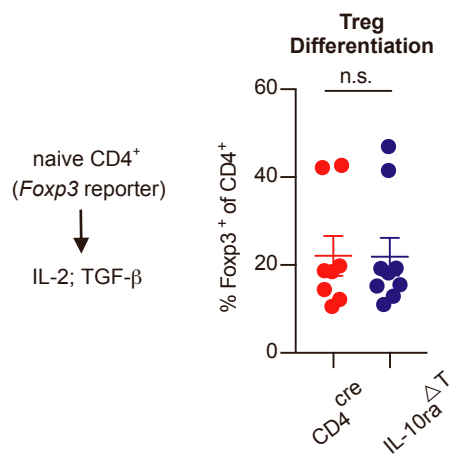
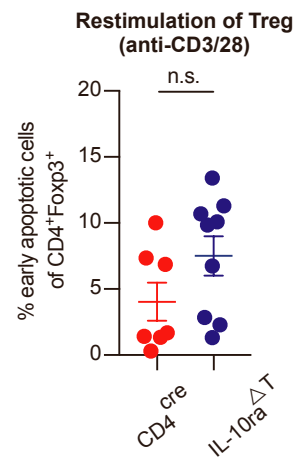
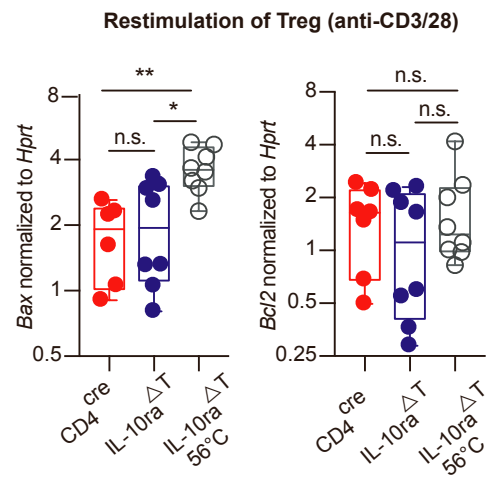
A**B****C****D**

Figure S3

Figure S3 related to Figure 3: IL-10 signaling in Treg is dispensable for their survival *in vitro*.

(A) FACSsorted naïve CD4 T cells, isolated from the FoxP3-GFP reporter mice were plated on 96 well plate (2×10^5 /well, triplicates) in serum free/TGF β -free medium. Cells were stimulated with anti CD3/CD28, TGF β at the indicated concentrations, and treated with either rIL-10 (20ng/ml), anti-IL-10 (1 μ g/ml) or without further stimuli. 72hrs post stimulation, the triplicates were pooled, stained and analyzed by FACS. Data shown as percentages of GFP $^+$ iTreg of total viable CD4 $^+$ T cells. One representative experiment of two. (B) Flow cytometric analysis of the frequencies of induced *Foxp3* RFP $^+$ Treg after 5 days of *in vitro* differentiation from naïve IL10R $\alpha^{\Delta T}$ T cells (blue; n=9) and CD4 Cre CD4 $^+$ T cells (red; n=8). (C) FACS analysis of early apoptotic Annexin $^+$ Pacific Orange $^-$ iTreg (left) and (D) mRNA expression levels of the pro-apoptotic gene *Bax* and anti-apoptotic gene *Bcl2* in sorted IL-10R $\alpha^{\Delta T}$ and CD4 Cre iTreg after 16hr (for FACS analysis) and 12 hrs (for qPcR analysis) of restimulation with plate-bound 1 μ g/ml anti-CD3 and soluble anti-CD28 mAb. Grey bars show gene expression after heat induced apoptosis induction as positive control.

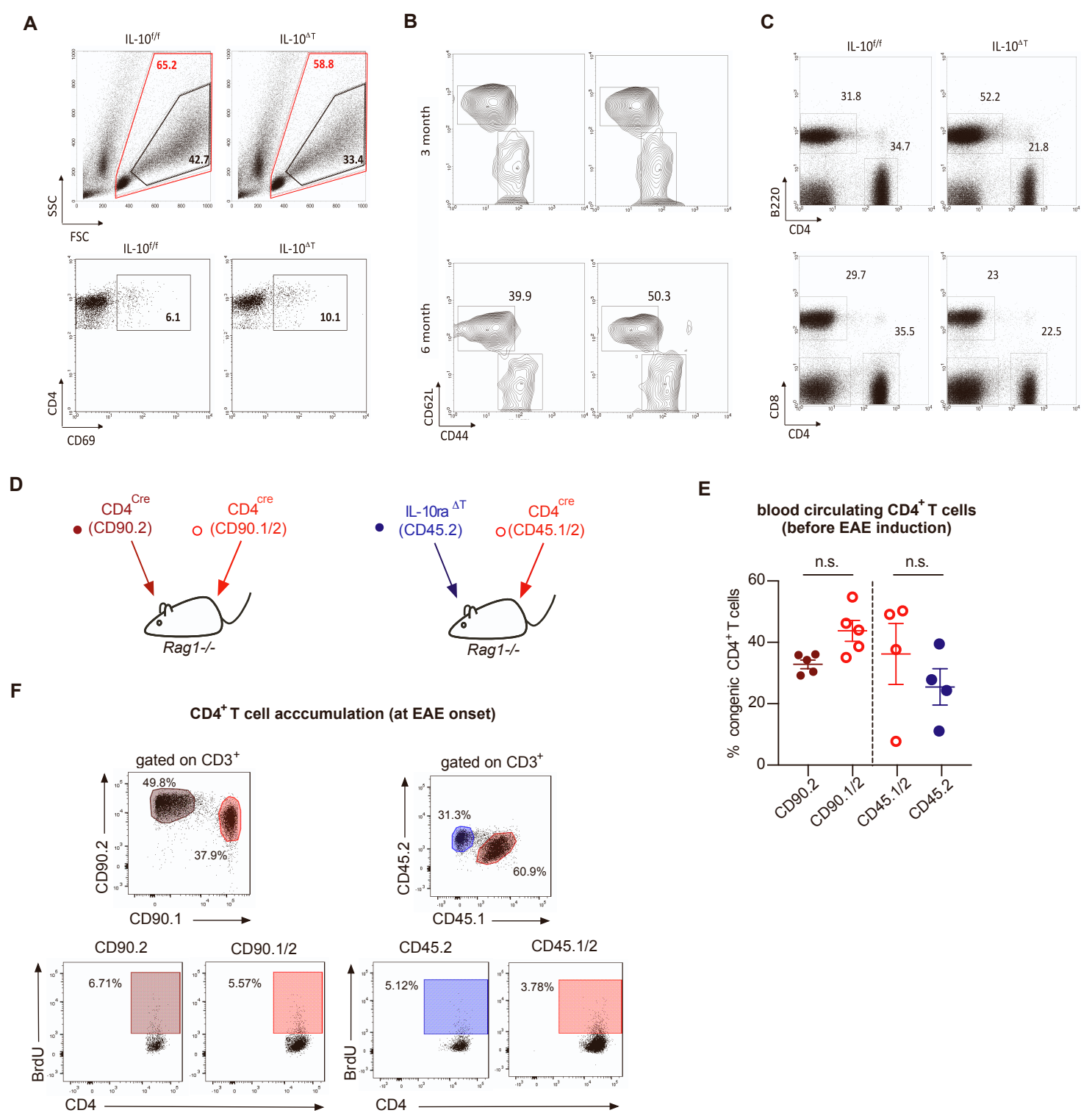


Figure S4

Figure S4 related to Figure 4: Loss of effector *Il10*-deficient T cells *in vivo* and reconstitution efficiency after co-transfer of *Il10ra*-proficient and -deficient T cells into *Rag1*^{-/-} mice.

(A) MACS-enriched CD4⁺ T cells isolated from ing.LN of EAE mice (day 10-post immunization) and co-cultured with MOG₃₅₋₅₅-pulsed BMDCs, IL-2, IL-12 and IL-23 for 5 days. (Upper panel) percentage of total viable cells (red) or cell-blasts (black). (Lower panel) percentage of activated (CD69⁺) CD4⁺ T cells. (B) Percentage of naïve CD4⁺ T cells found in LN of healthy, non-challenged, young and old mice (3 vs 6 month-old, respectively). (C) B cell (B220⁺) to CD4⁺ T cell ratio (upper panel), and CD8⁺ to CD4⁺ T cell ratio (lower panel), as seen in healthy, non-challenged, young mice. (D) Experimental layout of CD4⁺ T cell co-transfer into *Rag1*^{-/-} mice: Co-transfer of CD4⁺ T cells derived from littermate control mice (*CD4*^{Cre}*Il10ra*^{wt/wt}, short *CD4*^{Cre}), expressing either CD90.2 or CD90.1/2 congenic marker (dark red, full circles red vs red, empty circles, respectively). (right) Co-transfer of CD4⁺ T cells derived from either *CD4*^{Cre} control and IL-10 $\alpha^{\Delta T}$ mice or littermate *CD4*^{Cre} control mice, expressing either CD45.2 or CD45.1/2 congenic marker (blue, full circle vs red, empty circle, respectively). (E) To determine the reconstitution efficiency of congenic *CD4*^{Cre} wild-type and IL-10 $\alpha^{\Delta T}$ CD4⁺ T cells peripheral blood was sampled 4 to 5 weeks after reconstitution and stained for CD4, CD90.1 and CD90.2 or CD4, CD45.1 and CD45.2, respectively. The Diagram shows the ratio of congenic CD4⁺ T cells after transfer of a mixture of equal numbers of *CD4*^{Cre}:*CD4*^{Cre} control cells (left) and after transfer of a mixture of equal numbers of IL-10 $\alpha^{\Delta T}$:*CD4*^{Cre} control cells (right). (F) After 4 to 5 weeks of reconstitution, the mice were immunized with MOG₃₅₋₅₅ as described in the Material and Methods section, and gating strategy of FACS analyses of T cell subpopulations were performed at the indicated time points, as shown here for BrdU staining. Co-transfer of CD4⁺ T cells derived from littermate control mice (*CD4*^{Cre.pos}*Il10ra*^{wt/wt}, short *CD4*^{Cre}), expressing either CD90.2 (brown) or CD90.1/2 (red) congenic marker (also labeled as full vs empty circles, respectively). (right) Co-transfer of CD4⁺ T cells derived from IL-10 $\alpha^{\Delta T}$ and *CD4*^{Cre} control mice, expressing either CD45.2 (blue) or CD45.1/2 (red) congenic marker (also labeled as full vs empty circle, respectively).